



HI932

Automatic Potentiometric Titrator

Dear Customer,

Thank you for choosing a Hanna Instruments® product.

This manual has been written for HI932 Automatic Potentiometric Titrator with software version 2.2.2 and higher.

Please read this instruction manual carefully before using this instrument as it provides the necessary information for correct use of this instrument and a precise idea of its versatility.

If you need additional technical information, do not hesitate to e-mail us at tech@hannainst.com.

Visit www.hannainst.com for more information about Hanna Instruments and our products.

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INTRODUCTION

The **HI932** is an automatic potentiometric titrator with high accuracy, great flexibility and repeatability.

The titrator is designed to perform a variety of potentiometric titrations, allowing the user to obtain good results and high-speed analysis.

The main attributes of the **HI932** titrator are:

- Small footprint, requires minimal bench space
- Casing made with strong, chemically resistant plastic
- Flexible electrode holder supports up to 3 electrodes, 4 dispensing tubes, 1 temperature sensor and removable stirrer
- Electrode holder positions electrodes in the center of beaker, allowing for smaller sample sizes
- Integrated peristaltic pump available for reagent addition
- Support for 100 titration methods and 30 autosampler sequences
- Digital inputs for Hanna Instruments[®] digital probes
- User-customizable reports
- Integrated research grade pH/mV/ISE meter
- Clearly displayed warning and error messages

This manual provides information regarding installation and functionality of the titrator and refined operation suggestions. Before using the titrator, it is recommended you become familiar with its various features and functionality.

This manual is divided into four parts:

PART 1: QUICK START GUIDE

Helps the user quickly setup and operate **HI932** Automatic Potentiometric titrator.

It covers basic connections, user interface, and how to run a titration.

PART 2: INSTRUCTION MANUAL

Provides a comprehensive description of the operating principles, user interface, general options, methods, titration mode, optimization, maintenance, autosampler etc.

PART 3: APPLICATIONS

Contains complete instructions for commonly-used analyses.

Additional methods and method packs are available. Contact your local Hanna Instruments office for more details.

PART 4: TITRATION THEORY

Outlines the principles of operation of the titrator.

It covers the chemistry of titrations, titration types and result calculations.

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PART 1. QUICK START GUIDE

1.1. SAFETY MEASURES

The following safety measures must be followed:

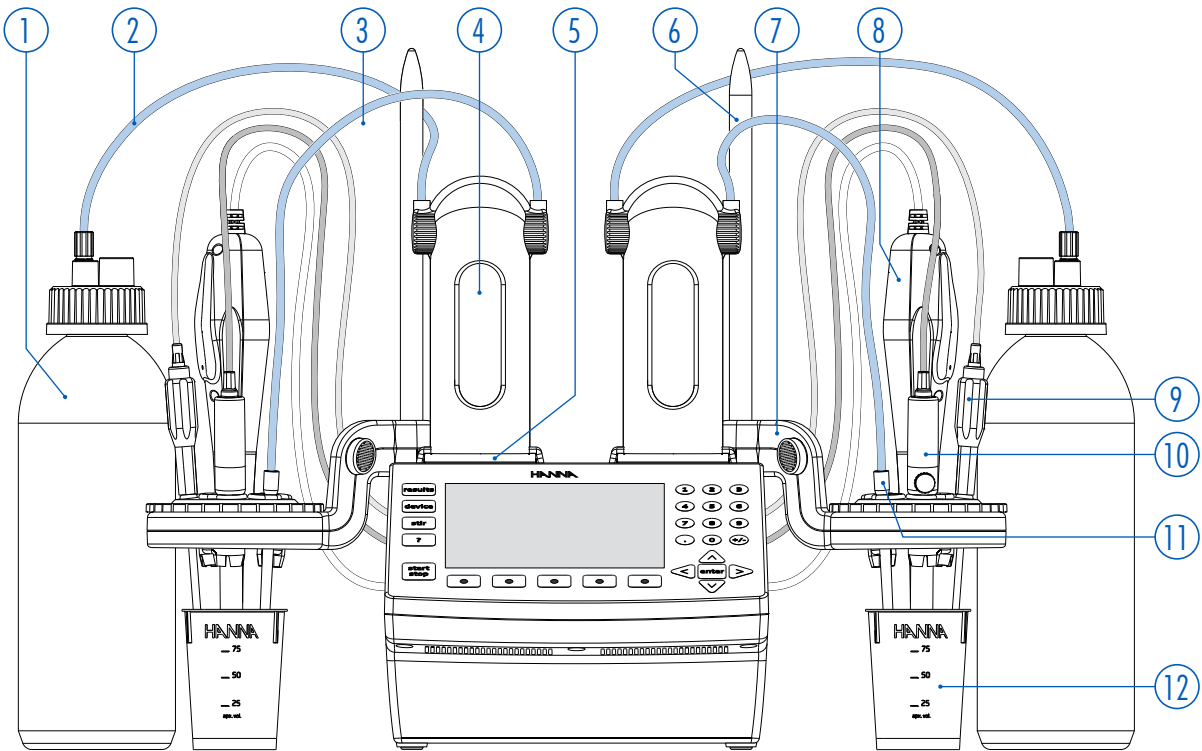
1. Never connect or disconnect the air pump and magnetic stirrer assembly or other peripheral with the titrator turned on.
2. Verify that the reagent and the attached tubing are assembled correctly.
3. Always check that the titrant, solvent and waste bottles, as well as the titration vessel are properly assembled.
4. Always wipe up spills and splashes immediately.
5. Avoid the following environmental working conditions:
 - Severe vibrations
 - Direct sunlight
 - Atmospheric relative humidity above 95% non-condensing
 - Environment temperatures below 10 °C (50 °F) and above 40 °C (104 °F)
 - Explosion hazards
 - Near heating or cooling sources
6. Have the titrator serviced by qualified service personnel only.
7. Avoid inhalation of reagent vapors.
8. Avoid contact with chemicals.

1.2. ABBREVIATIONS

ABS	Acrylonitrile Butadiene Styrene	mg / kg	Milligrams per kilogram
GLP	Good Laboratory Practice	mg / L	Milligrams per liter
PC-ABS	Polycarbonate-Acrylonitrile Butadiene Styrene	mmol / g	Millimoles per gram
PEI	Polyetherimide	mmol / kg	Millimoles per kilogram
PTFE	Polytetrafluoroethylene	mmol / L	Millimoles per liter
PVDF	Polyvinylidene fluoride	M (mol / L)	Molarity (moles per liter)
RPM	Revolutions per minute	mol / kg	Moles per kilogram
		mol / L	Moles per liter
eq / kg	Equivalents per kilogram	N (eq / L)	Normality (equivalents per liter)
eq / L	Equivalents per liter	ppb (μg / kg)	Parts per billion (micrograms per kilogram)
g / 100 mL	Grams per 100 milliliters	ppb (μg / L)	Parts per billion (micrograms per liter)
g / L	Grams per liter	ppm (mg / kg)	Parts per million (milligrams per kilogram)
μg / L	Micrograms per liter	ppm (mg / L)	Parts per million (milligrams per liter)
meq / kg	Milliequivalents per kilogram	ppt (g / kg)	Parts per thousand (grams per kilogram)
meq / L	Milliequivalents per liter	ppt (g / L)	Parts per thousand (grams per liter)
mg / 100 mL	Milligrams per 100 milliliters	% (g / 100 g)	Percent by weight (grams per 100 grams)
mg / g	Milligrams per gram	%w / v	Percent weight by volume

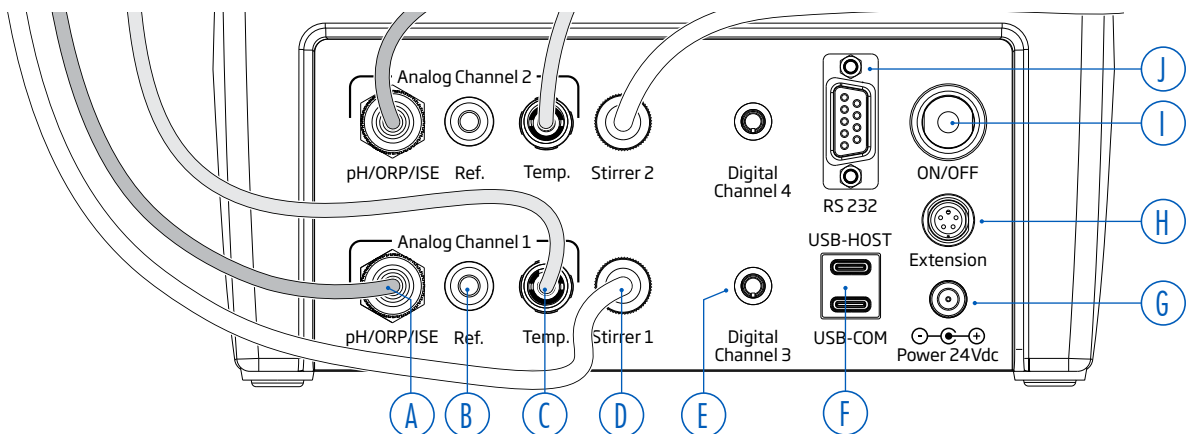
1.3. TITRATOR CONNECTIONS

Front View



- | | |
|-------------------------------------|-------------------------|
| 1. Titrant bottle | 7. Electrode holder |
| 2. Aspiration tube (titrant inlet) | 8. Stirrer |
| 3. Dispensing tube (titrant outlet) | 9. Temperature sensor |
| 4. Pump assembly | 10. pH or ORP electrode |
| 5. Burette assembly | 11. Dispensing tip |
| 6. Support rod | 12. Titration beaker |

Rear View

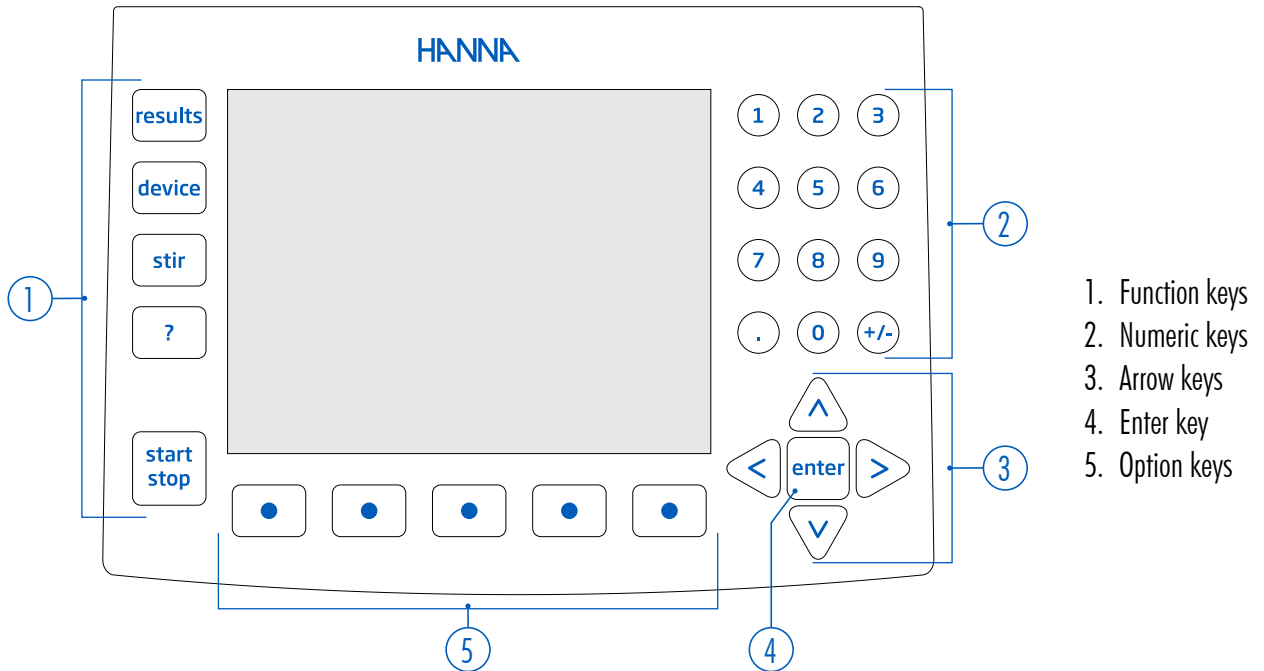


- | | |
|--|---|
| A. Probe connector (BNC) | F. USB-HOST: Keyboard, Photometric electrode power
USB-COM: Network box, PC connection |
| B. Reference electrode | G. Power input connector (24 VDC) |
| C. Temperature sensor | H. Extension: autosampler connection |
| D. Stirrer | I. Power switch |
| E. Digital probe connector (jack 3.5 mm) | J. Serial interface: balance |

1.4. USER INTERFACE

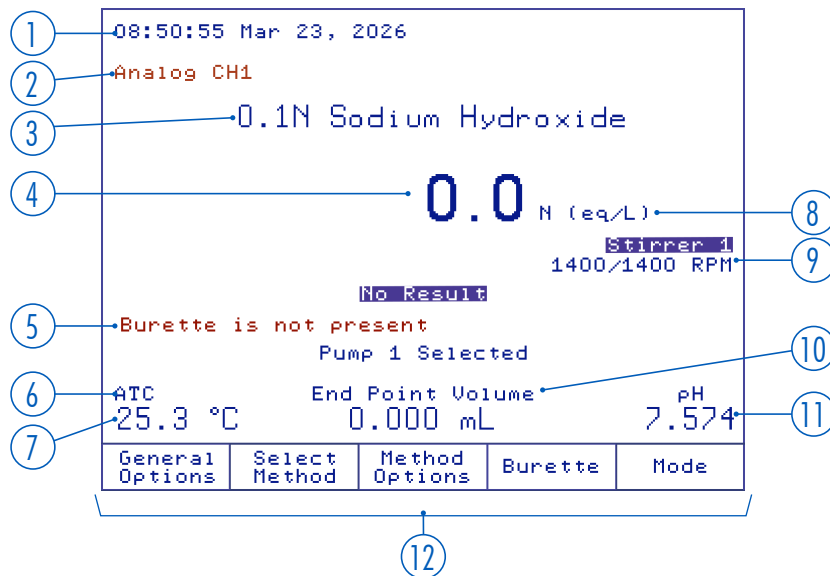
1.4.1. KEYPAD

The titrator's keypad has 27 keys grouped in five categories, as shown below.



1.4.2. DISPLAY

The titrator has a 5.7" graphical backlit color display.







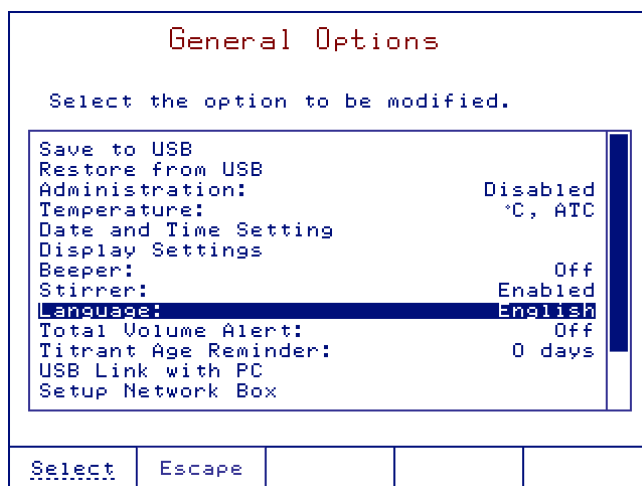
- | | |
|------------------------------------|--------------------------------|
| 1. Time and Date | 7. Temperature reading |
| 2. Active input channel | 8. Results units |
| 3. Method name | 9. Stirrer information |
| 4. Result | 10. End point titration volume |
| 5. Reminders or Warnings | 11. mV or pH reading |
| 6. Temperature compensation status | 12. Virtual option keys |

The user interface contains several screens. In each screen, many information fields are present at the same time. The information is displayed in an easy-to-read manner.

Virtual option keys describe the function performed when the corresponding soft key is pressed.

1.5. LANGUAGE

- To change the language, press  from the main screen.
- Highlight *Language* option.
- Use the  and  keys to select the language from the options listed in the **Set Language** screen.
- Press .
- Restart the titrator in order to apply the new language setting.



1.6. CONTEXTUAL HELP

Press  to access information about the titrator.

The contextual help can be accessed at any time and it provides useful information about the current screen.

1.7. METHODS

The **HI932** titrator can store up to 100 standard and user-defined methods.

Standard Methods

Each titrator is supplied with a package of standard methods.




Standard method packs are developed at Hanna Instruments® to meet analysis requirements of specific industries (e.g., water treatment, wine, dairy).

User-Defined Methods

User-defined methods allow the user to create and save their own methods.

Each new method is based on an existing method which is altered to suit a specific application.

1.8. HOW TO CALIBRATE A pH ELECTRODE

- To enter pH calibration mode, press .
- Next, press  followed by .

1.8.1. PREPARATION


- Pour small quantities of the buffer solutions into clean beakers. Use plastic beakers to minimize any EMC interferences. For accurate calibration and to minimize cross-contamination, use two beakers for each buffer solution: one for rinsing the electrode and one for calibration.
- If measuring in the acidic range, use pH 7.01 or 6.86 as the first buffer and pH 4.01/3.00 or 1.68 as the second buffer.
- If measuring in the alkaline range, use pH 7.01 or 6.86 as the first buffer and pH 10.01/9.18 or 12.45 as the second buffer.
- For extended range measurements (acidic and alkaline), perform a five-point calibration by selecting five buffers across the entire pH range.

1.8.2. CALIBRATION PROCEDURE

During calibration, the user has a choice of 8 standard buffers (pH 1.68, 3.00, 4.01, 6.86, 7.01, 9.18, 10.01, 12.45) and up to 5 custom buffers.





For accurate measurements it is recommended to perform a five-point calibration. However, at least a two-point calibration is suggested.

For pH titrations, the selected buffers should bracket the endpoint e.g. if endpoint value is at 8.5, use 7.01 or 6.86; and 9.18 or 10.01 for calibration.

1. Press  to begin calibration.

If the instrument was calibrated before, press  to clear previous calibration.

Note: It is very important to clear calibration history when a new electrode is used.

2. Immerse the pH electrode and the temperature probe approximately 4 cm (1.5") into a buffer solution and stir gently.
3. If necessary, select the pH calibration buffer value with  or .
4. Once the reading has stabilized, press  to update the calibration.
The calibration buffer will be added to the Calibrated Buffers section.
5. Rinse the pH electrode and the temperature probe.
6. Immerse pH electrode and the temperature probe into the next buffer solution and follow the above procedure.
Alternatively, press  to exit the calibration.




1.9. THE FIRST TITRATION

1.9.1. REQUIRED SOLUTIONS

- Titrant - 500 mL of 0.1 M (mol/L) Sodium Hydroxide (NaOH) in a titrant bottle
- Sample - 0.1 M (mol/L) Hydrochloric Acid (HCl)
- Distilled or deionized water

Note: Analytical grade reagents and water should be used for accurate results.


1.9.2. PRIMING THE BURETTE

1. Insert the aspiration tube in the titrant bottle and the dispensing tube in a waste beaker.
2. From the main screen, press .
3. Highlight the *Prime Burette* option, then press .
4. Enter the number of burette rinses.
At least 3 rinses are recommended.
5. Press  to start. The message "Executing..." will be displayed.

Note: Ensure continuous liquid flow inside the burette. For accurate results, the aspiration tube, the dispensing tube, and the syringe must be free of air bubbles.

1.9.3. METHOD SELECTION

For this analysis we will use the **HI1009EN Neutralization w/ NaOH** standard method.

1. To select this method, press  from the idle screen.
2. Use the  and  keys to highlight **HI1009EN Neutralization w/ NaOH** method.
3. Press .

1.9.4. SETTING METHOD PARAMETERS

1. Press Method Options to display the method parameters.
The **View/Modify Method** screen will be displayed.
Note: Only certain parameters can be changed.
2. For this titration, the NaOH titrant concentration and the size of the HCl sample need to be entered.
Highlight *Titrant Conc.* option, then press Select.
3. The Titrant Concentration screen will be displayed.
Enter the correct value, then press Accept.
4. Highlight *Analyte Size* option, then press Select.
5. Enter the volume of the sample (e.g.: 5 mL), then press Accept.
6. Press Escape then highlight *Save Method* option. Press Select.

Titrant Concentration

Enter the titrant concentration.

0.10678 M (mol/L)

Accept	Escape	Delete Digit		Exponent
--------	--------	--------------	--	----------

1.9.5. SETTING UP TITRATION REPORT

Users can select the information that is stored for each titration.

To obtain proper information at the end of the titration, perform the following operations:

1. From the main screen, press results and the **Data Parameters** screen will be displayed.
2. Highlight *Setup Titration Report* option and press Select.
3. Mark the fields to be included with the * symbol using the ^ and v keys.
4. Press Select to toggle the selection.
5. Press Save Report then press Escape to return to the main screen.

1.9.6. PREPARING THE SAMPLE

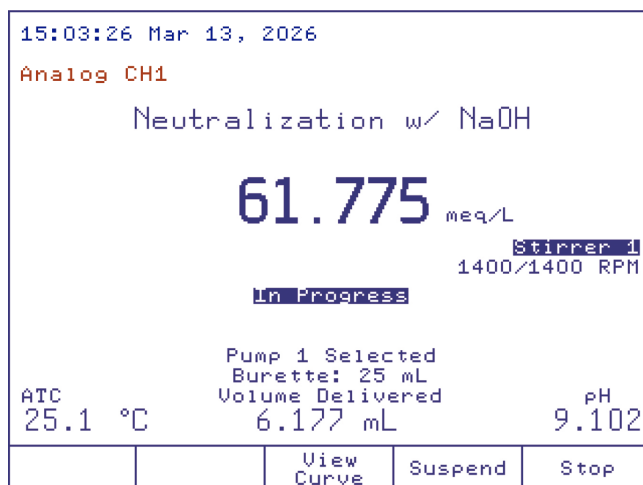
1. Add 50 to 65 mL of distilled / deionized water to the titration beaker.
2. Use a pipette (or burette) to add 5.0 mL of the sample (0.1M Hydrochloric Acid (HCl)) into the same beaker.
3. Slide the stirrer assembly up.
4. Place the beaker under the stirrer assembly.
5. Lower the stirrer assembly until the electrodes are submersed and the stirrer is close to the bottom of the beaker.
6. Adjust the level of the sample solution with distilled / deionized water so that the pH electrode bulb is completely immersed in the sample solution and the reference junction of the electrode is 5-6 mm below the surface.

1.9.7. PERFORMING A TITRATION

- From the main screen, press start stop. The user is prompted to enter the analyte size.
- Enter 5 mL and press enter. The titrator will start the analysis.
At the end of the titration, the message "Titration Completed" is displayed with the final concentration of the analyte in the sample and the equivalence endpoint volume.

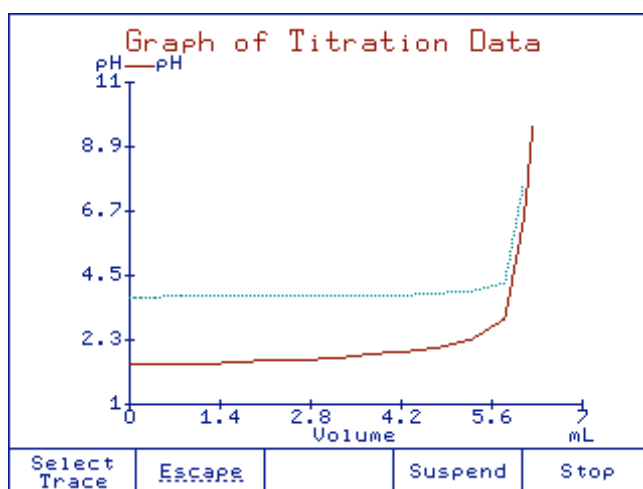
1.9.8. TITRATION SCREEN

During a titration, the following screen is displayed:



1.9.9. TITRATION GRAPH

- After a few doses are dispensed, View Curve will become active. Press View Curve to display the real-time titration graph. The curves displayed are plots of the pH and the 1st derivative versus titrant volume. The two graphs are scaled to fit in the same screen window. See [PART 2. Instruction Manual](#) for more information.
- Press Select Trace to change the y-axis scale to either the pH values or the 1st derivative values.



1.9.10. TITRATION TERMINATION

The titration is terminated when the conditions of the Termination Criteria have been met.

The titration is normally terminated when the first equivalence endpoint is detected according to the selected algorithm.

To ensure the correct detection and interpolation of the equivalence endpoint, the titrator will dispense a few additional doses after the endpoint was reached.

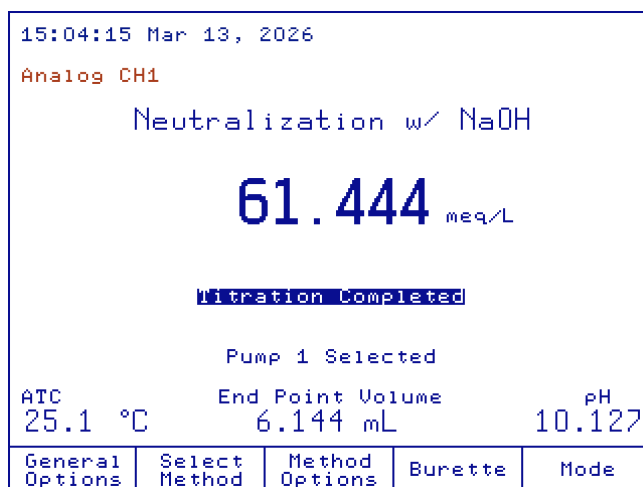
The titration result can be displayed either in the main screen or in the Graph of Titration Data screen.

When the titration has ended, the titrator will display the equivalence endpoint volume and the final concentration of the analyte together with the "Titration Completed" message.

To view the titration graph and/or results, press **results**.

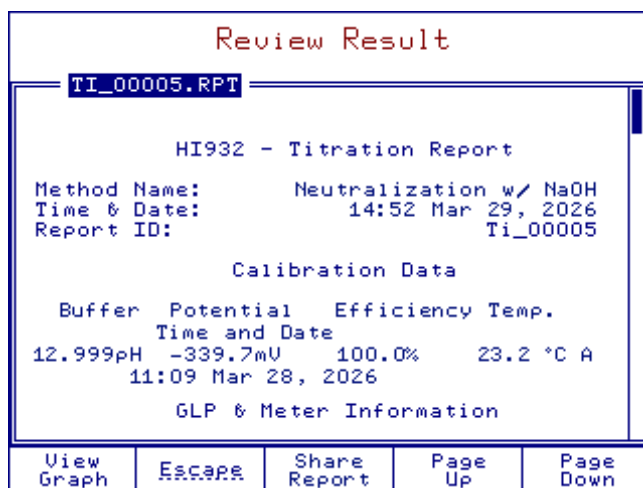
When the titration ends, an "x" will mark the endpoint on the pH versus titrant volume curve in the Graph of Titration Data screen.

The value of the endpoint volume is also displayed next to the endpoint.



1.9.11. RESULTS

The results obtained from titration are stored in a report file that can be displayed, transferred to a USB storage device or a PC, or shared via FTP/FTPS or SMB when connected to a network box.



1.9.12. VIEWING THE LAST TITRATION DATA

To view the last titration report:

1. From the main screen, press **results**.
The **Data Parameters** screen will be displayed.
2. From the **Data Parameters** screen, highlight *Review Last Analysis Report* option.
3. Press **Select**. The **Review Result** screen will be displayed.
4. Use the **Page Up** and **Page Down** keys to display information related to the last titration performed.

1.9.13. SHARING THE TITRATION REPORT

Note: The HI932933 Network Box must be connected, to print or transfer files via FTP/FTPS and SMB.

1. When in the **Review Report** screen, press **Share Report**.
2. Use the **▲** and **▼** keys to choose sharing type (Print, Add to summary, FTP/FTPS or SMB transfer, PDF, or Share as raw file to USB flash drive).
3. Report graphs can also be shared by pressing **View Graph**, then **Save as Bitmap**.
Press **Share** to select sharing type for the graph (Print, FTP/FTPS or SMB transfer, PDF, or Share as raw file to USB flash drive).
4. Press **Escape** to return to the Data Parameters screen.
5. Press **Escape** again to return to the main screen.

Share stored reports and graphs

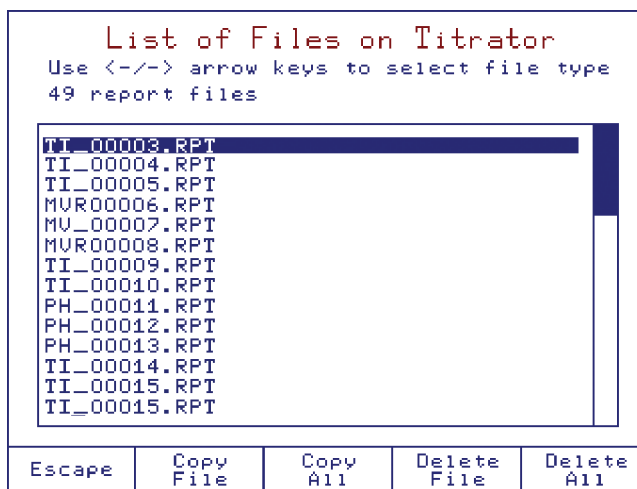
1. Navigate to *Review Available Reports* on the **Data Parameters** screen.
2. Use the **▲** and **▼** keys to select reports to view and/or share.

1.9.14. SAVING DATA TO USB STORAGE DEVICE

Note: The USB Storage Device has to be formatted FAT or FAT32.



This feature allows saving the results of titrations or logging sessions on a USB storage device.

1. From the main screen, press **General Options**. The **General Options** screen will be displayed.
2. Use the **▲** and **▼** keys to highlight *Save to USB Storage Device* option.
3. Insert the USB storage device into the USB socket.
4. Press **Select**. The **List of Files on Titrator** screen will be displayed.
5. Use the **▲** and **▼** keys to select the report files.



6. Press **Copy All** to transfer all available reports to the USB storage device, or highlight the name of the report file to be transferred.
7. Press **Copy File**.
Transferring a report file will automatically transfer the corresponding log file and titration graph (*.BMP file if applicable).
8. Press **Escape** to return to the **General Options** screen.
9. Press **Escape** again to return to the main screen.

1.9.15. TITRATION REPORT

While scrolling with the  and  keys, the fields below can be seen on the titrator display or shared. The same information is available on the saved report file (Ti_00011.rpt in this example, with all report fields selected).

HI932 - Titration Report

Method Name: Neutralization w/ NaOH
 Time & Date: 15:01 Jun 13, 2026
 Report ID: Ti_00011

Calibration Data

Buffer	Potential	Efficiency	Temp.
Time and Date			
4.010pH	169.3mV	98.8%	24.0°C A
			11:44 Jun 13, 2025
7.010pH	-5.8mV	98.7%	23.9°C A
			11:42 Jun 13, 2025
10.010pH	-180.7mV	98.7%	24.0°C A
			11:46 Jun 13, 2025

GLP & Meter Information

Sample Name:
 Company Name:
 Operator Name:
 Electrode Name:
 Field 1:
 Field 2:
 Field 3:
 Titrator Software Version: v2.2.2
 Base Board Software Version: v1.00
 Pump 1 Software Version: v1.00
 Pump 2 Software Version: v1.00
 Stirrer 1 Software Version: v1.00
 Titrator Serial Number: TT180525011
 Analog CH1 Serial Number: AB180525005
 Analog CH2 Serial Number: AB180525006
 Digital CH3 Serial Number: D180525007
 Digital CH4 Serial Number: D180525008
 Pump 1 Serial Number: DP180525004
 Pump 2 Serial Number: DP180525007
 Stirrer 1 Serial Number: OS180524001
 Analog CH1 Calibration Date: May 25, 2026
 Analog CH2 Calibration Date: May 25, 2026
 Digital CH3 Calibration Date: May 25, 2026
 Digital CH4 Calibration Date: May 25, 2026

Method Parameters

Name: Neutralization w/ NaOH
 Method Revision: 4.0
 Analysis Type: Standard Titration
 Measurement Input: Analog CH2
 Stirrer Configuration:
 Stirrer: Stirrer 1
 Stirring Speed: 1400 RPM

```

Pump Configuration:
  Titrant pump:           Pump 1
  Reagent Addition 1:    Disabled
  Reagent Addition 2:    Disabled
  Dosing Type:           Dynamic
  Min Vol:               0.050 mL
  Max Vol:               0.500 mL
  delta E:               20.000 mV
  End Point Mode:       pH 1EQ point,1st Der
  Recognition Options
  Threshold:             50 mV/mL
  Range:                 NO
  Filtered Derivatives: NO
  Pre-Titration Volume:  0.000 mL
  Pre-Titration Stir Time: 0 sec
  Measurement Mode:     Signal Stability
  delta E:               1.0 mV
  delta t:               2 sec
  Min wait:              2 sec
  Max wait:              15 sec
  Electrode Type:       pH
  Blank Option:         No Blank
  Calculations:         Sample Calc. by Volume
  Dilution Option:     Disabled
  Titrant Name:         0.1N NaOH
  Titrant Conc.:        0.1000 N (eq/L)
  Analyte Size:         10.0000 mL
  Analyte Entry:       Fixed
  Maximum Titrant Volume: 20.000 mL
  Potential Range:      -2000.0 to 2000.0 mV
  Volume/Flow Rate:     25 mL / 50.0 mL/min
  Signal Averaging:     1 Reading
  Significant Figures:   XXXXX

```

N (eq/L) --> meq/L

```

V eq 1000meq
--*--*-----
  L   eq
-----
mL   L
--*-----
  1000mL

```

V = volume dispensed in liters.
 0.100 eq/L -> titrant conc.
 10.000 mL -> sample volume

Nr	Volume[mL]	mV	pH	Graphic	Temp. [°C]	Time
0	0.000	274.4	2.219	0.0	24.9	A 00:00:00
1	0.050	274.4	2.220	1.0	25.0	A 00:00:07
2	0.100	274.4	2.220	0.0	25.0	A 00:00:10
3	0.200	274.3	2.222	-0.8	25.0	A 00:00:12
4	0.400	274.0	2.227	-1.6	25.0	A 00:00:15
5	0.800	273.2	2.241	-2.0	25.0	A 00:00:18
6	1.300	271.5	2.271	-3.4	25.0	A 00:00:24
7	1.800	269.5	2.304	-3.9	25.1	A 00:00:30
8	2.300	267.2	2.344	-4.7	25.1	A 00:00:37

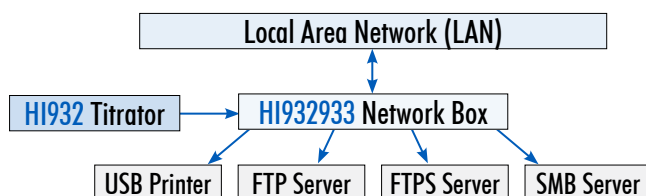
9	2.800	264.4	2.393	-5.7	25.1	A	00:00:43
10	3.300	260.8	2.455	-7.2	25.1	A	00:00:50
11	3.800	256.1	2.535	-9.3	25.1	A	00:00:58
12	4.300	250.3	2.635	-11.7	25.1	A	00:01:05
13	4.800	241.9	2.779	-16.8	25.1	A	00:01:14
14	5.300	228.3	3.011	-27.2	25.1	A	00:01:23
15	5.800	193.0	3.614	-70.5	25.1	A	00:01:31
16	6.077	21.0	6.556	-620.0	25.1	A	00:01:48
17	6.128	-38.2	7.568	-1183.2	25.1	A	00:02:03
18	6.177	-123.6	9.031	-1708.0	25.1	A	00:02:19
19	6.227	-157.7	9.616	-682.8	25.1	A	00:02:28
20	6.278	-174.5	9.903	-335.8	25.1	A	00:02:35
21	6.339	-187.8	10.130	-215.9	25.1	A	00:02:42

Titration Results

Method Name: Neutralization w/ NaOH
 Time & Date: 15:01 Jun 13, 2026
 Analyte Size: 10.0000 mL
 End Point Volume: 6.144 mL
 pH Equivalence Point: 8.063
 Result: 61.444 meq/L
 Initial & Final pH: 2.219 to 10.130
 Titration Duration: 2:42 [mm:ss]
 Titration went to Completion

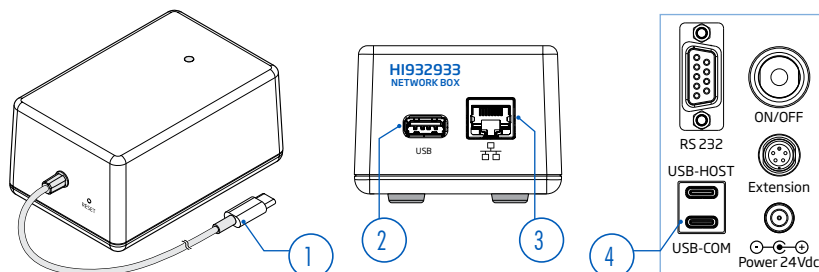
1.10. NETWORK BOX

The **HI932933** is a secure network device designed for **HI932** Automatic Potentiometric Titrator running software version 2.2.2 and higher. It enables secure Ethernet connection for data exchange and adds USB printer support, allowing direct printing of reports, methods, and data logs.



To connect the **HI932** titrator to its peripherals correctly:

1. Plug the USB-C end (1) into the **HI932** titrator USB-COM port (4).
2. Plug the printer into the USB port on the box (2).
3. Plug the Ethernet cable into the Ethernet connection port (3).



The device supports software updates for security patches and new features (see [2.3.13. Network Box Setup » Update Network Box Software](#)); and uses SMB and FTP/FTPS protocols for secure data transfer to PC and network servers.

The **HI932** Automatic Titrator allows network box configuration directly through its interface, without additional software. See [2.3.13. Network Box Setup](#).

PART 2. INSTRUCTION MANUAL

2.1. SETUP

2.1.1. UNPACKING

Remove the titrator and accessories from the packaging and examine it carefully.

For further assistance, please contact your local Hanna Instruments® office or email us at tech@hannainst.com.

Each [HI932](#) potentiometric titrator is supplied with:

- Titrator
- Pump assembly
- Burette assembly
 - » Burette with 25 mL syringe
 - » Aspiration tube with fitting and protection tube
 - » Dispensing tube with dispensing tip, protection tube, and tube guide
 - » Tube locks
 - » Tool for burette cap removal
 - » Light spectrum protection screen
- Electrodes holder and stirrer
 - » Electrode holder
 - » Overhead stirrer
 - » Propellers (3 pieces)
 - » Support rod
- Burette blank support
- Pump and burette locking screws with plastic head
- Temperature sensor
- Shorting cap
- Power adapter
- USB cable
- USB memory stick
- [HI900](#) PC application (installation kit on USB-C memory stick)
- Quality certificate
- Instruction manual

If any of the items are missing or damaged, please contact your local Hanna Instruments office or email us at tech@hannainst.com.

See [2.12.3. Titrator Components](#) section for component pictures.

Note: Save all packing materials until you are sure that the instrument functions correctly. Any damaged or defective items must be returned in their original packing materials together with the supplied accessories.

2.1.2. SAFETY MEASURES

The following safety measures must be followed:

- Never connect or disconnect the pump assembly or the air pump and magnetic stirrer assembly with the titrator turned on.
- Verify that the burette and the attached tubing are assembled correctly.
See [Burette Assembly](#) section for more information.
- Always check that the titrant bottle and the titration beaker are on a flat surface.
- Always wipe up spills and splashes immediately.
- Avoid the following environmental working conditions: severe vibrations, direct sunlight, atmospheric relative humidity above 95% non-condensing, environment temperatures below 10 °C (50 °F) and above 40 °C (104 °F), explosion hazards.
- Have the titrator serviced only by qualified service personnel.

2.1.3. HI932 TITRATOR TECHNICAL SPECIFICATIONS

Analysis Type	Standard titration (Standardization, Fixed pH / mV, Equivalence point pH / mV)		
	Back titration		
	Direct reading		
Endpoint Mode	Fixed mV		
	Fixed pH		
	mV equivalence point (up to 5 points, 1 st or 2 nd derivative)		
	pH equivalence point (up to 5 points, 1 st or 2 nd derivative)		
Burette	Size	5 mL, 10 mL, 25 mL, 50 mL	
	Resolution	0.001 mL	
	Flow rate	0.1/0.3 mL to 2 × burette volume per minute	
	Accuracy	±0.005 mL (5 mL burette)	
		±0.010 mL (10 mL burette)	
		±0.025 mL (25 mL burette)	
±0.050 mL (50 mL burette)			
Stirrer	Type	Overhead	
	Range	200 to 2500 RPM	
	Resolution	100 RPM	
mV	Range	−2000.0 to 2000.0 mV	
	Resolution	0.1 mV	
	Accuracy	±0.1 mV	
	Calibration	single point, offset	
pH	Range	−2.000 to 20.000 pH	
	Resolution	0.1 / 0.01 / 0.001 pH	
	Accuracy	±0.002 pH (±1 last significant digit)	
	Calibration	up to 5 points with standard or custom buffers	
ISE	Range	1×10^{-6} to 9.999×10^{10}	
	Resolution	1 / 0.1 / 0.01	
	Accuracy	±0.002 pH (±1 last significant digit)	
	Calibration	up to 5 points	

Temperature	Range	–5.0 to 105 °C 23.0 to 221.0 °F 268.2 to 378.2 K
	Resolution	0.1
	Accuracy	±0.1 °C / ±0.2 °F / ±0.1 K
Data Storage	Methods	Up to 100 titration methods (standard and user-defined) Up to 30 autosampler sequences
	Reports	Up to 100 titration and pH / mV / ISE reports Up to 40 autosampler tray reports (e.g. 720 reports for 18 beaker tray)
Connections	Measurement	2 × BNC socket (pH, ORP, ISE half-cell, and ISE combination electrodes) 2 × 4 mm banana socket (reference electrode) 2 × RCA socket (temperature sensor) 2 × 6-pin connector (stirrer) 2 × 3.5 mm Jack connector (digital sensor)
	Peripheral	1 × DB-9 socket (serial interface) 2 × USB type C (external PC keyboard, network box, PC connection) 1 × 5-pin connector (balance) 1 × USB standard A (USB flash drive)
Additional Specifications	Electrode holder	4 × multi-purpose slots (titrant / reagent tubes) 3 × 12-mm electrode slots 1 × temperature sensor slot 1 × overhead stirrer slot
	Display	5.7" graphical color display with backlight
	Power supply	100 - 240 VAC, 50 / 60 Hz
	Power draw	0.5 amps
	Enclosure material	ABS, PC-ABS, stainless steel
	Keypad	Polyester
	Dimensions	315 × 205 × 375 mm (12.4 × 8.1 × 14.8 ")
	Weight	Approximately 4.3 kg (9.5 lbs.) with 1 pump, stirrer, and sensors
	Operating environment	10 to 40 °C (50 to 104 °F); up to 95 % RH
Storage environment	–20 to 70 °C (–4 to 158 °F); up to 95 % RH	

2.1.4. HI922 AUTOSAMPLER TECHNICAL SPECIFICATIONS

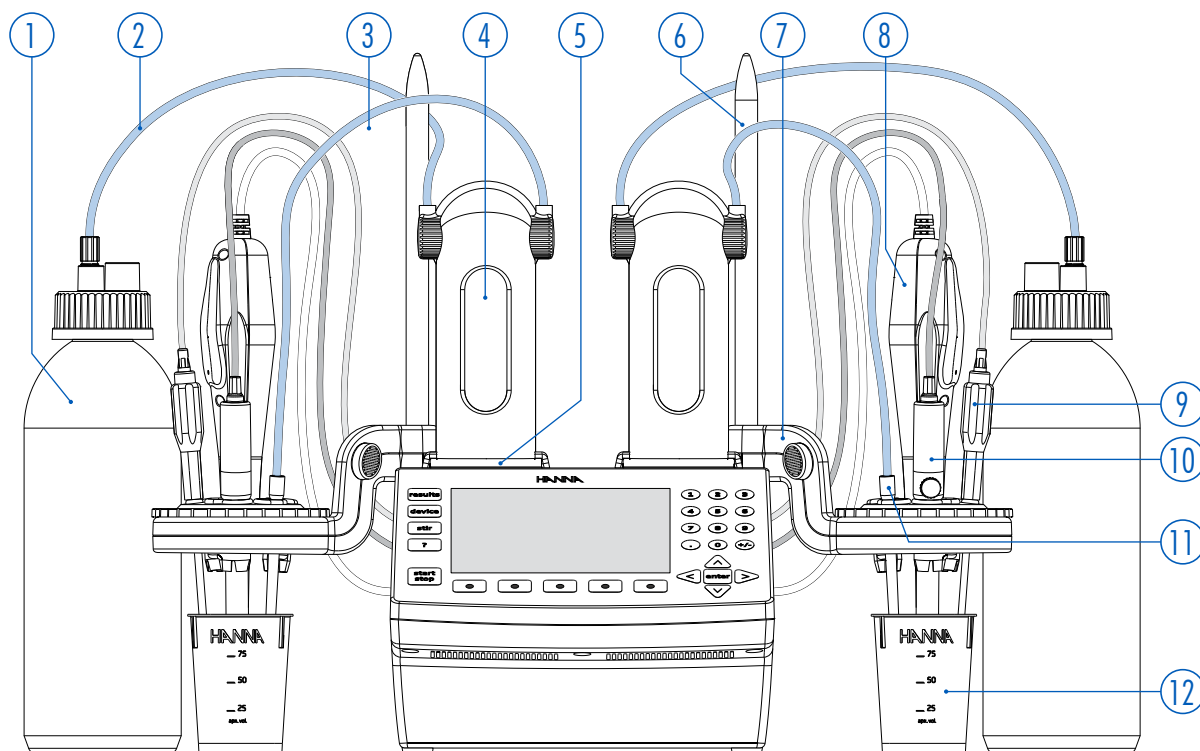
Electrode holder	5 × multi-purpose slots (titrant / reagent tubes) 3 × 12 mm electrodes slots 1 × overhead stirrer slot 1 × temperature sensor slot 1 × aspiration tube slot
Stirrer	Magnetic stirrer (built-in) Overhead stirrer (optional)
Temperature sensor	HI7662-AW (included)
Peristaltic pumps	Up to three (slot 1, 2, or 3)
Diaphragm pumps	One (slot 4)
Peripheral units	USB barcode reader
Trays	16 beakers × 150 mL with built-in RFID tag 18 beakers × 100 mL with built-in RFID tag
Beakers	ASTM short-form glass beakers, 100 & 150 mL HI920-060 150 mL plastic beakers HI920-053 100 mL plastic beakers
Control panel	Buttons for manual operation of tray Manual operation of peristaltic or diaphragm pumps Two-line backlit display with status information
Enclosure material	ABS plastic and steel
Electrode holder material	ABS plastic
Tray material	ABS plastic and acrylic
Keypad material	ABS plastic and polycarbonate
Weight	Approximately 13 kg (29 lbs)
Operating environment	10 to 40 °C (50 to 104 °F) ; up to 95% RH
Storage environment	−20 to 70 °C (−4 to 158 °F); up to 95% RH

2.1.5. HI932933 NETWORK BOX SPECIFICATIONS

Connection type	Attached USB-C cable
Power source	HI932 Automatic Potentiometric Titrator (USB-COM port)
Ethernet input	RJ-45 Ethernet connector (10/100 Mbps connection)
Status indicator	LED color-coded system
Dimensions	70 × 105 × 50 mm (2.7 × 4.1 × 1.9")
Weight	150 g (0.33 lb)

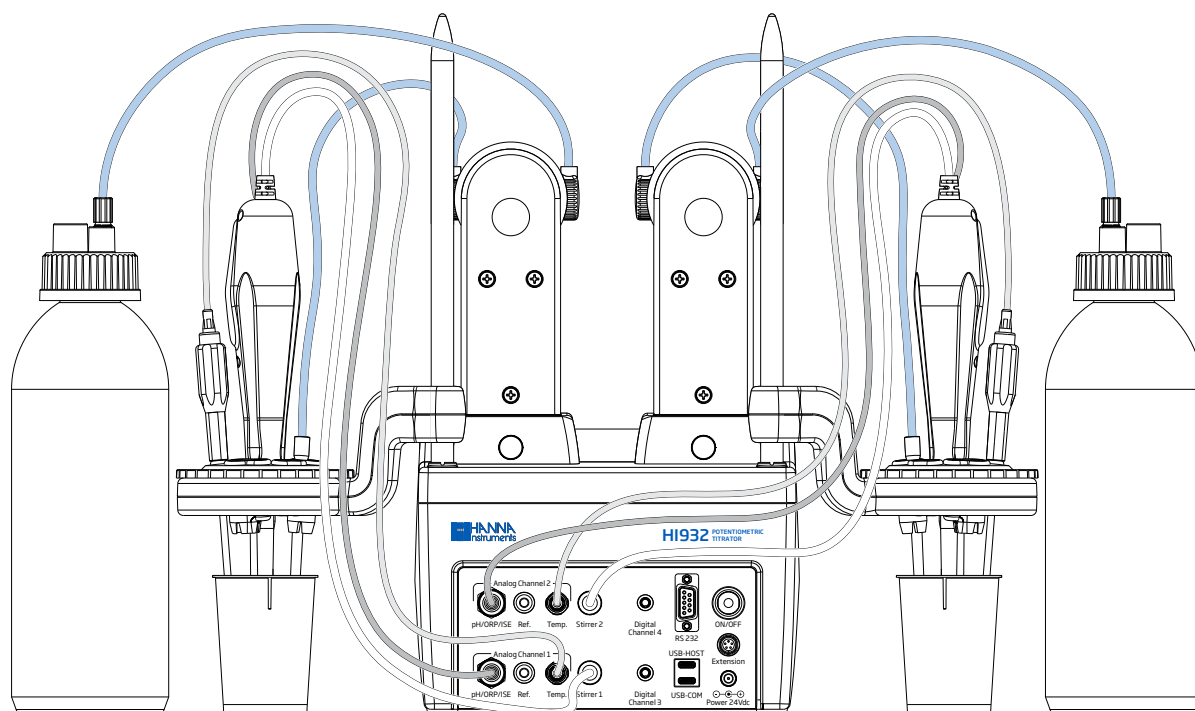
2.1.6. INSTALLATION

2.1.6.1. Titrator Front View



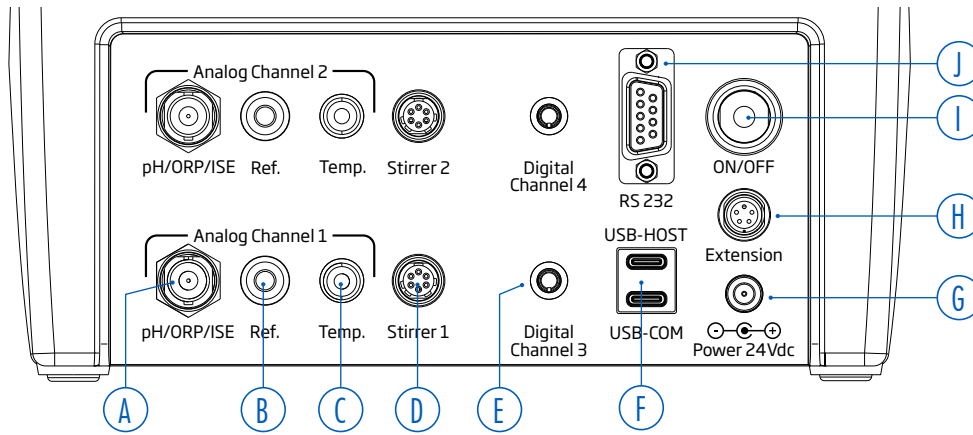
- | | |
|-------------------------------------|-----------------------|
| 1. Titrant bottle | 7. Electrode holder |
| 2. Aspiration tube (titrant inlet) | 8. Stirrer |
| 3. Dispensing tube (titrant outlet) | 9. Temperature sensor |
| 4. Pump assembly | 10. pH electrode |
| 5. Burette assembly | 11. Dispensing tip |
| 6. Support rod | 12. Titration beaker |

2.1.6.2. Titrator Rear View



2.1.6.3. Titrator Rear Connections

1. Connect the electrode to the BNC connector (A) and the temperature sensor to the RCA connector (C), or connect the digital sensor to the 3.5 mm jack (E).
2. Connect the stirrer to the mini-DIN connector (D).
3. Connect the power adapter cable to the power input connector (G).

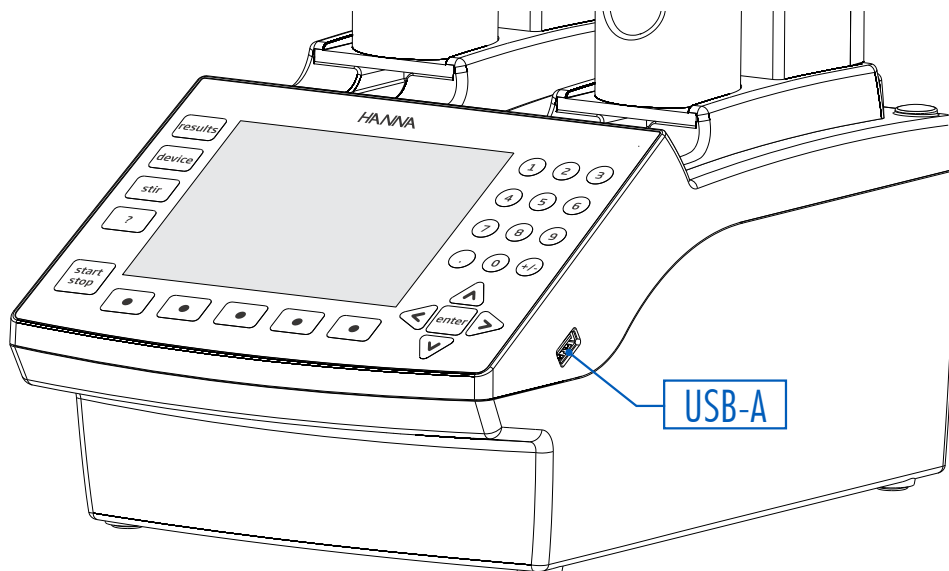


Function

Connector Type

A. Connection for pH, ORP, ISE half-cell, ISE combination electrodes	BNC socket
B. Reference electrode	Ø 4 mm banana socket
C. Temperature sensor	RCA socket
D. Stirrer	6-pin connector
E. Digital channel input	3.5 mm jack
F. Keyboard (HOST), Network box (USB-COM), PC (COM) Photometric electrodes (HOST or COM)	2×USB type C
G. Power input connector (24 VDC)	DC power jack connector
H. Autosampler interface (extension)	5-pin connector
I. Power switch	-
J. Balance interface	DB-9 socket (RS-232)

2.1.6.4. Titrator Side View



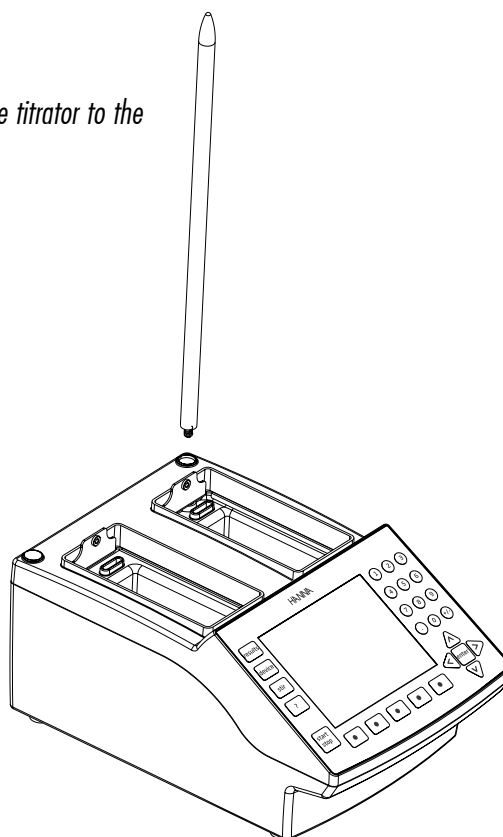
2.1.6.5. Titrator Assembly

Note: Assembly operations must be completed before connecting the titrator to the power supply!

Assembling Support Rod

To insert support rod into the titrator case:

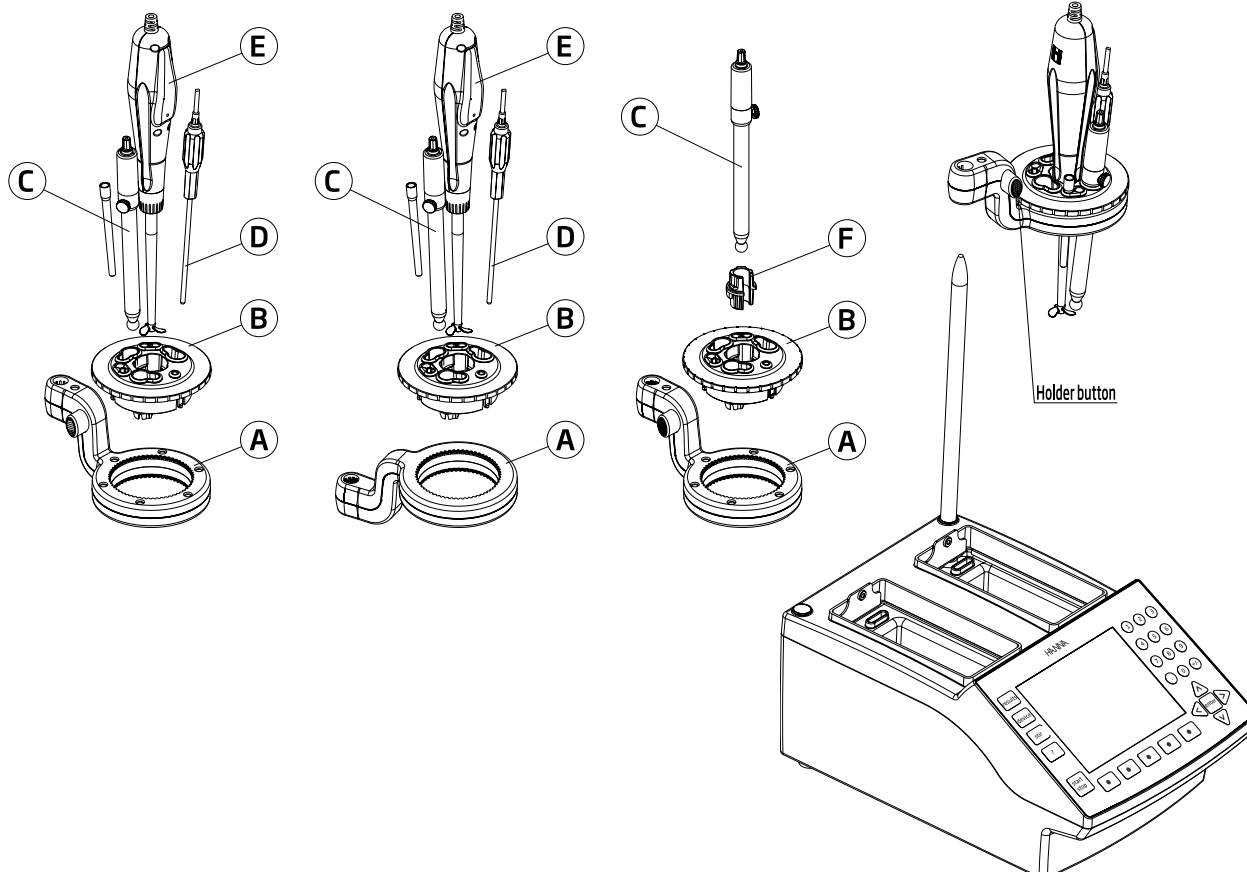
1. Remove protective cap from titrator case
2. Insert the support rods into the titrator case
3. Turn the support rod clockwise to secure it in place



Attaching Stirrer & Electrode

1. Place the electrode holder (B) in the stirrer support housing (A).
The stirrer support housing can be inverted if necessary.
2. Slide the electrode holder into the support rod and set the desired height using the **holder button**.
3. Insert electrode (C), temperature sensor (D) and stirrer (E) into the dedicated holes in the electrode holder.
Push them until they are in a stable position.

Note: For small sample sizes, use the electrode adapter (F) in the center of the holder.



Attaching the Photometric Electrodes

The HI90060X series of photometric electrodes can be connected to the HI932 Titrator by plugging the BNC connector into the pH/ORP/ISE port (A) and the PS/2 connector into the USB-HOST or USB-COM port (B).

1. Turn off titrator when connecting a photometric electrode to ensure proper initialization.

2. **BNC connector**

The BNC electrode connector is plugged into the designated pH/ORP/ISE port (A).

3. **PS/2 connector**

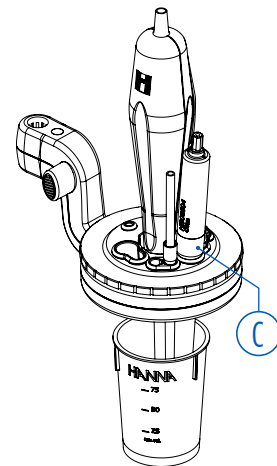
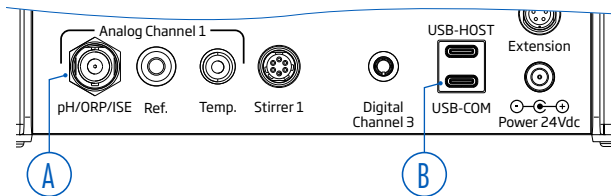
The PS/2 connector is plugged into the female port of the PS/2 to USB converter (HI932934).

Align the flat plastic tab on the PS/2 plug with the notched slot inside the converter's female port.

Gently insert the connector. Do not force it.

4. Plug the USB end of the converter into the USB-HOST or USB-COM port (B) on the HI932 Titrator.
5. To ensure the HI932 Titrator detects the electrode, plug into the converter then restart the Titrator.

See [2.12. Accessories](#) » [2.12.2. Sensors](#) and [2.12. Accessories](#) » [2.12.3. Titrator Components](#) for ordering information.

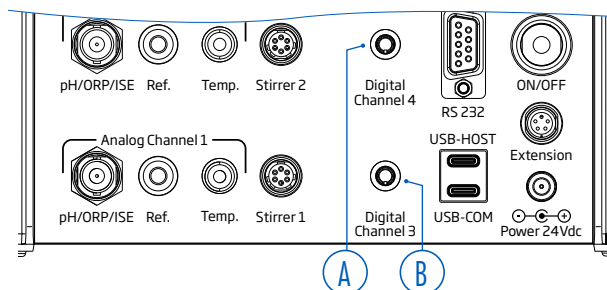


The photometric electrode (C) can be installed in two locations on the HI932 electrode holder. Both of these place the probe as far downstream from the clockwise-spinning titrant as possible.

Attaching the Digital Probes

HI932 utilizes two rear-panel digital inputs to connect Hanna Instruments® digital probes. When connected, the titrator automatically recognizes the sensor and its serial number.

1. Turn off titrator before connecting a digital probe to ensure proper initialization.
2. Insert the 3 mm probe jack into one of the rear panel digital inputs (A or B).
Ensure the probe is securely seated.



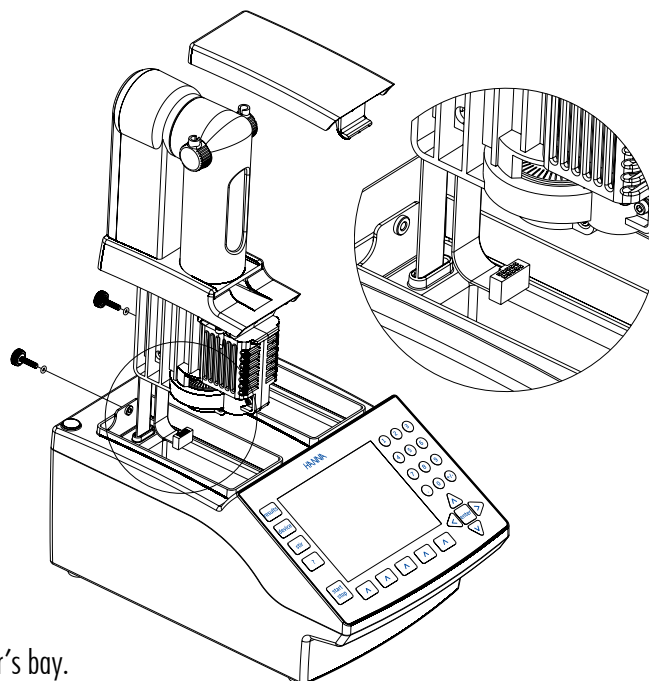
3. Select the active input channel (see [2.6. pH Mode](#) section).

For available digital probes please refer to [2.12. Accessories](#) » [2.12.2. Sensors](#) section.

Connecting the Pump

1. Retrieve the pump cable from inside the bay.
The pump 1 connector is located in the left bay and the pump 2 connector in the right bay.
2. Connect the cable to the pump as shown here.
The pump connector is located on the bottom of the pump.
3. Lower the pump into the titrator, then slide it towards the front of the titrator case until it is firmly latched.
4. Secure the pump with the locking screw.

This procedure can be repeated to connect a second pump.

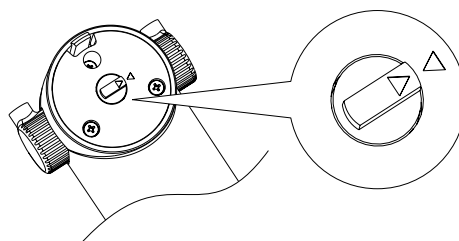


Attaching Burette Blank Support

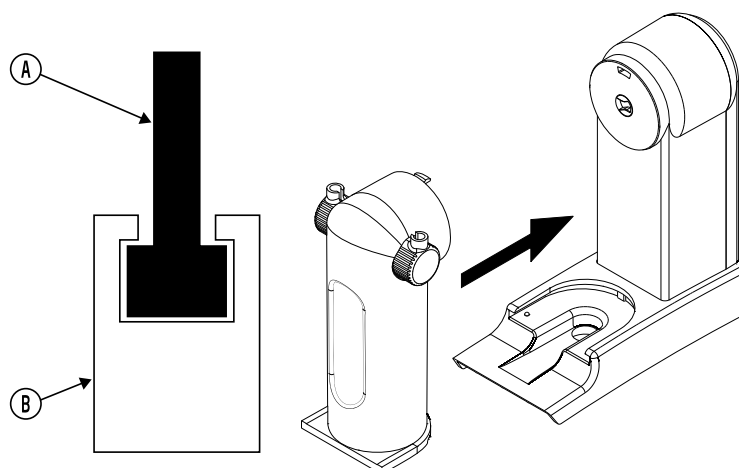
1. Insert and lower the burette blank support into the titrator's bay.
2. Slide it towards the front of the titrator case until it is firmly latched.
3. Secure the burette blank support with the locking screw.

Attaching the Burette

1. Make sure that the mark from the valve actuating cap and from the burette body are aligned.



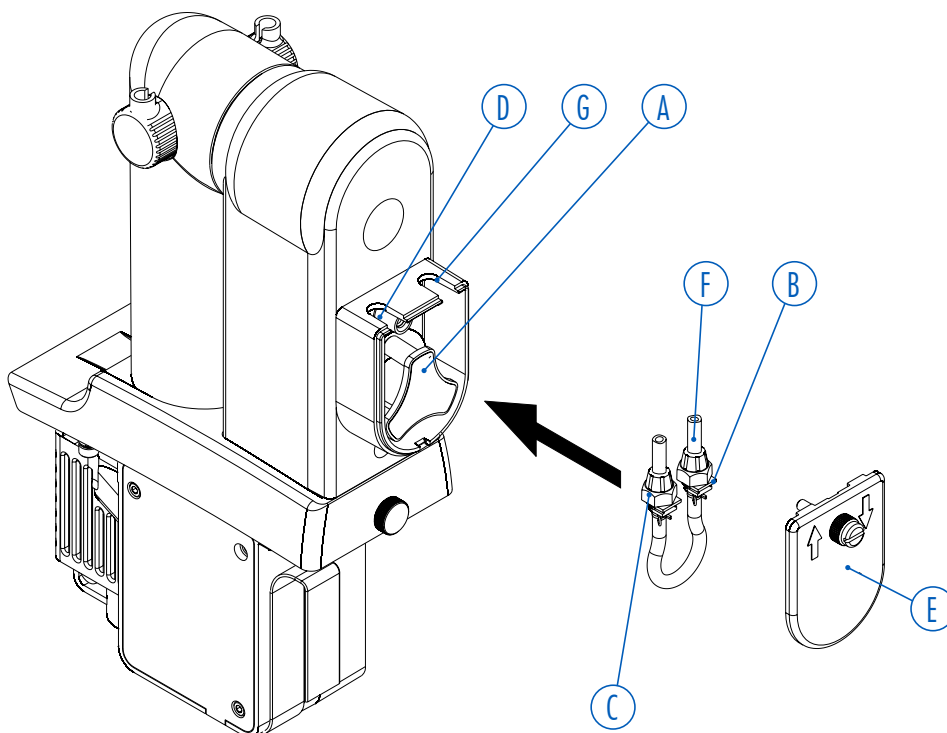
2. Slide the burette into the support on the burette pump. Ensure correct coupling between the syringe plunger (A) and the pump piston (B).



Connecting Peristaltic Pump Tubing

To attach the pump tubing to the burette pump with the built-in peristaltic pump:

1. Use a screw driver to remove the plastic cover (E) from the pump.
2. Remove the blue tube connectors (F).
3. Insert the roller tube (C) into the left side of the holder (D).
The fitting on the top of the roller tube will sit on the top of the housing.
4. Manually rotate the pump (A) counterclockwise until the tubing is mounted on the pump.
5. Insert the roller tube (B) into the right side of the holder (G).
The fitting on the top of the roller tube will sit on the top of the housing.
6. Attach aspiration and dispensing tubing to the roller tubing and replace the blue tube connectors (F).
7. Replace the plastic cover (E).

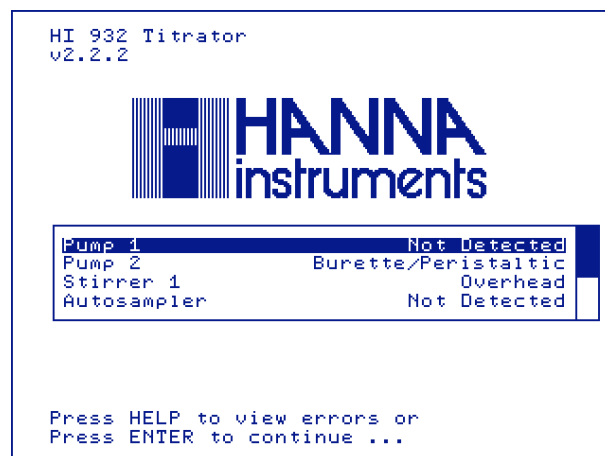
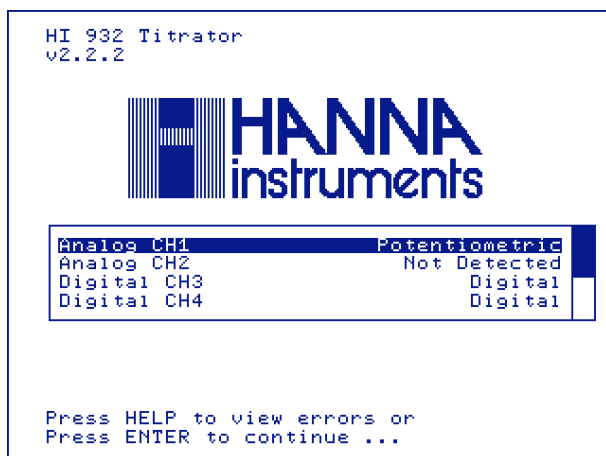


2.2. USER INTERFACE

2.2.1. START UP

Once the instrument is assembled and installed, follow the steps below to start the titrator.

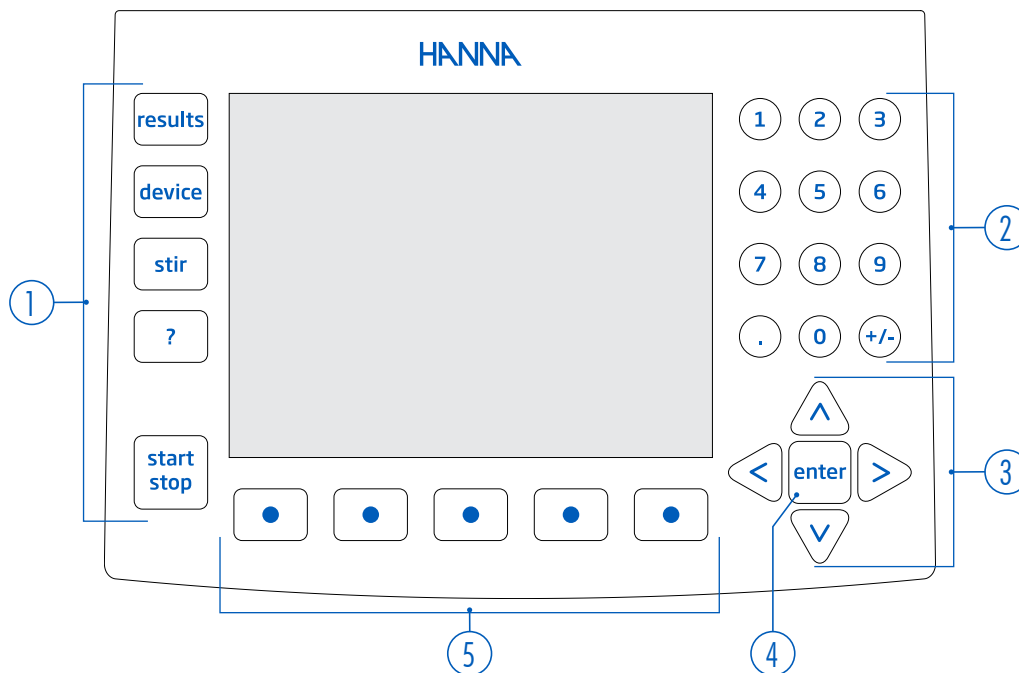
1. Connect the titrator to a power outlet with the supplied power adapter.
2. Turn on the titrator using the power button located on the back of the instrument.
3. Wait until the titrator finishes the initialization process.
4. Press when prompted or wait a few seconds for titrator to start.



Note: All the performed initialization processes must be successfully completed. If one fails, restart the titrator. If the problem persists, contact your nearest Hanna Instruments® Service Center.

2.2.2. KEYPAD

The titrator's keypad is grouped into five categories.



1. Function keys
2. Numeric keys
3. Arrow keys
4. Enter key
5. Option keys

Function Keys

If one of the function keys is pressed, the associated function is immediately performed.

Some of the keys are active only in specific screens.



Starts or stops a titration process



Turns the selected stirrer on and off



Access the autosampler



Access the data parameters menu (reports, GLP, meter information, report setup)



Displays contextual help

Option Keys

Option keys are assigned to the virtual keys on the display.

Their functions are listed in the boxes above the buttons and vary depending on the displayed screen.

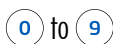
An underlined virtual key can also be activated by pressing .

Arrow Keys

Arrow keys have the following functions:

- move the on-screen cursor
- increase or decrease the stirrer speed and other settings
- select a character (alphanumeric screen only)
- navigate through menu options

Numeric Keys



Used for numeric entries.



Toggles between positive and negative values.



Used for decimal point.

Enter Key

This key has the following functions:

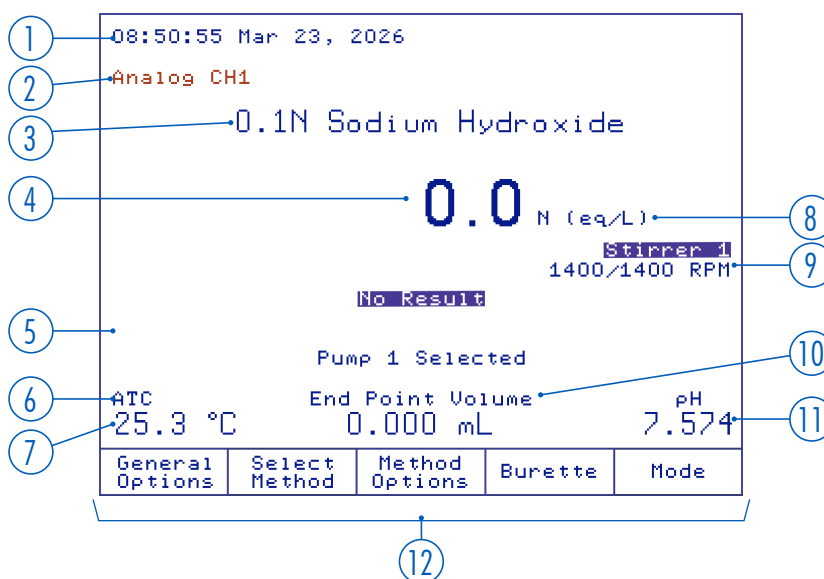
- accepts alphanumeric data entry
- executes the default (underlined) virtual option key

2.2.3. DISPLAY

The titrator has a large color graphical display.

The main screen is shown below with short explanations of the screen segments.

The Main Screen



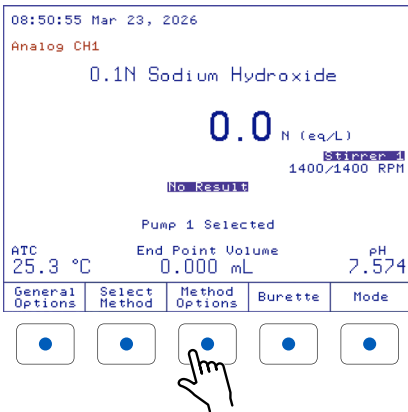
- | | |
|------------------------------------|--------------------------------|
| 1. Time and date | 7. Temperature reading |
| 2. Active input channel | 8. Results units |
| 3. Method name | 9. Stirrer information |
| 4. Result | 10. End point titration volume |
| 5. Reminders or Warnings | 11. mV or pH reading |
| 6. Temperature compensation status | 12. Virtual option keys |

The user interface contains several screens. For each titrator function, several screens may be used.

Method name	Displays the name of the selected method.
Time and date	Displays the current date and time.
Temperature reading	Displays the measured temperature.
ATC	Automatic temperature compensation
Manual	Manual temperature compensation
Manual	Temperature probe is not connected, manual temperature compensation
Stirrer information	The selected stirrer, actual and set stirrer speed is displayed in RPM. When stirrer is off, the stirrer information is not displayed.
Endpoint volume	Displays the volume delivered to reach the titration endpoint. When no titration has been performed, the displayed volume is "0.000 mL".
Result	Displays the titration result or the direct reading measurement.
mV or pH reading	Displays the current reading. The reading will be in mV or pH.
mV	Indicates actual potential reading.
rel mV	Indicates relative potential reading.
pH	Indicates actual pH value.
Titration status	Displays the status of the selected titration. No Result is displayed when a titration has not been performed.
Reminders	Indicates when a task needs to be performed and displays errors.
Pump # selected	Displays the active pump.
Analog CH #/ Digital CH#	Active measurement input.

2.2.4. MENU NAVIGATION

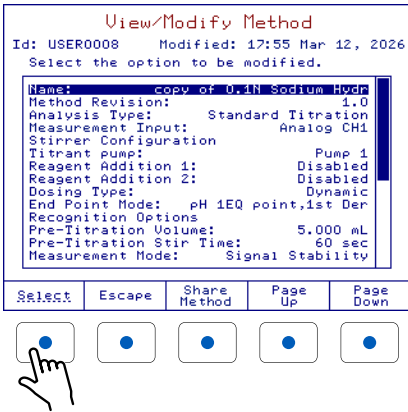
Selecting an Option



Press the option key below the virtual key.

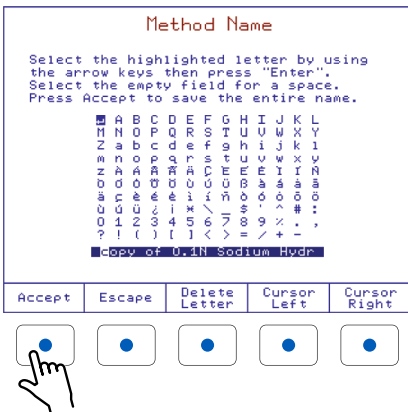
For example, to access the **Method Options** screen, press the option key below it.

Selecting a Menu Item

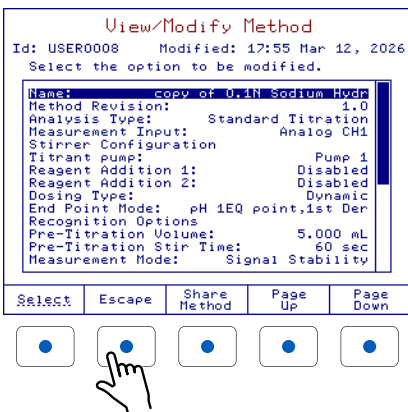


- Use the and arrow keys to move the cursor. When the menu is larger than the display, a scroll bar is active on the right side.
- Use the and keys to scroll through the pages.
- Press or to activate the selected menu item.

Entering Text

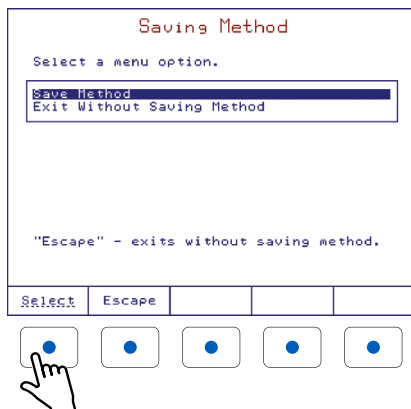


- Use to erase previous text.
- Use the arrow keys to highlight the letter then press . Use the same procedure to enter the whole name.
- Use the and keys for editing. When editing is complete, press . The method name will be updated and displayed in the name field of the **View / Modify Method** screen.



When all the desired parameters have been set, press .

Saving Modifications



The **Saving Method** screen allows the user to save the modifications.

- To save the modifications, highlight *Save Method* option, then press `Select`.
- To exit without saving, press `Escape`.

Alternatively, highlight *Exit Without Saving Method* option then press `Select`.

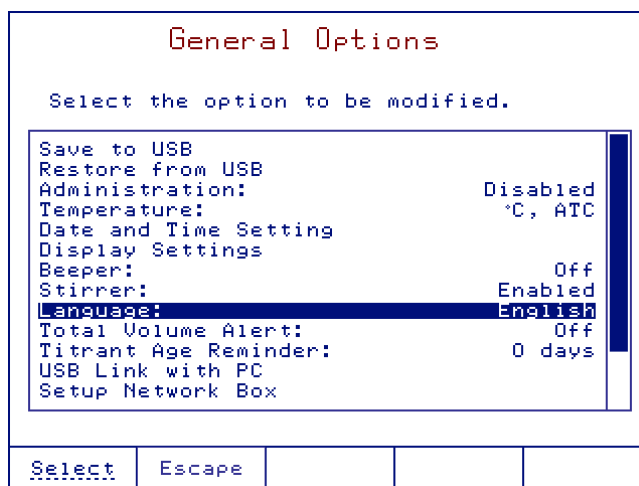
Note: To access the contextual help menu, press `?` at any time. Help is related to the displayed screen.

Press `Escape` or `?` to return to the previous screen.

2.3. GENERAL OPTIONS

The **General Options** screen gives access to options that are not directly related to the titration process or pH/mV/ISE measurement.

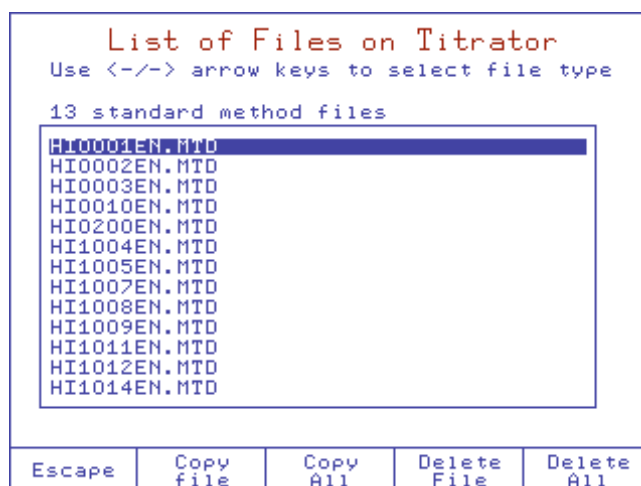
To access this screen, press `General Options` from the main screen.



2.3.1. SAVE TO USB

This option allows the user to save files from the titrator to a USB storage device.

Note: The USB storage device has to be formatted FAT or FAT32.








On the titrator, the available file types are as follows:

Standard method HIXXXXXY.MTD (e.g.: [HI0001EN.MTD](#), [HI1004EN.MTD](#))
User-defined method USERXXXX.MTD (e.g.: USER0001.MTD)
Report Ti_XXXXX.RPT, mV_XXXXX.RPT, pH_XXXXX.RPT, ISEXXXXX.RPT, mVrXXXXX.RPT (e.g.: Ti_00001.RPT, mV_00001.RPT, pH_00001.RPT, ISE00001.RPT, mVr00001.RPT)

- Insert the USB storage device into the USB port on the right side of the titrator.
- Use the < and > keys to select the file type.
The number of files and the file names will be displayed.
- Use the ^ and v keys to scroll through the list.

The option keys allow the following operations:

 key returns the user to the **General Options** screen.
 key copies highlighted file from the titrator to USB storage device.
 key copies all displayed files from the titrator to USB storage device.
 key deletes the highlighted file.
 key deletes all displayed files.

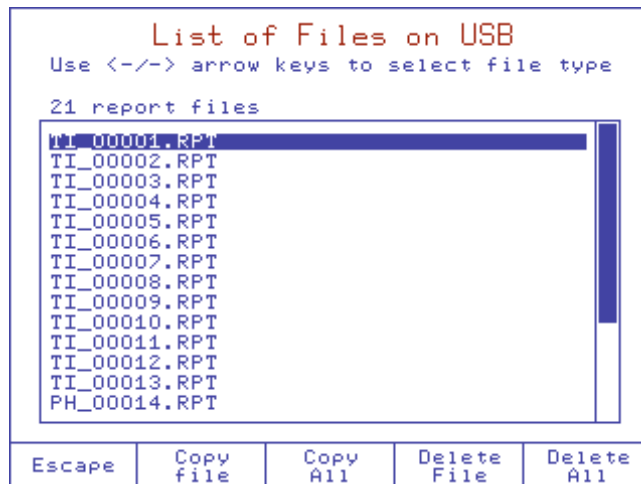
*Note: The saved files will be stored on the USB key in the **HI932R** folder, as follows:*

Methods USB Drive\HI932R\Methods*.mtd

Reports USB Drive\HI932R\Reports*.rpt

2.3.2. RESTORE FROM USB

This screen allows the user to transfer files from the USB storage device to the titrator.








The file types that can be transferred are:

Standard method HIXXXXXY.MTD (e.g.: [HI0001EN.MTD](#), [HI1004EN.MTD](#))
User-defined method USERXXXX.MTD (e.g.: USER0001.MTD)
Report Ti_XXXXX.RPT, mV_XXXXX.RPT, pH_XXXXX.RPT, ISEXXXXX.RPT, mVrXXXXX.RPT (e.g.: Ti_00001.RPT, mV_00001.RPT, pH_00001.RPT, ISE00001.RPT, mVr00001.RPT)

- Insert the USB storage device into the USB port on the right side of the titrator.
- Use the < and > keys to select the file type.
The number of files and the file names will be displayed.
- Use the ^ and v keys to scroll through the list.

The option keys allow the following operations:

-  key returns the user to the **General Options** screen.
-  key copies the highlighted file from the USB storage to the titrator.
-  key copies all displayed files from the USB storage to the titrator.
-  key deletes the highlighted files from the USB storage device.
-  key deletes all displayed files from the USB storage device.

Note: To restore files from USB key, please ensure that the methods and/or reports required for transfer to the titrator are in the correct folder:

Methods USB Drive\HI932R\Methods*.mtd

Reports USB Drive\HI932R\Reports*.rpt

2.3.3. ADMINISTRATION

A 4-digit numeric PIN can be set to prevent unauthorized changes from being made.

When the user enters administration mode and a pin has not been set, the user will be prompted to enter a new PIN.



Titration Administration

Administrator PIN has not been set.
Enter a 4-digit PIN to enable
Administrator function.

Enter PIN:

Confirm PIN:

Your PIN must be 4-digits long.

Next	Escape	Delete Digit	
------	--------	-----------------	--

Once a PIN has been set, the titrator can be locked. When a titrator is locked, the users cannot modify methods or delete reports. Basic functions are still available (review reports, save to USB, etc.).



Titration Administration

Titrator is UNLOCKED.

Lock Titrator

Enter PIN:

ACCEPT	Escape	Delete Digit	
--------	--------	-----------------	--

To return to administration mode, enter the PIN to unlock the titrator.

Titrator Administration				
Titrator is LOCKED.				
Unlock Titrator	Escape			Recovery PIN

If the PIN is lost or forgotten, press Recovery
PIN and contact technical support to supply the required information.

Recovery PIN				
For recovery PIN, please contact your vendor. When requesting PIN please provide following information:				
Titrator Serial Number: 12345678				
Code: 0078				
Recovery PIN: -----				
Accept	Escape	Delete Digit		

2.3.4. TEMPERATURE

The temperature menu allows access to the three temperature-related menu options: source, setting, and units.

Temperature Menu							
Select temperature option to be modified.							
<table border="1" style="width: 100%;"> <tr> <td style="background-color: black; color: white;">Temperature Source</td> </tr> <tr> <td>Manual Temperature Setting</td> </tr> <tr> <td>Temperature Units</td> </tr> </table>					Temperature Source	Manual Temperature Setting	Temperature Units
Temperature Source							
Manual Temperature Setting							
Temperature Units							
Select	Escape						

Temperature Source

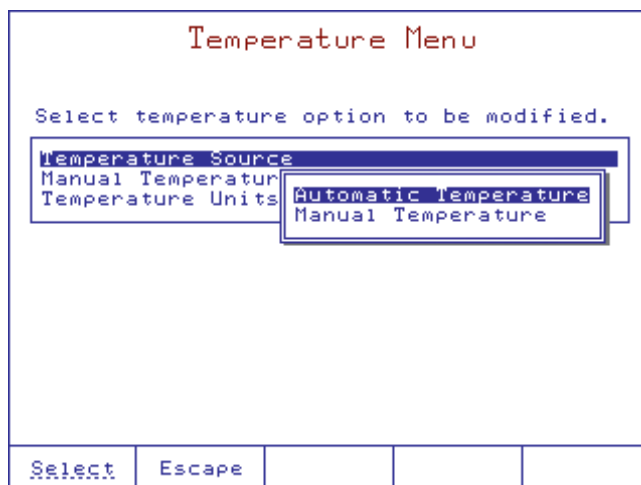
Option: Automatic Temperature or Manual Temperature

Select the temperature source used for temperature compensation.

When Automatic Temperature is selected, "ATC" is displayed on the main screen and temperature is read by the temperature probe.

When Manual Temperature is selected, "Manual" is displayed on the main screen and a preset temperature value is used for temperature compensation.

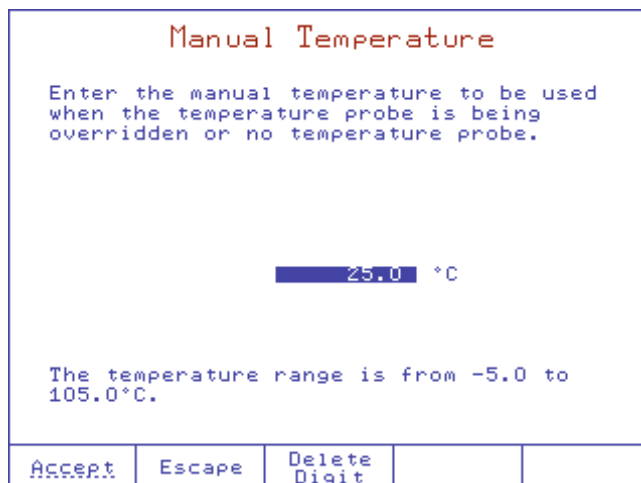
Note: The selected temperature source will be indicated in the report files: "A" for Automatic and "M" for Manual.



Manual Temperature Setting

Option: -5.0 to 105.0 °C (23.0 to 221.0 °F, 268.2 to 378.2 K)

If the temperature probe is not connected, the user can manually set the temperature used by the titrator for compensation.



Temperature Units






Option: °C, °F, K


The temperature ranges are as displayed in the **Temperature Units** screen.

Temperature Menu													
Select temperature option to be modified.													
Temperature Source													
Manual Temperature Setting													
Temperature Units													
<table border="1"> <tbody> <tr> <td>Celsius</td> <td>-5.0 to 105.0</td> <td>°C</td> </tr> <tr> <td>Fahrenheit</td> <td>23.0 to 221.0</td> <td>°F</td> </tr> <tr> <td>Kelvin</td> <td>268.2 to 378.2</td> <td>K</td> </tr> </tbody> </table>					Celsius	-5.0 to 105.0	°C	Fahrenheit	23.0 to 221.0	°F	Kelvin	268.2 to 378.2	K
Celsius	-5.0 to 105.0	°C											
Fahrenheit	23.0 to 221.0	°F											
Kelvin	268.2 to 378.2	K											
Select	Escape												

2.3.5. DATE & TIME SETTING





This screen allows the user to set the date and time.

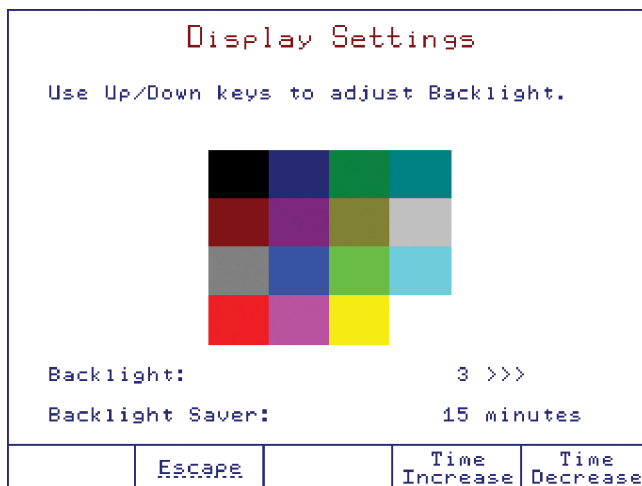
- Use the  and  keys or the numeric keys to modify the date and time.
- Use  to move the cursor to the next field.
- Use  or  to change the time format.

Date and Time Setting				
Enter the date.				
	03	2026		
day	month	year		
Enter the time.				
20	41	41		
hour	minute	second		
Press <Next> to move to the next entry.				
Accept	Escape	Delete Digit	Next	AM/PM

2.3.6. DISPLAY SETTINGS

This screen allows the user to customize the display settings.

- Use  to increase the backlight time-saver interval.
 - Use  to decrease the backlight time-saver interval.
 - Use the  and  keys to adjust the backlight intensity.
- There are 8 levels of backlight intensity, ranging from 0 to 7.



The displayed color palette allows for selection of appropriate backlight intensity.

The backlight time-saver option protects the display during standby periods, when no keys have been pressed for a set amount of time, the backlight will turn off.

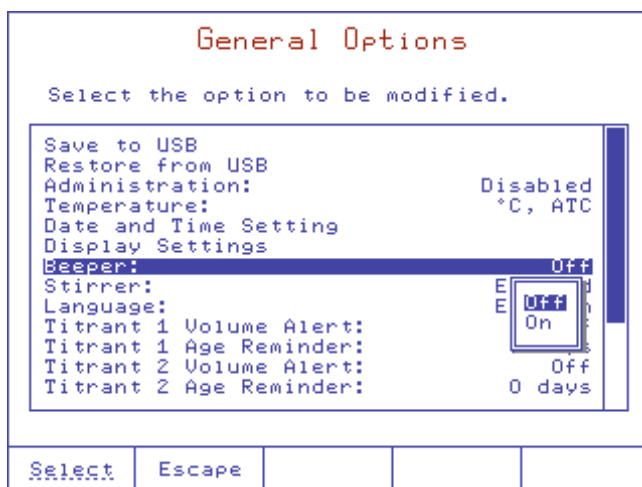
If the backlight is off, press any key to reactivate the backlight.

The range for backlight time-saver interval is between 1 and 60 minutes. To disable the backlight time-saver, increase the time to the maximum allowed, the Off indication will be displayed.

2.3.7. BEEPER

Option: On or Off

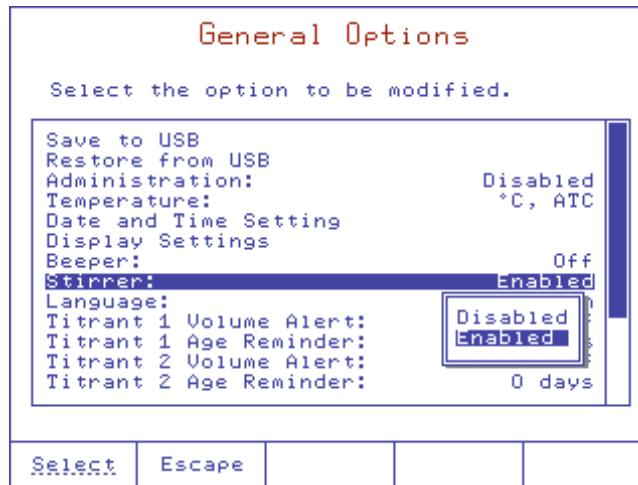
If enabled (on) an audible alert will sound after a titration is completed, when an invalid key is pressed, or when a critical error occurs during titration.






2.3.8. STIRRER

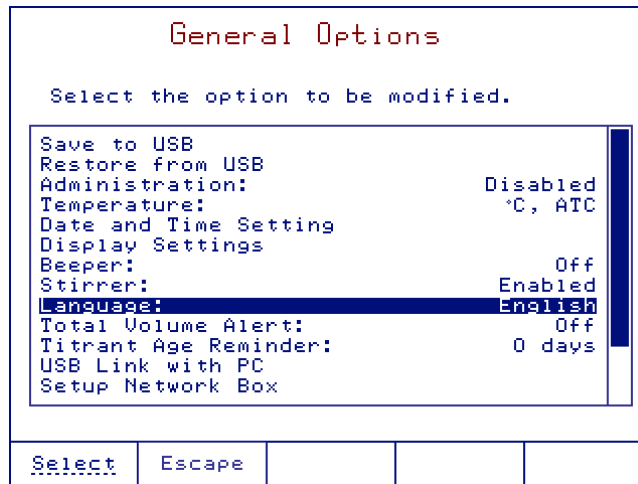
Option: Enabled or Disabled

If necessary, the stirrer can be disabled in individual titration method.



2.3.9. LANGUAGE

- Use the  and  keys to select the language from the options listed.
- Press .
- Restart the titrator in order to apply the new language setting.



2.3.10. TOTAL VOLUME ALERT

Option: 0 to 10000 mL, Off

This screen allows a programmable reminder to appear when the titrant reservoir is below 100 mL.

The titrant volume will decrease as the titrant is used.

After the new titrant volume has been entered in the **Total Volume Alert** screen, a warning message appears on the main screen reminding the user to re-standardize the newly added titrant.

Press to disable this option.

Titrant 1 Volume Alert				
Enter the amount of titrant available to the titration/reagent system from its reservoir. The mLs will decrease as the titrant/reagent is depleted.				
1000 mL				
A reminder will appear when less than 100 mLs of titrant volume is left.				
ACCEPT	Escape	Delete Digit		Off

2.3.11. TITRANT AGE REMINDER

Option: 0 to 31 days, Off

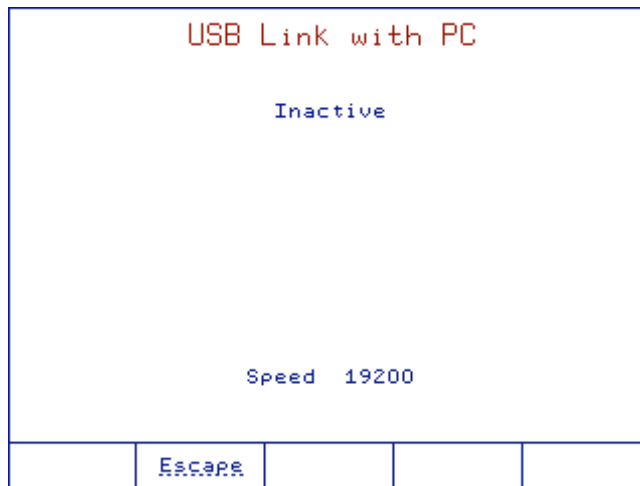
A programmable reminder will appear when it is time to verify the titrant concentration or to change the titrant.

Press to disable this option.

Titrant 1 Age Reminder				
Enter the number of days to pass since the last Titr. Vol. updating or the last Start pressing, whereafter the reminder appears.				
30 days				
The range is from 0 to 31 days.				
Start	Escape	Delete Digit		Off

2.3.12. USB LINK WITH PC

In order to use this feature, the USB cable needs to connect the titrator with the PC. Make sure that HI900 PC application is running on the PC.





“Active/Inactive” message shows the status of the USB link with the PC.

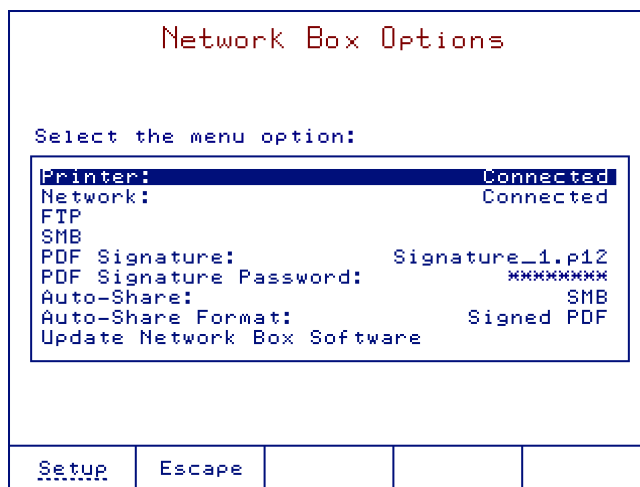
- » “Active” indicates that the titrator is using the USB communication with the PC and not with another device.
- » “Ready” indicates that the titrator is able to communicate with the PC.
- » During transfer of information between the PC and the titrator, press “Transmit” and the status is displayed.

2.3.13. NETWORK BOX SETUP

In order to use this feature, the HI932933 Network Box needs to be connected to the HI932 Titrator. The Network Box menu allows access to:

- Printer settings
- Network configuration
- FTP and SMB sharing
- PDF signature options
- Auto-Sharing
- Network Box software updates.

Use the  and  keys to highlight item to be modified.



Printer Setup

Option: Printer Name, Manufacturer, Page Size

Printer Setup

Select the option to be modified.

Printer Name:	Canon MF210 Series
Manufacturer:	Canon
Page Size:	A4

A5
 A4
 A3

Select Escape

If no printer is detected, Printer Name and Manufacturer fields are blank.

Network Setup

Option: Ethernet, Disabled

- Use the and keys to highlight option and press .

Network Setup

Select the option to be modified.

Network:	Disabled
----------	----------

Disabled
 Ethernet

Select Escape

With Ethernet connection established, IP assignment can be set as:

- Dynamic » IP Address, Gateway, Netmask, DNS Server are auto assigned
 Network Setup screen displays the currently assigned IP address, which is automatically provided by the DHCP server.
- Static » network details are filled in manually

Network Setup

Select the option to be modified.

Network Type	Ethernet
IP:	Dynamic
IP Address:	
Gateway:	
Netmask:	
DNS:	
MAC:	01:a2:b3:c4:d5:e6

Dynamic
 Static

Select Escape

Network Setup

Select the option to be modified.

Network Type	Ethernet
IP:	Static
IP Address:	192.168.1.100
Gateway:	192.168.1.1
Netmask:	255.255.255
DNS:	8.26.56.20
MAC:	01:a2:b3:c4:d5:e6

Select Escape

FTP Setup

FTP Requirements: Server name/IP address, Username, Password, Protocol, TLS Certificate, Remote Dir.

- Use the Δ and ∇ keys to highlight item to be modified.

Server name /IP address Fill in server name or IP address where the server is located.

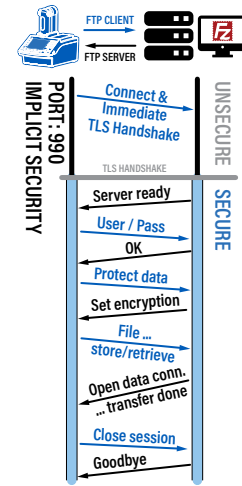
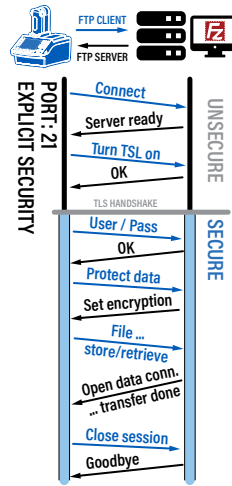
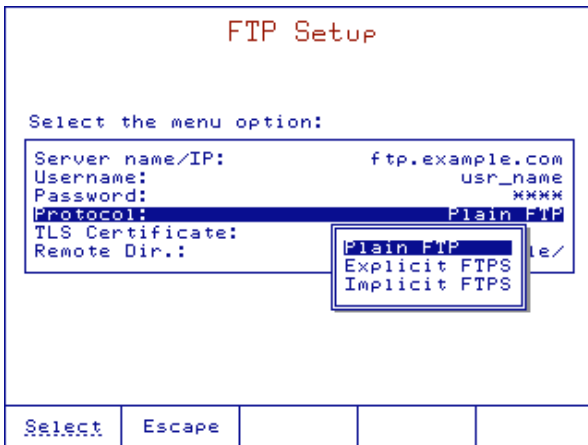
Username Fill in specific username for the server.

Password Enter user password for the server.

Protocol Select required FTP protocol (Plain FTP/Explicit FTPS/Implicit FTPS).

TLS Certificate TLS/SSL certificates are external certificates for FTPS, used to secure in-transit data during file transfer. TLS certificates are imported and installed from the USB device.

Remote Dir. Enter remote directory path, i.e. host system to which users are exporting.



The transition from unsecured to secured FTP file transfers is based on port selection and TLS certificate availability.

- Standard (Plain) FTP transfer operates without a TLS certificate, uses Port 21, all communication is unencrypted.
- Explicit FTPS transfer requires a TLS certificate, uses Port 21, handshake is not encrypted, file transfer is encrypted.
- Implicit FTPS transfer requires a TLS certificate, uses Port 990, both handshake and file transfer are encrypted.

Import TLS Certificate

- With the USB device plugged in, use $\left[\begin{smallmatrix} \text{Import} \\ \text{from USB} \end{smallmatrix} \right]$ to import a TLS certificate.

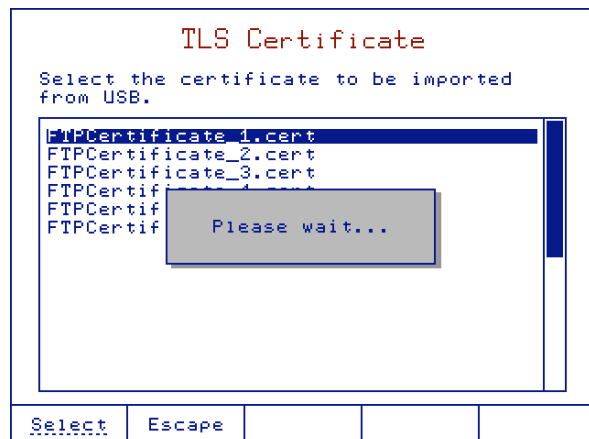
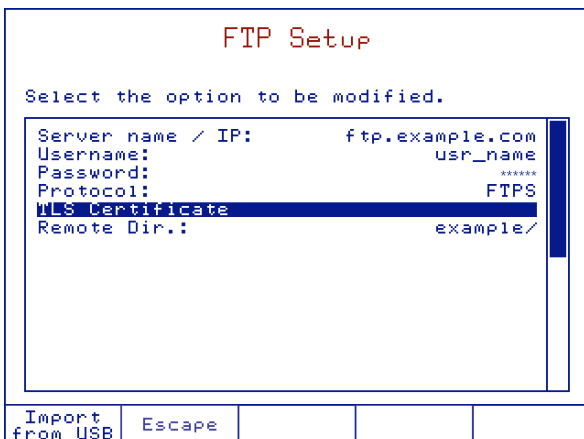
Certificates can be imported as .cer, .crt, or .pem files. Only one certificate can be loaded at a time.

“Please wait...” message is displayed as the file is being loaded.





A confirmation screen appears once the file is successfully copied.

- Use the \leftarrow and \rightarrow keys to select the certificate type, and the Δ and ∇ keys to select a certificate.
- Certificates must be stored in the following directory: USB Drive\HI932R\CERTS*.cert

Note: It is recommended to contact local IT department and have the server and certificate securely installed.



Clear TLS Certificate



- Use the  and  keys to highlight option.
- Use  to delete existing certificate file. A warning screen will appear.
- Use  to delete.

FTP Setup	
Select the option to be modified.	
Server name / IP:	ftp.example.com
Username:	usr_name
Password:	*****
Protocol:	FTPS
TLS Certificate:	CurrentBoxCertName
Remote Dir.:	example/
<input type="button" value="Clear"/> <input type="button" value="Escape"/> <input type="button" value=""/> <input type="button" value=""/> <input type="button" value=""/>	

WARNING	
Clear TLS Certificate	
Are you sure you want to delete the certificate currently used by the Network Box?	
<input type="button" value="Continue"/> <input type="button" value="Escape"/> <input type="button" value=""/> <input type="button" value=""/> <input type="button" value=""/>	

SMB (Server Message Block) Setup

SMB Requirements: Server name/IP address, Username, Password, SMB Domain, SMB Share, Remote Dir., Port

- Use the  and  keys to highlight option to be modified.

Server name /IP address Fill in server name or IP address where the server is located.

Username Fill in specific username for the server.

Password Enter user password for the server.

SMB Domain Fill in domain used for centralized authentication (Active Directory domain).

SMB Share Enter the name of the local folder path for the shared folder on the server.
The name should not contain spaces or start with a dot.

Note: ensure the user permissions have been set on the server to allow access with the user information entered above.

Remote Dir. Enter name of remote directory path, local directory tree on the host system to which users are exporting files.
A directory will be created if it does not already exist.

Port Port number is defaulted to 445.

Sever, Domain, and Share are the three components central to SMB transfer.

The minimum configuration for SMB to function requires Server, Share, and Domain components.

The SMB Server component must be active to share resources.

The SMB Share component is the designated shared resource.

The password opens access to the shared component (either for local users or for centralized Active Directory domains).

Note: The network box supports transfer to encrypted SMB. No certificates are required to transfer files over encrypted SMB (3.0). It is recommended to contact local IT department regarding proper SMB setup.

SMB Setup	
Select the option to be modified.	
Server name / IP:	ftp.example.com
Username:	usr_name
Password:	*****
SMB Domain:	LAB
SMB Share:	example
Remote Dir.:	example/
Port:	445
<input type="button" value="Select"/> <input type="button" value="Escape"/> <input type="button" value=""/> <input type="button" value=""/> <input type="button" value=""/>	

PDF Signature

PDF signatures are imported from a USB device to authenticate and validate digital files.

The USB device contains a private key and a digital certificate, which, when accessed, allows for electronic signature.

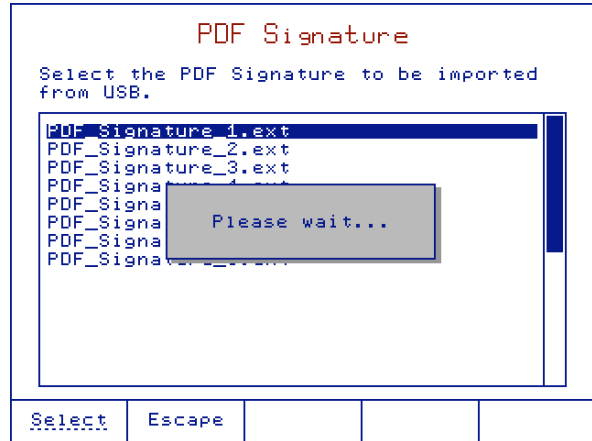
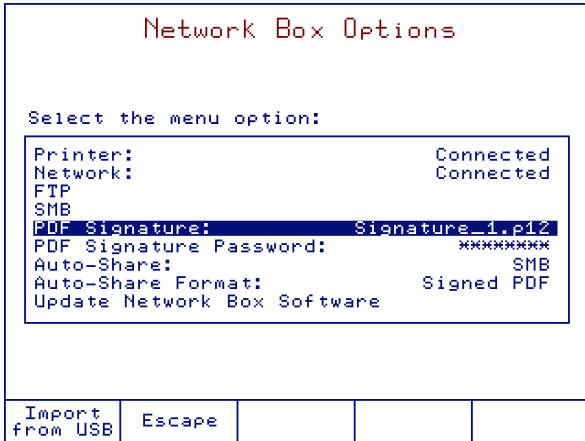
The signature is a cryptographic operation that ensures the document has not been altered since it was signed.

1. Plug in the USB device.

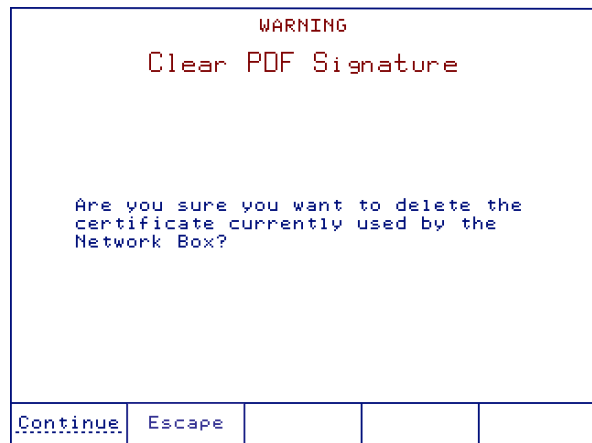
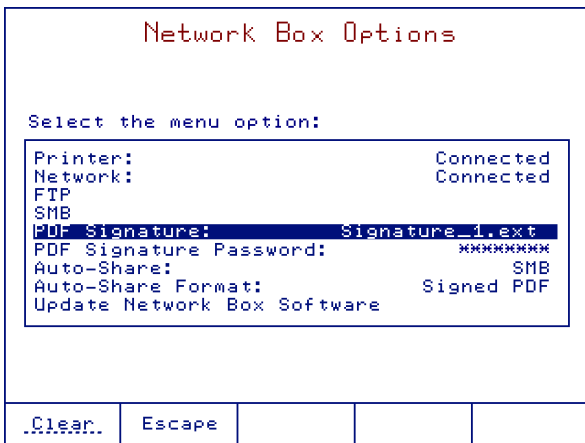
Certificates must be stored in the following directory: USB Drive\HI932R\CERTS*.p12

2. Once the USB is detected, "Please wait..." message is displayed while the file is being uploaded.
3. The document is scanned and the signature applied.

The signed PDF is automatically sent to the designated FTP or SMB server.

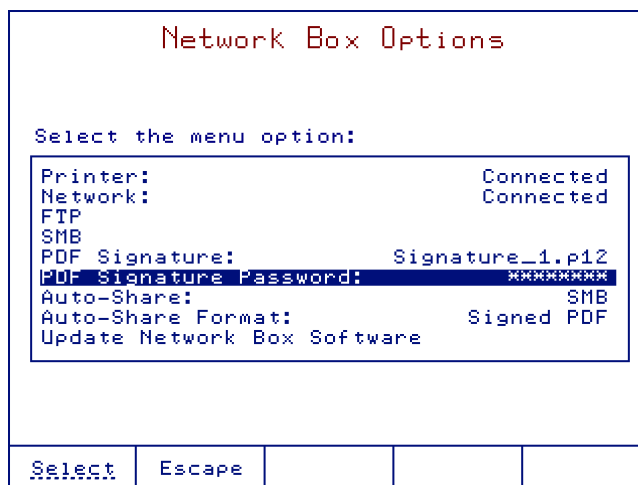


4. To delete a digitally signed certificate, use Clear key. A warning screen will appear.
5. Use Continue to delete.



PDF Signature Password

A PDF digital signature password protects the private key used in a digital certificate to sign documents, ensuring only the authorized user can apply it.



Auto-Share

Option: FTP, SMB, Disable

When auto-share option is enabled, data automatically generated (titration reports, direct readings, automatic logs) are instantly routed to configured network destination.

The feature supports both FTP (File Transfer Protocol) for remote transfers and SMB (Server Message Block) for local network integration.

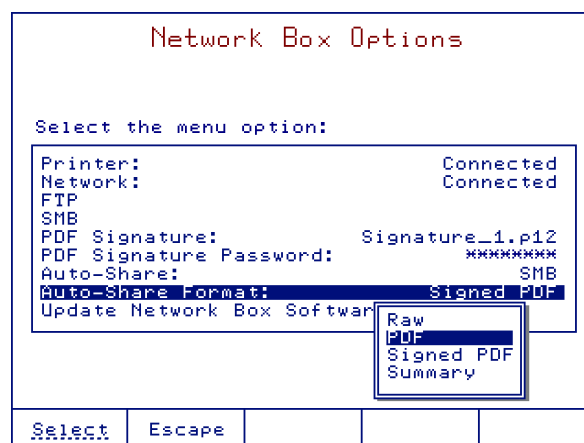
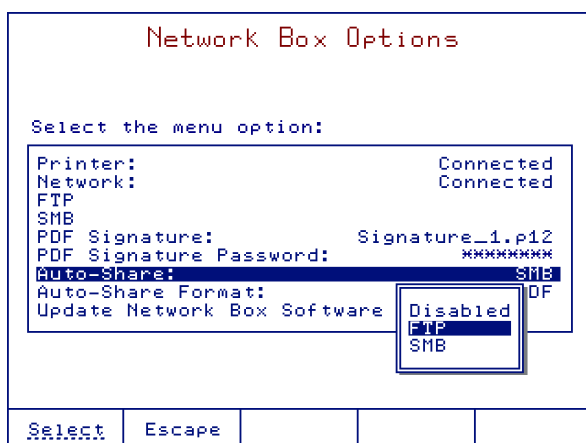
- Reports are shared automatically at the end of titration using the setup configuration.
- Tray reports (Autosampler) are batch-processed and shared after the last report is generated.
- An actionable 5-10 second cancel timeout prevents transfer of erroneous data.
- When connection fails before or during a file transfer, the system automatically displays an error message and saves an error report.

Auto-Share Format

Option: Raw, PDF, Signed PDF, Summary

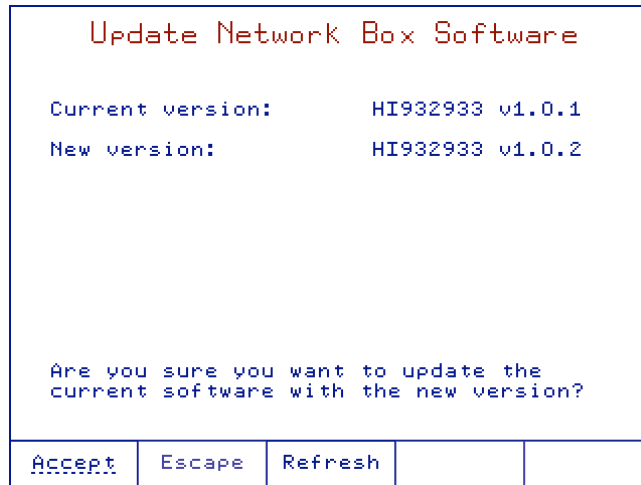
Users can select how the data is saved (file format):

- Raw (text format)
- PDF
- Signed PDF (if a valid signature file has been imported)
- Summary



Update Network Box Software

This screen allows the user to update the network box software from a USB device containing a software setup kit.



To check for new releases, scan the QR code or go to: <https://software.hannainst.com>.

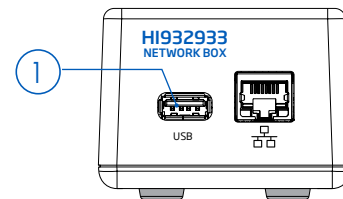
1. Scroll down the software downloads page to find the **Instrument Software** list.
2. Connect the USB-A flash drive to PC.
3. Find the software version needed for download, then click on the Download icon (↓). Wait for the download to complete.
4. Copy the file to a USB storage device.



To update the software:

On HI932933 network box

5. Locate the rear panel of the network box.
6. Plug the USB key directly into the designated USB port (1) on the network box.



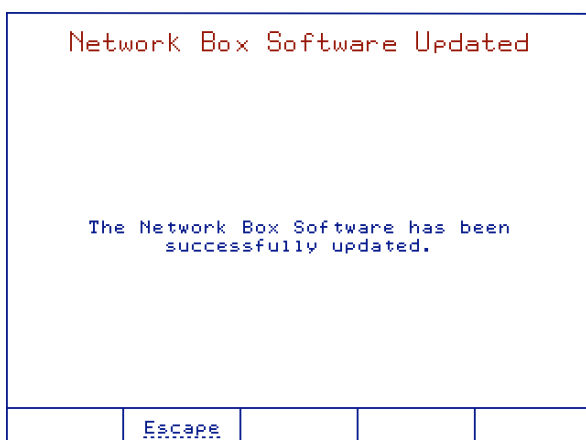
On HI932 titrator

7. Navigate to Network Box Options screen.
8. Use the and keys to highlight *Update Network Box Software* option.
9. Press .

The screen will display both current and new (if available) network box software versions.

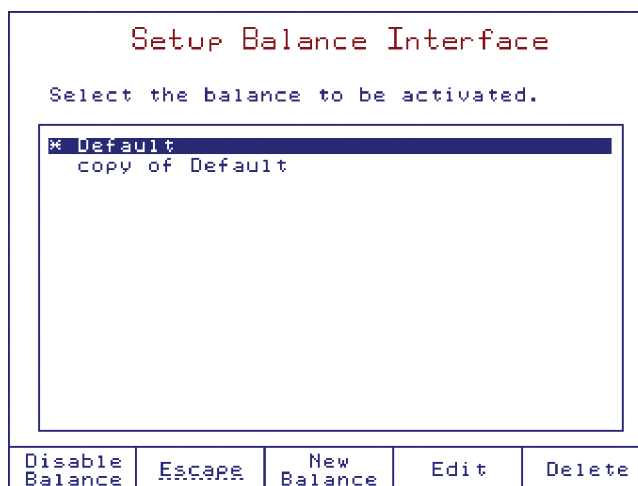
10. Press . "Please Wait" screen is displayed while the update is in progress.

A confirmation screen appears once the update has completed successfully, otherwise, an error message is displayed.









2.3.14. SETUP BALANCE INTERFACE

This screen allows the user to setup an analytical balance for automatic acquisition of sample mass prior to titration or standardization.




The balance is connected to the titrator via RS 232 interface.

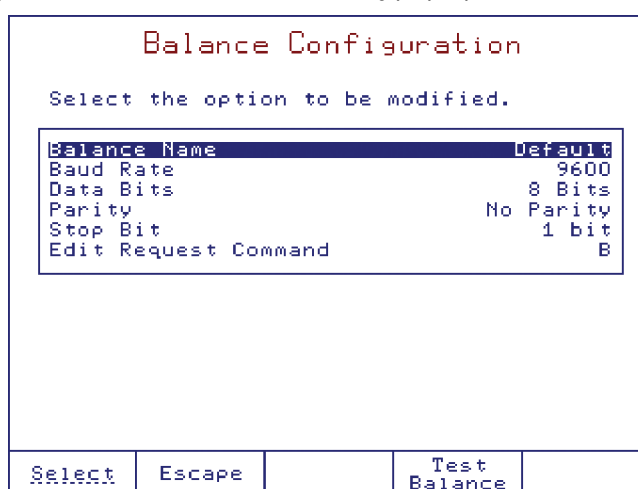
- Press  to enable the selected balance.
- Press  to disable the selected balance.
Automatic weight acquisition is not available.
- Press  to return to the **General Options** screen.
- Press  to add a new balance to the list.
- Press  to customize the serial communication parameters.
The **Balance Configuration** screen will open.
- Press  to delete the highlighted balance.

Note: At least one balance must be in the list.

Be sure that the balance configuration settings match the settings of your balance.

It may be necessary to change settings on your balance or titrator. Users should consult their balance instruction manual.

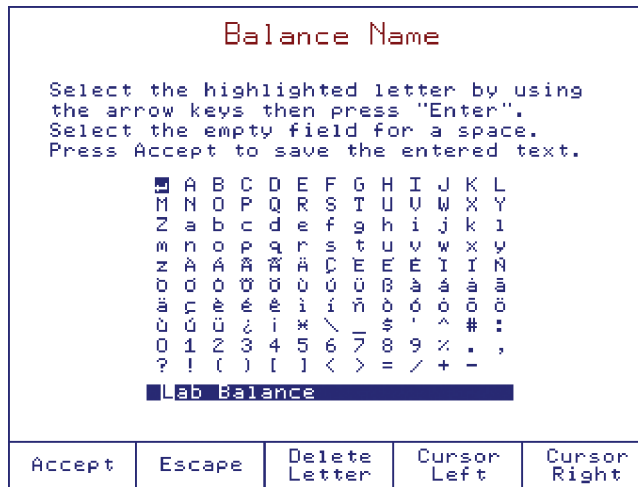
Press the  key to verify connection with the balance is working properly.



Balance Name

Option: Up to 24 characters

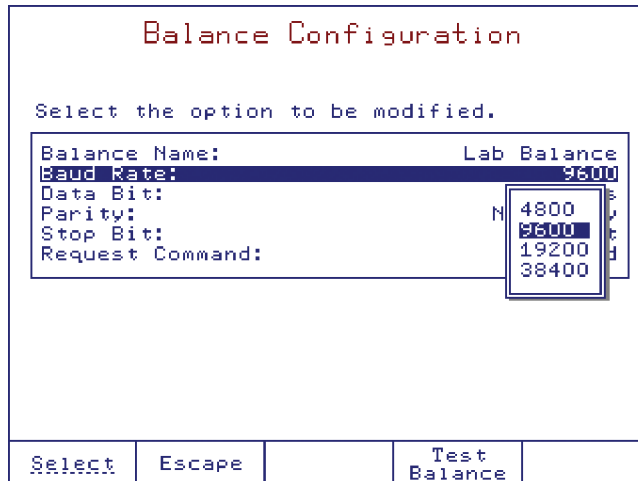
Assign a name for your customized balance.



Baud Rate

Option: 4800, 9600, 19200, 38400

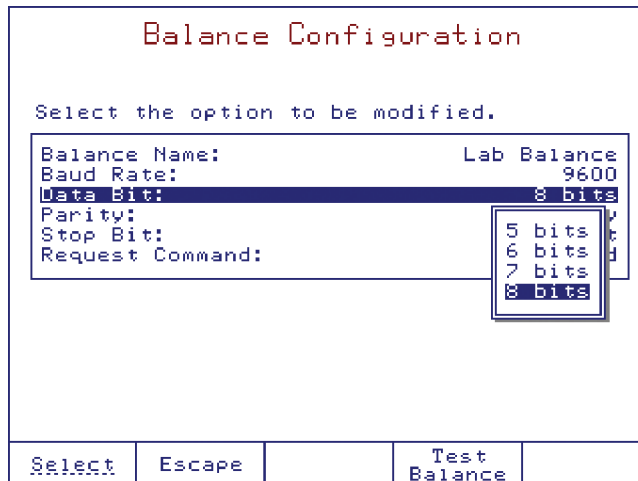
Set the serial communication baud rate.



Data Bits

Option: 5, 6, 7, 8 bits

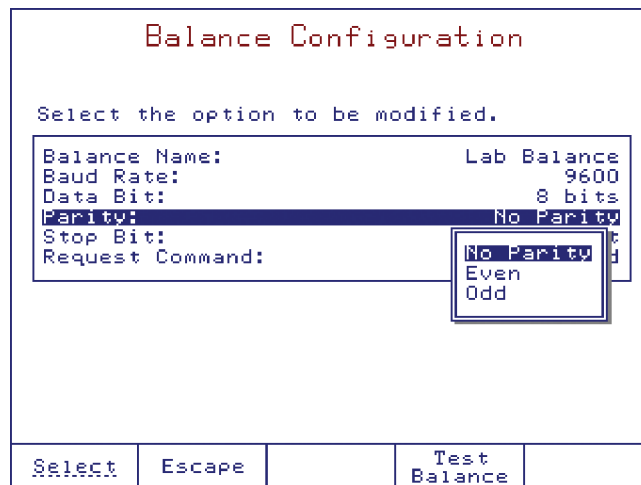
Set the number of data bits.



Parity

Option: No Parity, Even, Odd

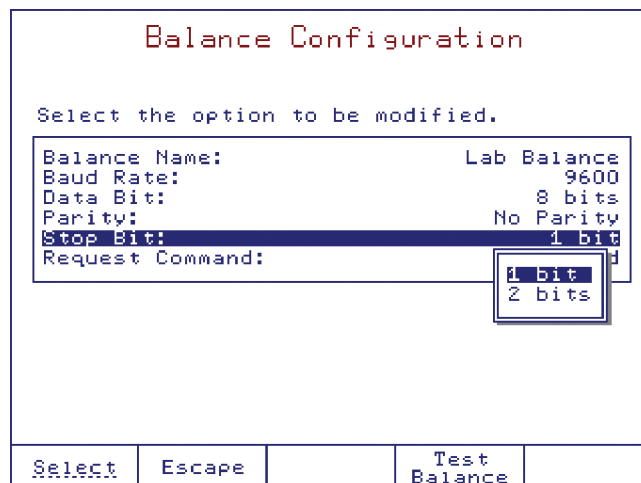
Set the parity of data packet.



Stop Bit

Option: 1 bit or 2 bits

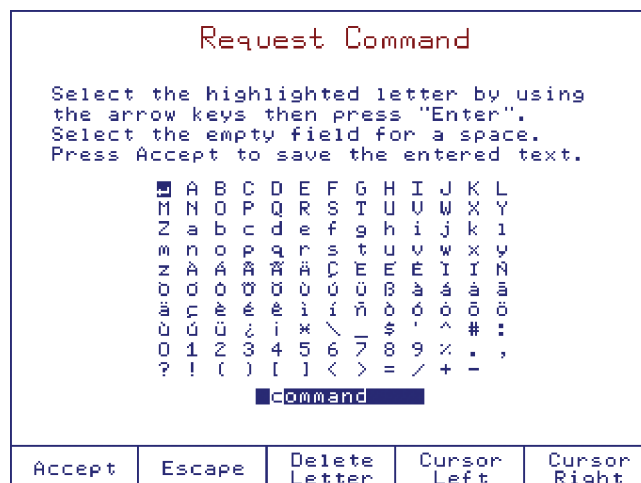
Set the number of stop bits.



Edit Request Stop

Option: Up to 10 characters

Type the syntax for weight request command.



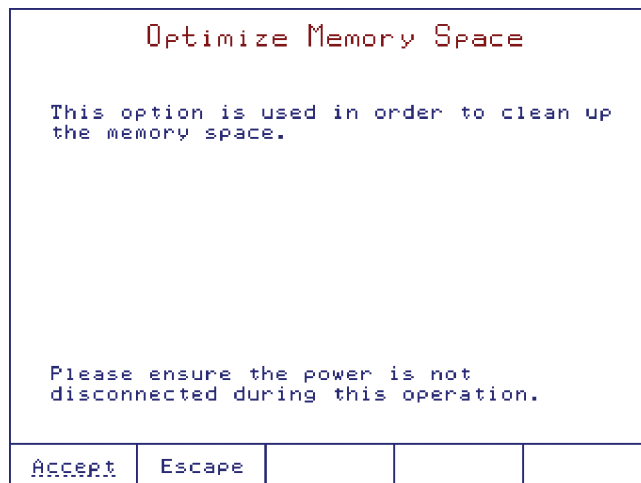
2.3.15. RESET TO DEFAULT SETTINGS

This will delete all user-defined methods and restore all manufacturer settings such as titrator configuration, standard method parameters, etc.



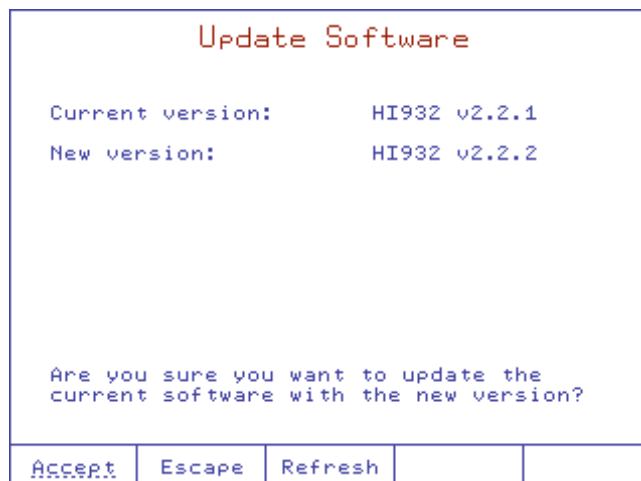
2.3.16. OPTIMIZE MEMORY SPACE

This screen allows the user to run a memory defragmentation utility to increase access speed to memory storage. Press Accept and then restart the titrator. Do not disconnect the power supply during this operation.



2.3.17. UPDATE SOFTWARE

This screen allows the user to update the titrator software from a USB storage device containing a software setup kit.



To check for new releases, scan the QR code or go to: <https://software.hannainst.com>.



To update the software:

1. Scroll down the software downloads page to find the **Instrument Software** list.
2. Connect the USB storage device to PC.
3. Find the software version needed for download, then click on the Download icon (↓).
4. Wait for *.zip file download to complete.
5. Unzip the file and copy the obtained "UPD932R" folder to a USB storage device.
6. Insert the USB storage device into the USB port.
7. Go to **General Options**, then **Update Software**.

The titrator will display the current and new software versions.

8. Press .

When prompted, remove the USB storage device and restart the titrator.

2.4. TITRATION METHODS

All parameters required to complete an analysis are grouped into a method.

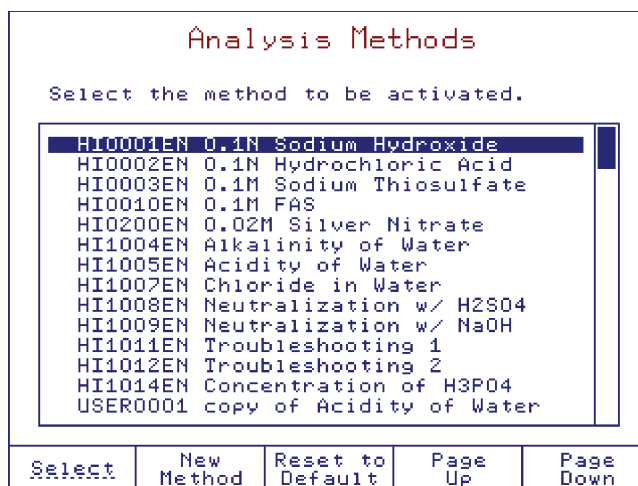
The titrator is supplied with a pack of standard methods. These methods have been developed by Hanna Instruments® and can be used to create user-defined methods.

Standard and user-defined methods can be upgraded, saved, or deleted by connecting the titrator to a PC using the **HI900** PC application or a USB flash drive.

2.4.1. SELECTING METHODS

To select a method, press from the main screen.

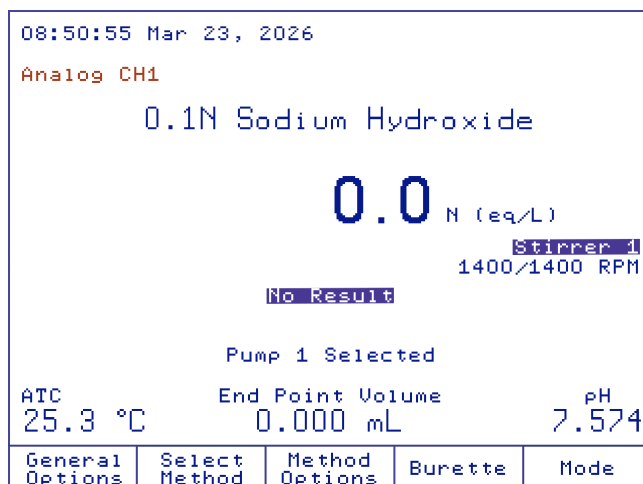
A list of available methods will be displayed.



The list of all available methods (standard and user-defined methods) can be viewed in the **Analysis Methods** screen.

To select a method, highlight the method then press .

The name of the selected method will be displayed on the main screen.



2.4.2. STANDARD METHODS

The standard methods are developed for the most common types of analysis and can be used as templates to create new user-defined methods. Only specific method parameters can be modified by the user.

See [2.4.5. Method Options](#) section for more information.

Upgrading Standard Methods

To upgrade the titrator with new standard methods, follow the steps below.

From USB storage device

1. Insert the USB storage device into the USB port, located on the right side of the titrator.
2. Press from the main screen.
3. Use and keys to highlight *Restore from USB* option. Choose .
4. Use and keys to navigate through file types to find “standard method files”.
5. Press the or key to upgrade the titrator with the standard methods.
6. Press to return to **General Options** screen.

From PC

Using the **HI900** PC application, the titrator can be upgraded with standard methods from a PC.

See [2.3.12. USB Link with PC](#) section for more information.

Deleting Standard Methods

Standard methods can be removed from the titrator by following one of the procedures below.

From General Options Screen

1. Use the and keys to highlight *Save to USB* option. Press .
2. Use the and keys and navigate through the file types menu to find the list of “standard method files”.
3. Press the or keys to remove unnecessary standard methods.
4. Press to return to the **General Options** screen.

From PC

The not required standard methods can be removed from the titrator using the **HI900** PC application.

See [2.3.12. USB Link with PC](#) section for more information.

Restoring the Standard Methods to the Manufacturer Settings

To restore the standard methods to the default settings, press  and then press .






Confirmation of Reset Methods				
Are you sure you want to reset all Standard Methods to default?				
Reset	Escape			

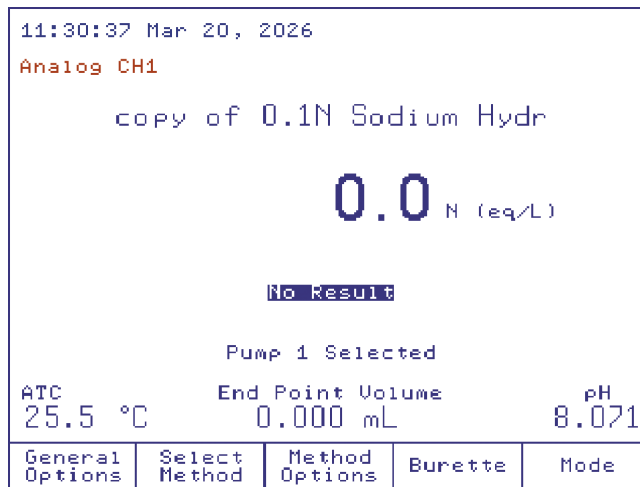
2.4.3. USER-DEFINED METHODS

User-defined methods are created by users, by modifying a standard method or previously created user-defined method. All method parameters can be modified to suit user-specific requirements.

Creating User-Defined Methods





To create a new user-defined method, start from a standard or previously generated user-defined method and follow the steps below.

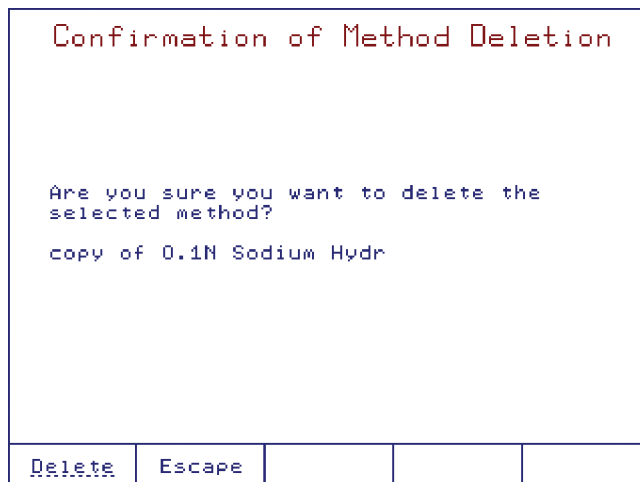
1. Press  from the main screen.
 2. Use the  and  keys to highlight an existing method from the method list.
 3. Press .
- A new user-defined method will be generated.
4. Press  to activate the new method.



Note: The titrator can hold 100 methods (standard and user-defined). When the limit is reached, a warning message is displayed.

Deleting User-Defined Methods

1. To remove a user-defined method, press  from the main screen.
 2. Highlight the user-defined method that you want to delete and press .
- A confirmation screen will appear.
3. Press  again to confirm, or press  to cancel the operation.



2.4.4. VIEWING / MODIFYING METHOD

- To modify the method parameters, press Method Options from the main screen.
A list of all the parameters for the selected method will be displayed.
- Press the ▲ and ▼ keys to highlight the option you want to modify.
- Choose Select.

View/Modify Method

Id: USER0008 Modified: 17:55 Mar 12, 2026

Select the option to be modified.

Name:	copy of 0.1N Sodium Hydr
Method Revision:	1.0
Analysis Type:	Standard Titration
Measurement Input:	Analog CH1
Stirrer Configuration	
Titration pump:	Pump 1
Reagent Addition 1:	Disabled
Reagent Addition 2:	Disabled
Dosing Type:	Dynamic
End Point Mode:	pH 1EQ point,1st Der
Recognition Options	
Pre-Titration Volume:	5.000 mL
Pre-Titration Stir Time:	60 sec
Measurement Mode:	Signal Stability

Select	Escape	Share Method	Page Up	Page Down
--------	--------	--------------	---------	-----------

- To exit the **View/Modify Method** screen, press the Escape key.
- Next, highlight *Save Method* or *Exit Without Saving Method*.

Saving Method

Select a menu option.

Save Method
Exit Without Saving Method

"Escape" - exits without saving method.

Select	Escape			
--------	--------	--	--	--

- Use Select to save modifications.
- Use Escape to discard the changes.

2.4.5. METHOD OPTIONS

Note: Not all method options can be changed for standard methods.

2.4.5.1. Name

Option: Up to 24 characters

Method Name				
Select the highlighted letter by using the arrow keys then press "Enter". Select the empty field for a space. Press Accept to save the entire name.				
<pre> A B C D E F G H I J K L M N O P Q R S T U V W X Y Z a b c d e f g h i j k l m n o p q r s t u v w x y z À Á Â Ã Ä Å Ç È É I Î Ñ ò ó ô õ ö ù ú û ü ß à á â ã ä å ç è é ê ë ì í ñ ò ó ô õ ö ù ú û ü ÿ i * \ = \$ ' ^ # : 0 1 2 3 4 5 6 7 8 9 % & ' () ? ! () [] < > = / + - </pre>				
copy of 0.1N Sodium Hydr				
Accept	Escape	Delete Letter	Cursor Left	Cursor Right

2.4.5.2. Method Revision

Option: Up to 3 characters

Method Revision				
Select the highlighted letter by using the arrow keys then press <Enter>. Select the empty field for a space. The revision string format is "X.X".				
<pre> B C D E F G H I J K L M N O P Q R S T U V W X Y Z a b c d e f g h i j k l m n o p q r s t u v w x y z À Á Â Ã Ä Å Ç È É I Î Ñ Ó ò ó ô õ ö ù ú û ü ß à á â ã ä å ç è é ê ë ì í ñ ò ó ô õ ö ù ú û ü ÿ i * \ = \$ ' ^ # : 0 1 2 3 4 5 6 7 8 9 % & ' () ? ! () [] < > = / + - * / \ _ ^ ` ' : </pre>				
1.0				
Accept	Escape	Delete Letter	Cursor Left	Cursor Right

2.4.5.3. Analysis Type

Option: Standard Titration, Back Titration, Direct Reading

Analysis Type				
Select the analysis type.				
<pre> Standard Titration Back Titration Direct Reading </pre>				
Select	Escape			

Standard Titration

- A titration with a pH or mV equivalence point detection (single or multiple equivalence points).
- A titration with fixed pH or mV endpoint.
- A titrant standardization.

Back Titration

A titration with a pH or mV equivalence point detection consisting of two titration phases:

- Phase 1
The sample is consumed by a known volume and concentration of titrant 1.
A sufficient amount of titrant 1 is dispensed to surpass the equivalence point in order to react quickly with the sample.
- Phase 2
The excess of titrant 1 is titrated with the titrant 2 to the equivalence point.
The concentration of the sample is determined by the amount of titrant used in phase 2.

Break At Titrant Changing

Option: Yes or No

Select "Yes" to stop the titration temporarily between the titration phases.

This allows users to perform a task related to the analysis (e. g. boiling the sample to remove carbon dioxide, pH adjustment).

Break at Titrant Changing						
Select the option.						
<table border="1"> <tr> <td>NO</td> </tr> <tr> <td>YES</td> </tr> </table>					NO	YES
NO						
YES						
"NO" - without break at titrant changing. "YES" - with break at titrant changing.						
Select	Escape					

Direct Reading

A direct pH, mV or ISE reading with an optional reagent addition.

The titrator will take the measurement automatically once a stable reading has been obtained.

2.4.5.4. Measurement Input

Option: Analog CH1, Analog CH2, Digital CH3, Digital CH4

View/Modify Method																																
Id: USER0003 Modified: 09:40 Mar 23, 2026																																
Select the option to be modified.																																
<table border="1"> <tr> <td>Name:</td> <td>copy of 0.1N Sodium Hydr</td> </tr> <tr> <td>Method Revision:</td> <td>1.0</td> </tr> <tr> <td>Analysis Type:</td> <td>Standard Titration</td> </tr> <tr> <td>Measurement Input:</td> <td>Analog CH1</td> </tr> <tr> <td>Stirrer Configuration</td> <td></td> </tr> <tr> <td> Titrant 1 pump:</td> <td>Analog CH1</td> </tr> <tr> <td> Titrant 2 pump:</td> <td>Analog CH2</td> </tr> <tr> <td> Reagent Addition 1:</td> <td>Digital CH3</td> </tr> <tr> <td> Reagent Addition 2:</td> <td>Digital CH4</td> </tr> <tr> <td>Dosing Type:</td> <td></td> </tr> <tr> <td>End Point Mode:</td> <td>pH 1EQ point,1st Der</td> </tr> <tr> <td>Recognition Options</td> <td></td> </tr> <tr> <td> Pre-Titration Volume:</td> <td>5.000 mL</td> </tr> <tr> <td> Pre-Titration Stir Time:</td> <td>60 sec</td> </tr> </table>					Name:	copy of 0.1N Sodium Hydr	Method Revision:	1.0	Analysis Type:	Standard Titration	Measurement Input:	Analog CH1	Stirrer Configuration		Titrant 1 pump:	Analog CH1	Titrant 2 pump:	Analog CH2	Reagent Addition 1:	Digital CH3	Reagent Addition 2:	Digital CH4	Dosing Type:		End Point Mode:	pH 1EQ point,1st Der	Recognition Options		Pre-Titration Volume:	5.000 mL	Pre-Titration Stir Time:	60 sec
Name:	copy of 0.1N Sodium Hydr																															
Method Revision:	1.0																															
Analysis Type:	Standard Titration																															
Measurement Input:	Analog CH1																															
Stirrer Configuration																																
Titrant 1 pump:	Analog CH1																															
Titrant 2 pump:	Analog CH2																															
Reagent Addition 1:	Digital CH3																															
Reagent Addition 2:	Digital CH4																															
Dosing Type:																																
End Point Mode:	pH 1EQ point,1st Der																															
Recognition Options																																
Pre-Titration Volume:	5.000 mL																															
Pre-Titration Stir Time:	60 sec																															
Select	Escape	Share Method	Page Up	Page Down																												

2.4.5.5. Stirrer Configuration

Use the arrow keys to select the menu option.

Stirrer Configuration				
Select a menu option.				
Stirrer:		Stirrer 1		
Stirring Speed:		1400 RPM		
Select	Escape			

Stirrer

Option: Stirrer 1, Stirrer 2 (if available), Disabled

Stirrer Configuration				
Select a menu option.				
Stirrer:		Stirrer 1		
Stirring Speed:		1400 RPM		
Select	Escape			

Disabled
Stirrer 1
Stirrer 2

Stirrer Speed

Option: 200 to 2500 RPM

The stirrer will remain on for as long as the method is active.

When the stirrer is running, use the \triangle and ∇ keys to adjust the speed.

Stirring Speed				
Enter the speed of the stirrer within below range.				
1400 RPM				
The range is from 200 to 2500 RPM.				
Accept	Escape	Delete Digit		

2.4.5.6. Pump Configuration

Option: Pump 1, Pump 2 (if installed)

Note: For back titrations, the pumps for titrant 1 and titrant 2 need to be selected.

View/Modify Method				
Id: USER0003 Modified: 09:40 Mar 23, 2026				
Select the option to be modified.				
Name: copy of 0.1N Sodium Hydr				
Method Revision: 1.0				
Analysis Type: Standard Titration				
Measurement Input: Analog CH1				
Stirrer Configuration				
Titrant 1 pump: Pump 1				
Titrant 2 pump: Pump 1				
Reagent Addition 1: Pump 2				
Reagent Addition 2: Pump 2				
Dosing Type:				
End Point Mode: pH 1EQ point,1st Der				
Recognition Options				
Pre-Titration Volume: 5.000 mL				
Pre-Titration Stir Time: 60 sec				
Select	Escape	Share Method	Page Up	Page Down

2.4.5.7. Reagent Addition

Option: Burette, Peristaltic Pump, Disabled

Reagent Addition 1				
Select the option to be modified.				
Reagent Pump: Disabled				
Disabled				
Burette 1				
Peristaltic 1				
Burette 2				
Peristaltic 2				
Select	Escape			

Burette (Volume Addition)

Use the numeric keypad to enter the volume to be dispensed.

Addition Volume				
Enter the addition reagent volume to be added to the sample.				
5.000 mL				
Press Help to view the valid ranges for the addition volume.				
Accept	Escape	Delete Digit		

The volume dispensed must be between the limits shown here:

5 mL burette 0.001 to 4.750 mL

10 mL burette 0.001 to 9.500 mL

25 mL burette 0.005 to 23.750 mL

50 mL burette 0.005 to 47.500 mL

Peristaltic Pump (Time Addition)

Option: 0 to 1800 seconds

Enter the dispensing time required to add the desired amount of reagent.

Note: The user should determine this value experimentally. The approximate dispensing rate is 200 mL per minute.

Dispensing Time				
Enter the period of time for running auxiliary pump.				
<div style="background-color: black; color: white; display: inline-block; padding: 2px 10px;">5</div> sec				
Low limit: 0 second High limit: 1800 seconds				
Accept	Escape	Delete Digit		

Stirring Time

Option: 0 to 1800 seconds

The timer will start after the reagent has been added.

Stirring Time				
Please enter the stirring time in seconds.				
<div style="background-color: black; color: white; display: inline-block; padding: 2px 10px;">2</div> sec				
Low limit: 0 second High limit: 1800 seconds				
Accept	Escape	Delete Digit		

Waiting Time

Option: 0 to 1800 seconds

The timer will start after the stirring timer.

Wait Time				
Please enter the wait time in seconds.				
<div style="background-color: black; color: white; display: inline-block; padding: 2px 10px;">2</div> sec				
Low limit: 0 second High limit: 1800 seconds				
Accept	Escape	Delete Digit		

2.4.5.8. Measurement Parameter (Direct Reading Only)

Option: pH, ISE (with analogic probes only), mV

Select the measurement parameter for the direct reading.

Setup screen for the selected parameter is visible in the method options.

View/Modify Method																										
Id: USER0002 Modified: 16:11 Mar 26, 2026																										
Select the option to be modified.																										
<table border="1" style="width: 100%;"> <tr> <td>Name:</td> <td>copy of 0.1N Sodium Hydr</td> </tr> <tr> <td>Method Revision:</td> <td>1.0</td> </tr> <tr> <td>Analysis Type:</td> <td>Direct Reading</td> </tr> <tr> <td>Measurement Input:</td> <td>Analogic pH</td> </tr> <tr> <td>Stirrer Configuration:</td> <td>mV</td> </tr> <tr> <td>Reagent Addition 1:</td> <td>Dispense ISE</td> </tr> <tr> <td>Reagent Addition 2:</td> <td>Dispense</td> </tr> <tr> <td>Measurement Parameter:</td> <td>pH</td> </tr> <tr> <td>Electrode Type:</td> <td>pH</td> </tr> <tr> <td>Linked To:</td> <td>No Link</td> </tr> <tr> <td colspan="2">pH Setup</td> </tr> </table>					Name:	copy of 0.1N Sodium Hydr	Method Revision:	1.0	Analysis Type:	Direct Reading	Measurement Input:	Analogic pH	Stirrer Configuration:	mV	Reagent Addition 1:	Dispense ISE	Reagent Addition 2:	Dispense	Measurement Parameter:	pH	Electrode Type:	pH	Linked To:	No Link	pH Setup	
Name:	copy of 0.1N Sodium Hydr																									
Method Revision:	1.0																									
Analysis Type:	Direct Reading																									
Measurement Input:	Analogic pH																									
Stirrer Configuration:	mV																									
Reagent Addition 1:	Dispense ISE																									
Reagent Addition 2:	Dispense																									
Measurement Parameter:	pH																									
Electrode Type:	pH																									
Linked To:	No Link																									
pH Setup																										
Select	Escape	Share Method	Page Up	Page Down																						

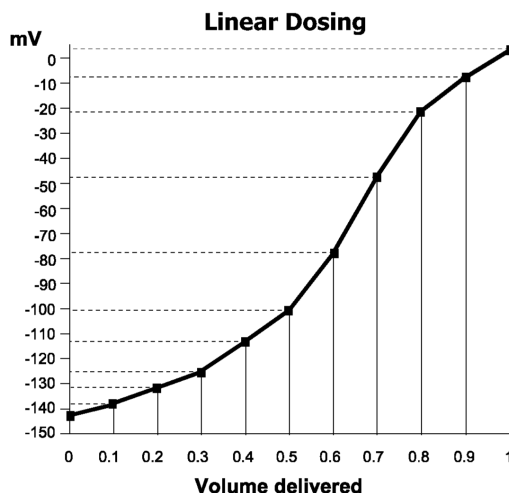
2.4.5.9. Dosing Type

Option: Linear Dosing or Dynamic Dosing

Dosing Type						
Select the dosing type.						
<table border="1" style="width: 100%;"> <tr> <td>Linear Dosing</td> </tr> <tr> <td style="background-color: black; color: white;">Dynamic Dosing</td> </tr> </table>					Linear Dosing	Dynamic Dosing
Linear Dosing						
Dynamic Dosing						
Select	Escape					

Linear Dosing

Linear dosing dispenses a pre-defined volume of titrant with every addition.



Linear dosing is recommended for titrations with a slower reaction rate, difficult nonaqueous titrations, and specific applications.

Note: For steep and normal titration curves, smaller volume increments are recommended, to obtain many points around the equivalence point.

For flat titration curves, larger volume increments are recommended for equivalence point detection.

To set the dosing volume, select Linear Dosing and enter the optimum dose.

Dosing volume ranges are given here.

5 mL burette 0.001 to 4.750 mL

10 mL burette 0.001 to 9.500 mL

25 mL burette 0.005 to 23.750 mL

50 mL burette 0.005 to 47.500 mL

Dynamic Dosing

The titrator determines the titrant dose by trying to maintain a certain potential change (ΔE) with each addition.

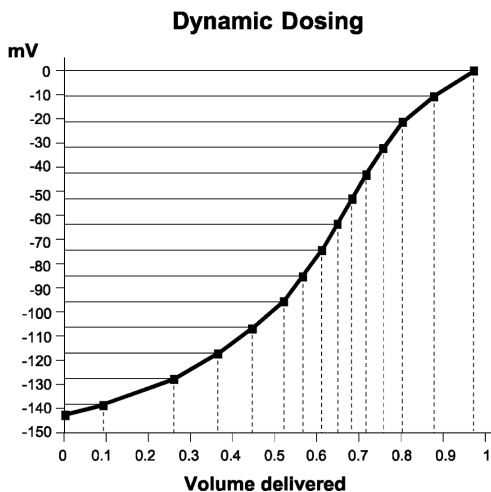
After a titrant dose, if the potential change is lower than the set ΔE , the next dose will be progressively increased until *max Vol* is attained. If the potential change is still lower than the set value, the titration will continue with *max Vol* doses.

After a titrant dose, if the potential change is higher than the set ΔE , the next dose will be progressively decreased until *min Vol* is attained. If the potential change is still higher than the set value, the titration will continue with *min Vol* doses.

The titrant is added in volumes that depend on the proximity of the endpoint as shown in the graph below.

Dynamic dosing allows for larger doses far from the endpoint, reducing the total titration time.

Closer to the endpoint, smaller doses are made, providing more data and improved accuracy.



Dynamic Dosing

Enter min Vol, max Vol and delta E.

0.030 mL - min Vol

0.500 mL - max Vol

4.500 mV - delta E

Press Next to move to the next entry.

Accept	Escape	Delete Digit	Next
--------	--------	--------------	------

The following parameters must be set:

- min Vol** The smallest dose to be dispensed during a titration.
 The *min Vol* must be greater than or equal:
 5 mL and 10 mL burette 0.001 mL
 25 mL and 50 mL burette 0.005 mL
- max Vol** The largest dose to be dispensed during a titration.
 The *max Vol* must be less than or equal to 4.000 mL.
- delta E** Sets the fixed potential jump that has to be achieved after each titrant dose.
 The allowed range is between 0.1 and 99.999 mV.

Recommendations for dosing parameters

For steep and normal titration curves, the recommended settings are:

- delta E** 3.5 to 9 mV
min Vol 0.010 to 0.025 mL (25 mL burette)
max Vol 0.075 to 0.250 mL (25 mL burette)

For flat titration curves, the recommended settings are:

- delta E** 10 to 15 mV
min Vol 0.050 to 0.150 mL (25 mL burette)
max Vol 0.400 to 0.600 mL (25 mL burette)

To achieve the highest levels of accuracy and reproducibility, it is recommended that 20 to 80% of the nominal burette volume used for each titration is consumed. If lower volumes of titrant are required, a smaller burette can be used.

2.4.5.10. Endpoint Mode

Option: Equivalence Endpoint (pH or mV) or Fixed Endpoint (pH or mV)

Titration End Point Mode

Select the end point detection.

Equivalence End Point (pH)

Equivalence End Point (mV)

Fixed End Point (pH)

Fixed End Point (mV)

Select	Escape			
--------	--------	--	--	--

Fixed Endpoint (pH or mV)

Fixed Endpoint (pH)

Option: -2.000 to 20.000 pH

The titration is terminated when the preset pH value has been exceeded.

The endpoint volume is a calculated value based on the dispensed volume when pH is under the preset value and the dispensed volume when pH exceeds the preset value.

Preset pH End Point

Enter the end point pH value.

8.600 pH

The range is from -2.000 to 20.000 pH.

Accept	Escape	Delete Digit	
--------	--------	-----------------	--

Fixed Endpoint (mV)

Option: -2000.0 to 2000.0 mV

The endpoint detection algorithm is the same as for pH, but the threshold value is expressed in mV.

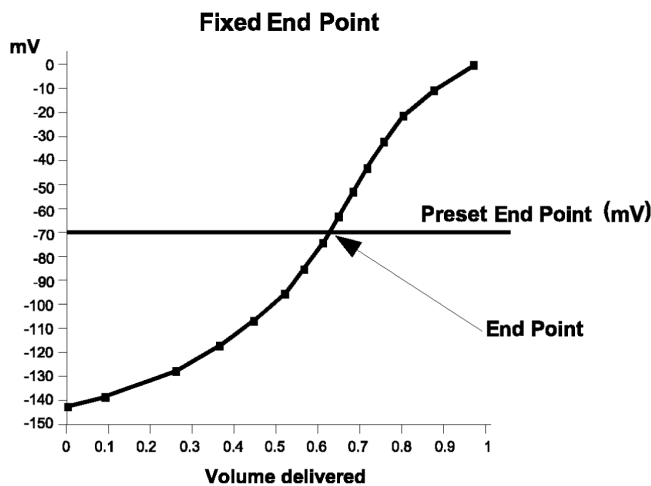
Preset mV End Point

Enter the end point mV value.

0.0 mV

The range is from -2000.0 to 2000.0 mV.

Accept	Escape	Delete Digit	
--------	--------	-----------------	--



Equivalence Endpoint (pH or mV)

The titration is terminated when the equivalence point is detected (the point where the added quantity of titrant equals the quantity of analyte present in the sample).

Number of Equivalence Points

Option: 1 to 5

Number of Equivalence Points				
Enter the number of equivalence points to be found.				
3 points				
The range is between 1 and 5 equivalence points.				
Accept	Escape	Delete Digit		

Endpoint Determination

Option: 1st derivative or 2nd derivative

End Point Determination				
Select the end point determination.				
1st derivative				
2nd derivative				
Select	Escape			

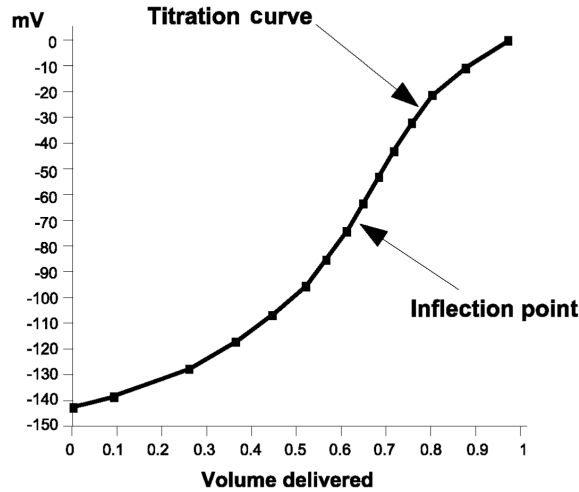
The equivalence point detection algorithm requires three additional titrant doses to be dispensed after the equivalence point is reached.

The reported endpoint volume is a calculated value based on a number of points around the equivalence point.

The potentiometric titration curve is the response in mV potential or pH, between the indication of the electrode versus the volume of titrant added.

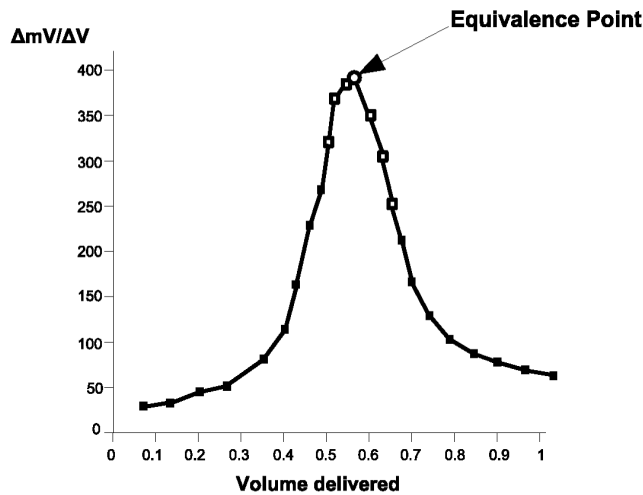
The inflection point of the titration curve is assumed to be the equivalence point of the chemical reaction.

For non-symmetric titration curves, the theoretical error can be reduced by using the dynamic dosing.



1st Derivative

When 1st derivative is used to recognize the equivalence point, the titration curve inflection point (EQP) is the point where the 1st derivative reaches its maximum value.

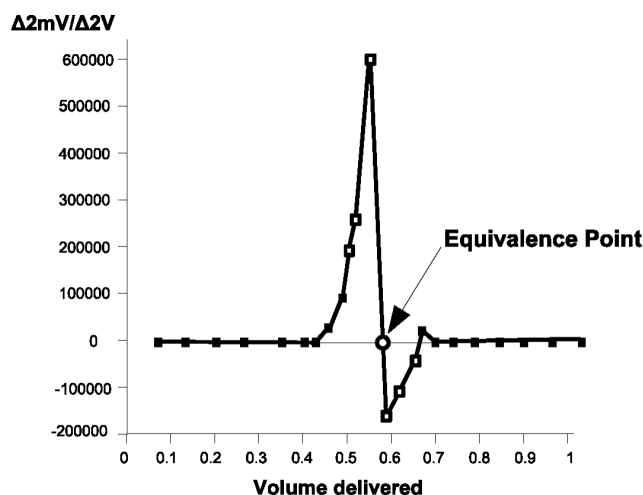


The detection algorithm looks for the maximum value of the 1st derivative. The 1st derivative must be greater than the threshold value at the maximum point.

See [2.4.5.11. Recognition Options \(Equivalence Endpoint Only\)](#) section for more information.

2nd Derivative

When 2nd derivative is used to recognize the equivalence point, the titration curve inflection point (EQP) is the point where the second derivative crosses zero.



The detection algorithm looks for the point where the second derivative changes sign.

The checked point, or 1st derivative, must be greater than the threshold value.

See [2.4.5.11. Recognition Options \(Equivalence Endpoint Only\)](#) section for more information.

2.4.5.11. Recognition Options (Equivalence Endpoint Only)

The **Recognition Options** screen is a set of parameters used to avoid false detection of the equivalence point due to the chemical system (titrant/sample species and concentrations) and/or electrode response.

Recognition Options																			
Select the options for equivalence point recognition.																			
<table border="1"> <tr> <td>Threshold</td> <td>500</td> <td>mV/mL</td> <td></td> <td></td> </tr> <tr> <td>Range</td> <td></td> <td></td> <td>NO</td> <td></td> </tr> <tr> <td>Filtered Derivatives</td> <td></td> <td></td> <td>NO</td> <td></td> </tr> </table>					Threshold	500	mV/mL			Range			NO		Filtered Derivatives			NO	
Threshold	500	mV/mL																	
Range			NO																
Filtered Derivatives			NO																
Select	Escape																		

Threshold

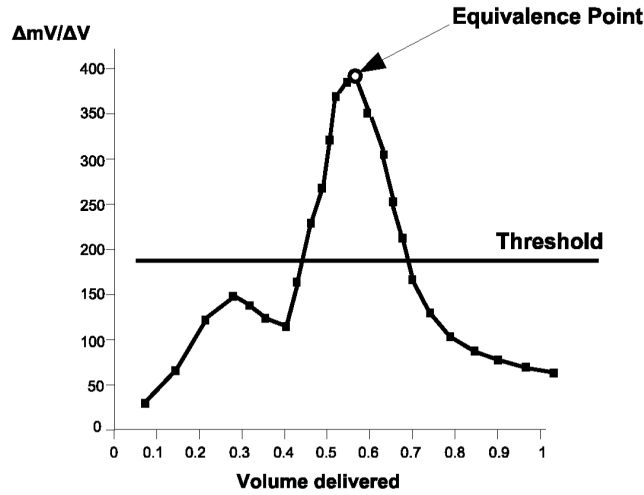
Option: 1 to 9999 mV / mL

This parameter must be set by the user according to the analysis.

The threshold represents the absolute value of the 1st derivative, expressed in mV / mL, which the detection algorithm does not search for the equivalence point.

Threshold				
Enter the threshold for equivalence point detection.				
EQ 1 Threshold: <input type="text" value="500"/> mV/mL				
Recommended value is between: 1 and 450 mV/mL for FLAT Curve, 450 and 1800 mV/mL for NORMAL Curve, 1800 and 9999 mV/mL for STEEP Curve.				
ACCEPT	Escape	Delete Digit		Next Threshold

The recommended value is 40% of the absolute value of the 1st derivative.



Depending on the titration curve profile, the following guide can be used:

- Flat** 1 to 450
- Normal** 50 to 1800
- Steep** 1800 to 9999

Range

Option: -2.000 to 20.000 pH or -2000.0 to 2000.0 mV

Range is an optional feature for equivalence point recognition.

Select "Yes" in the Range Options screen to enable.

The titrator will only look for an equivalence point between the set values.

Range Limits

Enter Limit 1 and Limit 2 for range.

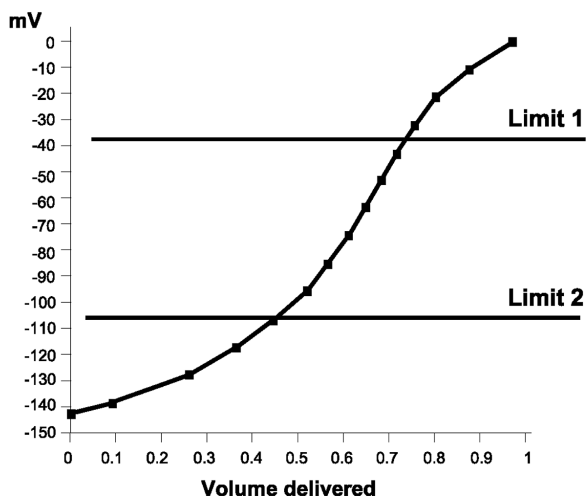
-2000 mV - EQ 1 Limit1

2000 mV - EQ 1 Limit2

Press <Next EQ Range> for the next range.

Accept	Escape	Delete Digit	Next Limit	Next EQ Range
--------	--------	-----------------	---------------	------------------

The Limit 2 value must not be equal to the Limit 1 value.



Filtered Derivatives

Option: Yes or No

This option adds a filtering procedure in the 1st and 2nd derivative computation algorithm that reduces the influence of pH or mV noise.

Select "Yes" in the Filtered Derivative Option to enable.

Filtered Derivatives Option

Select option for filtered derivatives.

NO

YES

"NO" - without filtered derivatives.
"YES" - with filtered derivatives.

Select	Escape			
--------	--------	--	--	--

Noise can be due to:

- Chemical system properties (sample, titrant, solvent), such as slow chemical reactions or unbuffered samples such as wastewater, tap water, wine
- Electrode response
- Incorrect method parameters settings such as *Signal Stability*, *Stirring Speed*
- Insufficient titrant additions

Note: A shift in the endpoint volume by one or two doses may be seen due to filtering.

2.4.5.12. Pre-Titration Volume

During a titration, the equivalence point is reached after many titrant doses. These doses take up extra time while having no relevance for equivalence point detection.

Pre-titration volume adds a large initial dose to jump directly to the proximity of the equivalence point.

This first dose occurs after the pre-titration stir time is completed.

The ranges for pre-titration volumes are shown here.

5 mL burette 0.001 to 4.750 mL

10 mL burette 0.001 to 9.500 mL

25 mL burette 0.005 to 23.750 mL

50 mL burette 0.005 to 47.500 mL

Pre-Titration Volume				
Enter the initial titrant volume to be dispensed.				
9.000 mL				
Press Help to view the valid ranges for the pre-titration volume.				
Accept	Escape	Delete Digit		

To disable a pre-titration volume, enter 0.000 mL.

Note: A pre-titration volume is highly recommended whenever possible. Fewer doses will considerably shorten the overall titration duration.

2.4.5.13. Pre-Titration Stir Time

Option: 0 to 180 seconds

When enabled, the sample is mixed for a set period of time before any titrant is added. This allows the sample to become homogeneous.

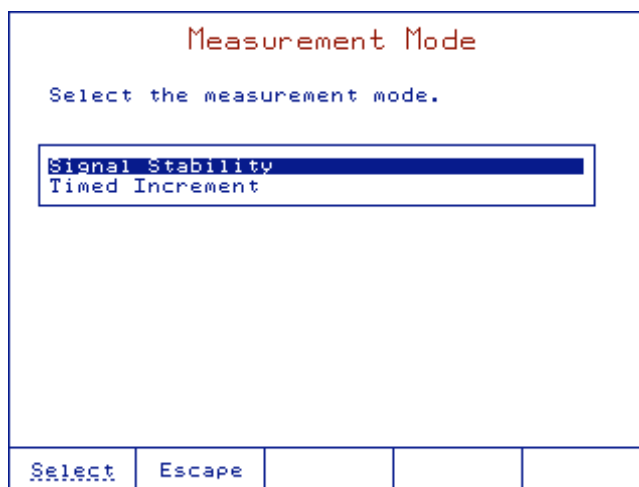
Pre-Titration Stir Time				
Enter the initial mixing time prior to the start of the titration.				
10 seconds				
The range is from 0 to 180 seconds.				
Accept	Escape	Delete Digit		

Pre-titration stir time is disabled if 0 seconds is entered.

2.4.5.14. Measurement Mode

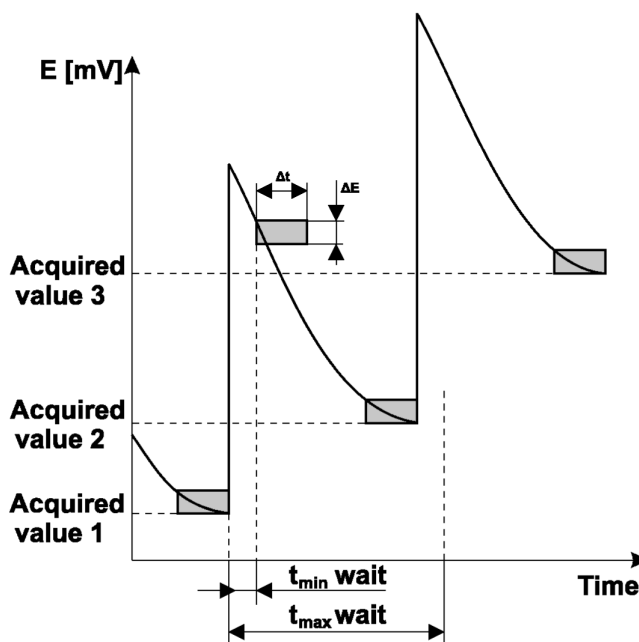
Option: Signal Stability or Timed Increment

During titration the acquisition of the potential (mV) value of the solution can be done by using either *Signal Stability* or *Timed Increment* option.



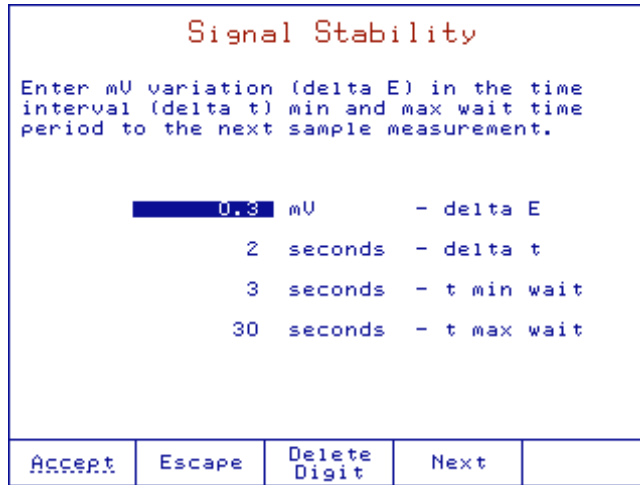
Signal Stability

When *Signal Stability* is selected, the titrator acquires the potential (mV) only when stable conditions are reached. The principles of signal stability are plotted below.



The signal stability window (condition) represents the time interval (Δt) during which the potential measured in solution (mV) is confined inside the potential interval (ΔE).

The new signal value is acquired if the stability condition is reached after the minimum (t_{\min}) wait time.
 If the stability condition is not reached and the maximum (t_{\max}) wait time has elapsed, the potential is acquired.

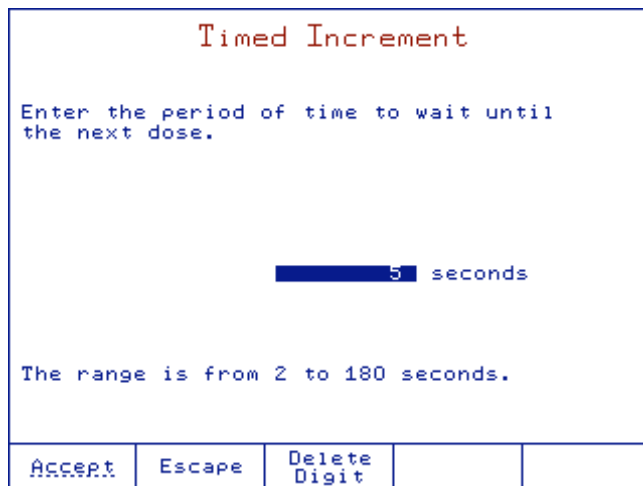
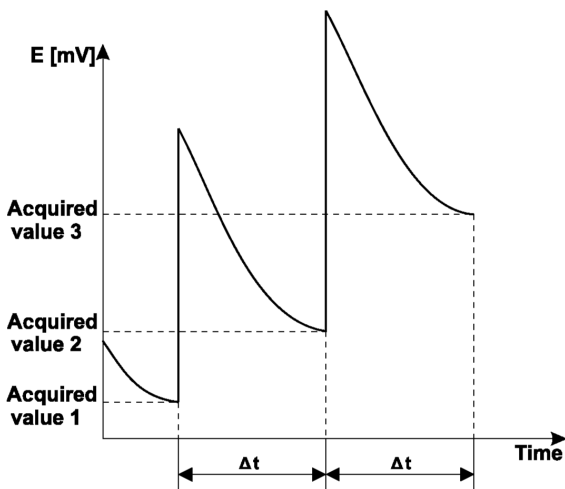


- delta E** Maximum change in potential during *delta t*
 The range is from 0.1 to 99.9 mV.
- delta t** The time interval during which the potential is measured.
 The range is from 1 to 10 seconds.
- t min wait** The minimum elapsed time before a stability check.
 This is also the minimum elapsed time between two doses.
 The range is from 2 seconds to *t max wait* time.
- t max wait** The maximum elapsed time between two successive doses.
 If the *t max wait* has elapsed, a new dose is added even if the signal stability condition is not reached.
 The range is from *t min wait* time to 180 seconds.

Timed Increment

Option: 2 to 180 seconds

When *Timed Increment* is selected, the titrator acquires the potential (mV) at a fixed time interval (no signal stability check).
 The time period between two acquisitions must be set according to the reaction and the response time of the electrode.



2.4.5.15. Electrode Type

Option: Up to 20 characters

Electrode Type

Select the highlighted letter by using the arrow keys then press "Enter".
Select the empty field for a space.
Press Accept to save the electrode type.

■	A	B	C	D	E	F	G	H	I	J	K	L
M	N	O	P	Q	R	S	T	U	V	W	X	Y
Z	a	b	c	d	e	f	g	h	i	j	k	l
m	n	o	p	q	r	s	t	u	v	w	x	y
z	À	Á	Â	Ã	Ä	Å	Æ	Ç	È	É	Ê	Ë
Ì	Í	Î	Ï	Ñ	Ò	Ó	Ô	Õ	Ö	Ù	Ú	Û
Ü	Ý	Þ	ß	à	á	â	ã	ä	å	ö	÷	ø
ù	ú	û	ü	ý	ÿ	ı	•	ˆ	˜	˘	˙	˚
0	1	2	3	4	5	6	7	8	9	.	,	:
?	!	()	[]	<	>	=	/	+	-	

■ PL

Accept	Escape	Delete Letter	Cursor Left	Cursor Right
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2.4.5.16. Blank Option

Option: Disabled, V-Blank, Blank-V

This feature allows the user to select the procedure for the blank calculations (where V is the volume of titrant dispensed during the titration and blank is the volume of titrant consumed by the blank sample).

View/Modify Method

Id: USER0003 Modified: 09:40 Mar 23, 2026

Select the option to be modified.

Analysis Type:	Standard Titration
Measurement Input:	Analog CH1
Stirrer Configuration	
Titration pump:	Pump 1
Reagent Addition 1:	Disabled
Reagent Addition 2:	Disabled
Dosing Type:	Dynamic
End Point Mode:	mV 3EQ points, 1st Der
Recognition Options	
Pre-Titration Volume:	U - Blank
Pre-Titration Stir Time:	Blank - U
Measurement Mode:	Sign: No Blank
Electrode Type:	
Blank Option:	No Blank

Select	Escape	Share Method	Page Up	Page Down
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If one of the options (V-Blank or Blank-V) is selected in the **View / Modify Method** screen, the *blank value* will be active on the **View / Modify Method** screen and the value of the blank can be set (in liters).

Blank Value

Enter the blank volume in liters.

0.00125 L

Accept	Escape	Delete Digit		Exponent
--------	--------	--------------	--	----------

The blank volume must be between 0.000001 and 0.1 L.

2.4.5.17. Calculations

The final result is calculated using the endpoint volume (titrant volume at the equivalence point or at the fixed endpoint), and a formula selected by the user.

Calculations				
Select either the calculation to be performed or modify the variables.				
<div style="border: 1px solid black; padding: 5px;"> Edit Variable Values No Formula (mL only) No Formula (L only) Sample Calc. by Weight Sample Calc. by Volume Stdz. Titrant by Weight Stdz. Titrant by Volume Generic Formula </div>				
Select	Escape			

Standard Titration Calculations

Edit Variable Values

Edit the variables in a previously selected calculation.

For each formula, selected variables can be changed.

No Formula (mL only)

Only the volume of titrant (mL) required to reach the endpoint will be displayed.

No Formula (L only)

Only the volume of titrant (L) required to reach the endpoint is displayed.

Sample Calculations by Weight

Titrant units

Option: M (mol / L), N (eq / L), g / L, mg / L

Final result units

Option: ppt (g / kg), ppm (mg / kg), ppb ($\mu\text{g} / \text{kg}$), % (g / 100 g), mg / g, mg / kg, mol / kg, mmol / g, eq / kg, meq / kg

This calculation is used when the concentration of an analyte is determined by the weight of the sample.

The results are based on the initial sample weight (in grams).

The titrator will calculate the results based on the titrant and sample units selected.

A formula example is shown below using M (mol / L) as the titrant unit and ppt (g / kg) as the final result unit.

Variables can be set according to the amount of sample and titrant used.

Calculating Sample Concentration				
M (mol/L) --> ppt (g/kg)				
The calculation is:				
$\frac{U \times \frac{\text{mol}}{\text{L}} \times \frac{\text{mol}}{\text{mol}} \times \frac{\text{g}}{\text{g}}}{\frac{\text{g}}{\text{g}} \times \frac{\text{kg}}{1000\text{g}}}$				
Select the variables to change value. U = volume dispensed in liters.				
<div style="border: 1px solid black; padding: 5px;"> 1.000 mol/L -> titrant conc. 1.000 mol/mol -> (sample/titrant) 1.000 g/mol -> mw of sample 1.000 g -> sample weight </div>				
Select	Escape	Save / Exit		

Sample Calculations by Volume

Titrant Units

Option: M (mol / L), N (eq / L), g / L, mg / L

Final Result Units

Option: ppt (g / L), ppm (mg / L), ppb (μg / L), M (mol / L), N (eq / L), mg / L, μg / L, mmol / L, mg / mL, mg/100 mL, g / 100 mL, eq / L, meq / L

This calculation is used when the concentration of an analyte is determined in terms of the volume of sample.

The results are based on the initial sample volume (in milliliters).

The titrator will calculate the results based on the selected units.

A formula example is shown below using N (eq / L) as the titrant units and g / L as the final result units.

Variables can be set according to the amount of sample and titrant used.

Calculating Sample Concentration

M (mol/L) --> mol/L

The calculation is:

$$\frac{V \times \frac{\text{mol}}{\text{L}} \times \frac{\text{mol}}{\text{mol}}}{\text{mL} \times \frac{\text{L}}{1000\text{mL}}}$$

Select the variables to change value.
V = volume dispensed in liters.

1.000 mol/L -> titrant conc.

1.000 mol/mol -> (sample/titrant)

100.00 mL -> sample volume

Select	Escape	Save / Exit	
--------	--------	----------------	--

Standardize Titrant by Weight

Option: M (mol / L), N (eq / L), g / L, mg / L

This calculation is used when the concentration of the titrant is determined using a solid standard.

Determination of the titrant concentration is based on the primary standard weight (in grams).

The calculation is based on the selected titrant unit. If the titrant unit is M (mol / L), the formula used to calculate the result is displayed below.

Calculating Titrant Concentration

The titrant concentration unit is M (mol/L).

The calculation is:

$$\frac{g \times \frac{\text{mol}}{\text{g}} \times \frac{\text{mol}}{\text{mol}}}{V}$$

Select the variables to change value.
V = volume dispensed in liters.

0.200 g -> standard weight

204.23 g/mol -> mw of standard

1.000 mol/mol -> (titrant/standard)

Select	Escape	Save / Exit	
--------	--------	----------------	--

Standardize Titrant by Volume

Option: M (mol /L), N (eq / L), g / L, mg / L

This calculation is used when the concentration of the titrant is determined using a primary standard solution.

Determination of the titrant concentration is based on the primary standard volume (in milliliters).

The calculation is based on selected titrant unit. If the titrant unit is N (eq / L), the formula used to calculate the result is displayed below.

Calculating Titrant Concentration

The titrant concentration unit is N (eq/L).

The calculation is:

$$\frac{\text{mL} \times \frac{\text{L}}{1000\text{mL}} \times \frac{\text{eq}}{\text{L}}}{U}$$

Select the variables to change value.
U = volume dispensed in liters.

1.684 mL -> standard volume
2.375 eq/L -> standard conc.

Select	Escape	Save / Exit		
--------	--------	----------------	--	--

Generic Formula

Final results units

Option: ppt (g/kg), ppt (g/L), ppm (mg/kg), ppm (mg/L), ppb ($\mu\text{g/kg}$), ppb ($\mu\text{g/L}$), % (g/100 g), M (mol/L), mg/g, N (eq/L), g/L, mg/kg, mg/L, mol/kg, $\mu\text{g/L}$, mol/L, mmol/g, eq/kg, mmol/L, meq/kg, mg/mL, mg/100 mL, g/100 mL, eq/L, meq/L, no unit

Users can define their own calculation formula based on the final result units in a solid or liquid sample.

The titrator will calculate the results based on the selected unit.

The formula can be either for titrant standardization or sample analysis.

Calculating Sample Concentration

Final unit is mg/L.

The calculation is:

$$\frac{C \times U \times F1 \times F2 \times F3}{S}$$

Select the variables to change value.
U = volume dispensed in liters.

1.000 C -> (titrant conc.)
1.000 F1 -> (general factor)
1.000 F2 -> (general factor)
1.000 F3 -> (general factor)

Select	Escape	Save / Exit		
--------	--------	----------------	--	--

- C the concentration of the titrant
 F1, F2, F3 general factor
 S sample size, in grams or milliliters
 V the volume delivered, in liters, to reach the endpoint

General factors

- Weight conversion** mol/L, eq/L, g/L, mg/L
Reaction ratio mol/mol, mol/eq, eq/mol
Unit conversion L to mL, g to mg
Weight conversion kg, g, mg, μg , mole, mmole

Back Titrations Calculations

Calculations				
Select either the calculation to be performed or modify the variables.				
<div style="border: 1px solid black; padding: 2px;"> Sample Calc. by Weight Sample Calc. by Volume Generic Formula </div>				
Select	Escape			

Sample Calculations by Weight

Select the titrant 1 unit, titrant 2 unit, and final result unit.

Titrant Units

Option: M (mol /L), N (eq / L), g / L, mg / L

Final Result Units

Option: ppt (g/kg), ppm (mg/kg), ppb ($\mu\text{g}/\text{kg}$), % (g/100 g), mg/g, mg/kg, mol/kg, mmol/kg, eq/kg, meq/kg

A formula example is shown below using M (mol / L) as the titrant 1 units, M (mol / L) as the titrant 2 units, mg / g and the final result units. This formula is used to calculate the amount of titrant 1 to dispense.

Calc. Direct Titr. Volume				
Titr1 Unit: M (mol/L)-->Result Unit: L				
The calculation is:				
$\frac{\text{g} \times \frac{\text{mol}}{\text{g}} \times f}{\frac{\text{mol}}{\text{L}} \times \frac{\text{mol}}{\text{mol}}}$				
Select the variables to change value.				
<div style="border: 1px solid black; padding: 2px;"> 1.000 g -> sample weight 1.000 g/mol -> mw of sample 1.000 f ->(excess factor) 1.000 mol/L -> titrant 1 conc. </div>				
Select	Escape		Next	

The formula is based on the assumption that the sample concentration is 100% w / w.

The titrator will calculate the volume of titrant 1 needed to consume the sample and multiply it with the excess factor in order to raise or lower the amount of titrant 1 dispensed.

Variables can be set according to the amount of sample and titrant used.

Press Next to proceed to the next formula.

If you do not want the titrator to calculate the volume of titrant 1 to be added see [2.4.5.23. Titrant 1 Entry \(Back Titration Only\)](#) section for more information.

The remaining volume of titrant 1 will need to be calculated.

The following formula is used to calculate the remaining volume of titrant 1 after the reaction with the sample.

Calc. Excess Volume Of Titr1.

M (mol/L) + M (mol/L) --> L

The calculation is:

$$\frac{U2 \times \frac{\text{mol}}{\text{L}} \times \frac{\text{mol}}{\text{mol}}}{\frac{\text{mol}}{\text{L}}}$$

U1 = U1tot - U1 excess
 Select the variables to change value.
 U2 = backtitr. vol. dispensed in liters.

1.000 mol/L -> titrant 2 conc.

1.000 mol/mol -> (titrant 1/titrant 2)

1.000 mol/L -> titrant 1 conc.

Select	Escape		Next	
--------	--------	--	------	--

When all the variables are set, press Next to proceed with the "Calculating Sample Concentration" formula.

Sample Calculations by Volume

Select the titrant 1 unit, titrant 2 unit, and the final result unit.

Titrant Units

Option: M (mol/L), N (eq/L), g/L, mg/L

Final Result Units

Option: ppt (g/L), ppm (mg/L), ppb ($\mu\text{g/L}$), M (mol/L), N (eq/L), mg/L, $\mu\text{g/L}$, mmol/L, mg/mL, mg/100 mL, g/100 mL, eq/L, meq/L

After you have selected the titrant 1, titrant 2, and the final result units, the titrator will display the formula used to calculate the amount of titrant 1 (used in the first stage of back titration) to be dispensed.

Calc. Direct Titr. Volume

Titr1 Unit: M (mol/L)-->Result Unit: L

The calculation is:

$$\frac{\text{mL} \times \frac{\text{L}}{1000\text{mL}} \times \frac{\text{g}}{\text{L}} \times \frac{\text{mol}}{\text{g}} \times f}{\frac{\text{mol}}{\text{L}} \times \frac{\text{mol}}{\text{mol}}}$$

Select the variables to change value.

1.000 mL -> sample volume

1.000 g/L -> sample max conc.

1.000 g/mol -> mw of sample

1.000 f ->(excess factor)

Select	Escape		Next	
--------	--------	--	------	--

The formula is based on the assumption that the sample concentration is 100% v / v.

The titrator will calculate the volume of titrant 1 needed to consume the sample and multiply it with the excess factor in order to raise or lower the amount of titrant 1 dispensed.

Variables can be set according to the amount of sample and titrant used.

Press Next to proceed to the next formula.

If you do not want the titrator to calculate the volume of titrant 1 to be added see [2.4.5.23. Titrant 1 Entry \(Back Titration Only\)](#) section for more information.

The following formula is used to calculate the remaining volume of titrant 1 after the reaction with the sample.

Calc. Excess Volume Of Titr1.

M (mol/L) + M (mol/L) --> L

The calculation is:

$$\frac{U2 \times \frac{\text{mol}}{\text{L}} \times \frac{\text{mol}}{\text{mol}}}{\frac{\text{mol}}{\text{L}}}$$

U1 = U1tot - U1 excess
 Select the variables to change value.
 U2 = backtitr. vol. dispensed in liters.

1.000 mol/L -> titrant 2 conc.
1.000 mol/mol -> (titrant 1/titrant 2)
1.000 mol/L -> titrant 1 conc.

Select	Escape		Next
--------	--------	--	------

When all the variables are set, press Next to proceed with the "Calculating Sample Concentration" formula.

Calculating Sample Concentration

Final unit is g/L:

The calculation is:

$$\frac{U1 \times \frac{\text{mol}}{\text{L}} \times \frac{\text{mol}}{\text{mol}}}{\frac{\text{mL}}{\text{L}} \times 1000 \text{mL} \times \frac{\text{mol}}{\text{g}}}$$

Select the variables to change value.
 U1 = volume dispensed in liters

1.000 mol/L -> titrant 1 conc.
1.000 mol/mol -> (sample/titrant 1)
1.000 mL -> sample volume
1.000 g/mol -> mw of sample

Select	Escape	Save / Exit	
--------	--------	----------------	--

Generic Formula

Users can define their calculation formula for the "Direct Titration Volume", "Calculating Excess Volume of Titrant 1" and "Final Sample Concentration" in a solid or liquid sample.

2.4.5.18. Dilution Option

Option: Enabled or Disabled

When the initial sample is diluted, a titration is made with an aliquot of the diluted sample, dilution calculations can be used. The calculations are based on the original sample weight or volume in order to express the results for the initial sample.

Dilution Parameters

Select the option.

Final Dilution Volume:	100.000 mL
Aliquot Volume:	10.000 mL
Analyte size to be diluted:	1.0000 mL

Select	Escape		
--------	--------	--	--

- Final Dilution Volume** The volume of the sample after dilution
- Aliquot Volume** Volume of sample taken from the dilution for titration
- Analyte size to be diluted** The initial sample weight or volume

2.4.5.19. Titrant Name

Option: Up to 15 characters

Titrant1 Name

Select the highlighted letter by using the arrow keys then press "Enter".
 Select the empty field for a space.
 Press Accept to save the entered text.

■	A	B	C	D	E	F	G	H	I	J	K	L
M	N	O	P	Q	R	S	T	U	V	W	X	Y
Z	a	b	c	d	e	f	g	h	i	j	k	l
m	n	o	p	q	r	s	t	u	v	w	x	y
z	À	Á	Â	Ã	Ä	Å	Æ	Ç	È	É	Ê	Ë
Ì	Í	Î	Ï	Ð	Ñ	Ò	Ó	Ô	Õ	Ö	×	Ù
Ú	Û	Ü	Ý	Þ	ß	à	á	â	ã	ä	å	æ
ç	è	é	ê	ë	ì	í	î	ï	ð	ñ	ò	ó
ô	õ	ö	÷	¸	¹	º	»	¼	½	¾	¸	º
0	1	2	3	4	5	6	7	8	9	×	.	,
?	!	()	[]	<	>	=	/	+	-	

0.1N NaOH

Accept	Escape	Delete Letter	Cursor Left	Cursor Right
--------	--------	---------------	-------------	--------------

Note: For back titrations the name of titrant 1 and titrant 2 can be entered.

2.4.5.20. Titrant Concentration

Enter the concentration of the titrant to be used.

When determining the titrant concentration, only the concentration unit is displayed.

Note: For back titrations the concentration of titrant 1 and titrant 2 can be entered.

Titrant1 Concentration

Enter the titrant 1 concentration.

0.10676 M (mol/L)

Accept	Escape	Delete Digit		Exponent
--------	--------	--------------	--	----------

2.4.5.21. Analyte Size

Option: 0.001 to 250.0

Enter the size of the sample (for sample-concentration determination) or standard (for titrant-concentration determination).

Sample Volume				
Enter the initial sample volume in milliliters.				
1.0000 mL				
This volume will be used when fixed sample size is selected.				
ACCEPT	Escape	Delete Digit		Exponent

Note: For back titrations the concentration of titrant 1 and titrant 2 can be entered.

2.4.5.22. Analyte Entry

Option: Fixed, Manual, Same as Previous (linked methods only)

Analyte Entry							
Select the entry mode of analyte.							
<table border="1"> <tr> <td>Fixed Weight or Volume</td> </tr> <tr> <td>Manual Weight or Volume</td> </tr> <tr> <td>Same as Previous</td> </tr> </table>					Fixed Weight or Volume	Manual Weight or Volume	Same as Previous
Fixed Weight or Volume							
Manual Weight or Volume							
Same as Previous							
Verify the correct formula is being used, i.e. weight or volume analyte type.							
Select	Escape						

Fixed Weight or Volume

For each titration will use a set weight or volume in the calculations.

Manual Weight or Volume

For each titration the exact weight or volume can be entered at the beginning of each titration.

Same as Previous (Linked Method)

The same weight or volume is used for both methods.

2.4.5.23. Titrant 1 Entry (Back Titration Only)

Select the mode for evaluating the necessary quantity of titrant 1 used in the back titration process (phase 1).

Titrant 1 Entry						
Select the entry mode of titrant 1.						
<table border="1"> <tr> <td>Calculated By Formula</td> </tr> <tr> <td>Fixed By User</td> </tr> </table>					Calculated By Formula	Fixed By User
Calculated By Formula						
Fixed By User						
Select	Escape					

Calculated by Formula The volume of titrant 1 to be dispensed in the phase 1 of back titration will be calculated by the titrator. See [Back Titrations Calculations](#) section for more information.

Fixed by User A fixed volume of titrant 1 will be used during the first phase of back titration process (direct titration). The volume must be between 0.1 and 100 mL.

2.4.5.24. Maximum Titrant Volume

Option: 0.100 to 100.000 mL

The maximum titrant volume used in the titration must be set according to the analysis.

If the titration endpoint (fixed or equivalence point) is not reached, the titration will be terminated after the maximum titrant volume has been dispensed. The error message "Limits Exceeded" will appear on the display.

Maximum Titrant Volume						
Enter the maximum titrant volume to be dispensed.						
<table border="1"> <tr> <td>15.000</td> <td>mL</td> </tr> </table>					15.000	mL
15.000	mL					
Recommend the total volume of the burette.						
Accept	Escape	Delete Digit				

2.4.5.25. Potential Range

Option: -2000.0 to 2000.0 mV

The input potential range can be set by the user. The titration will be terminated and an error message will appear if the potential is outside these limits.

These limits provide protection against a titration that does not generate an endpoint due to potential over-range.

Potential Range				
Enter the upper and lower potential.				
<div style="border: 1px solid black; display: inline-block; padding: 2px;">2000.0</div> mV - Upper Limit -2000.0 mV - Lower Limit				
Press Next to move to the next entry.				
ACCEPT	Escape	Delete Digit	Next	

2.4.5.26. Volume / Flow Rate

The flow rate for the dosing system can be set by the user in an interval of 0.1/0.3 to 2 times the burette volume.

5 mL burette 0.1 to 10 mL/minute

10 mL burette 0.3 to 20 mL/minute

25 mL burette 0.3 to 50 mL/minute

50 mL burette 0.3 to 100 mL/minute

The flow rate is set for all burette operations.

Flow Rate				
Enter the titrant/reagent flow rate.				
<div style="border: 1px solid black; display: inline-block; padding: 2px;">50.0</div> mL/min				
The range is from 0.3 to twice the total volume of the burette.				
ACCEPT	Escape	Delete Digit		

Note: The titrator will automatically detect the burette size and display the correct high limit volume.

2.4.5.27. Signal Averaging

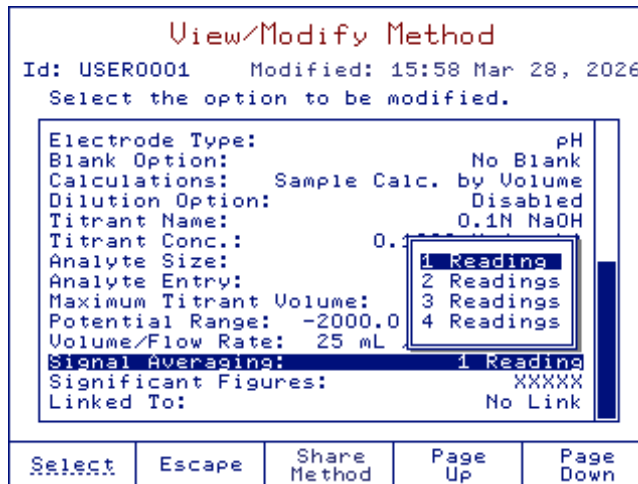
Option: 1, 2, 3, 4 readings

This option enables filtering on the mV / pH reading.

If 1 Reading is selected, the filtering is disabled. The titrator will take the last reading and place it into a "moving window" along with the last 2, 3, or 4 readings (depending on the selected option).

The average of those readings is displayed and used for calculations.

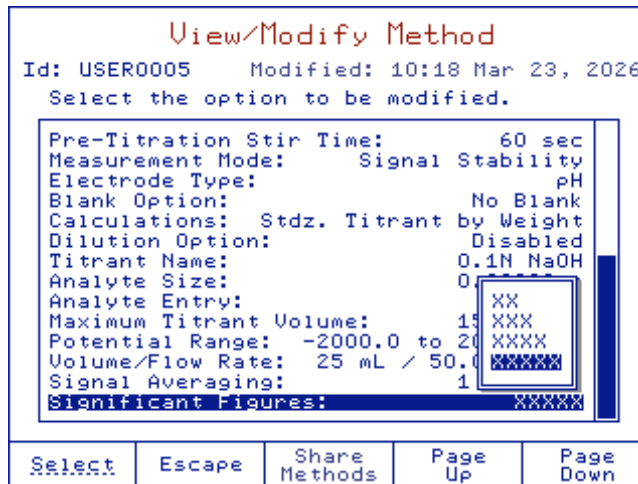
Averaging more readings is helpful when a noisy signal is received from the electrode.



2.4.5.28. Significant Figures

Option: Two (XX), Three (XXX), Four (XXXX) or Five (XXXXX)

This option allows the user to set the format for displaying the final titration result.

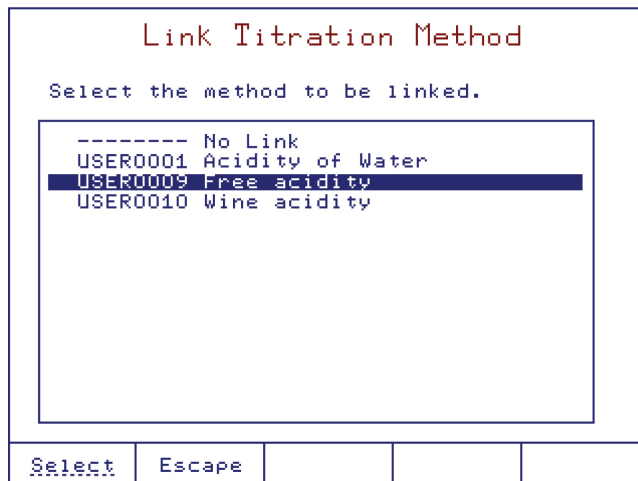


2.4.5.29. Linked Method

This option allows the user to link two titration methods.

If No Link is selected, only the current method will run.

If a method is selected, it will run after the current method.

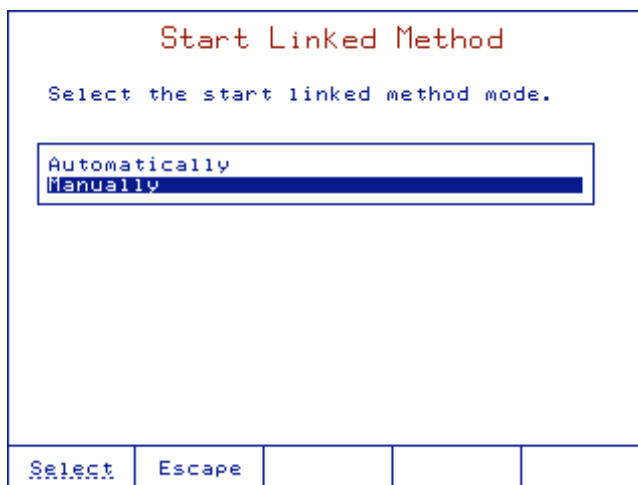


2.4.5.30. Start Linked Method (Linked Method Only)

Option: Manually or Automatically

Selecting Manually will stop the titration temporarily between the methods.

This break allows users to perform a task related to the analysis (e.g.: boiling the sample to remove carbon dioxide).



2.4.6. SHARING METHODS AND REPORTS

Titration methods, method parameters, titration reports, tray reports, and meter information can be printed via USB, shared over Ethernet using FTP/FTPS or SMB; or saved to a USB storage device. Files can be sent as raw text formats (i.e. .txt, .rpt) or converted to PDF or signed PDF format (if a valid signature file has been imported).

Titration reports can be compiled into a CSV summary file that can be transferred over FTP/FTPS or SMB.

Option: Print

Add to Summary

Share Summary over FTP

Share Summary over SMB

As Raw over FTP

As Raw over SMB

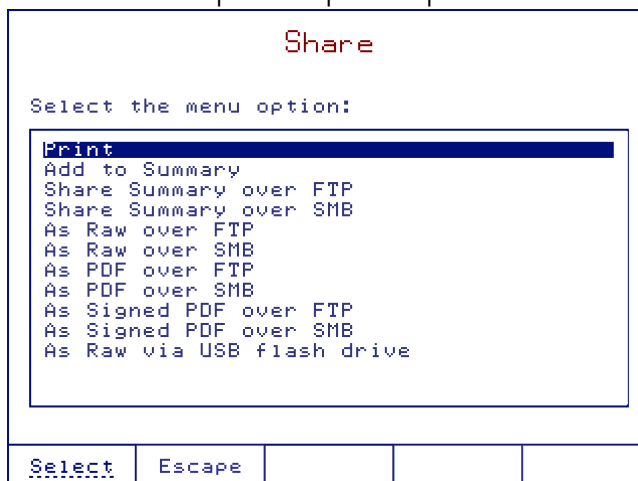
As PDF over FTP/As Signed PDF over FTP

As PDF over SMB/As Signed PDF over SMB

As Raw via USB flash drive

2.4.6.1. Printing

- To print method parameters, press Method Options from the main screen.
- Press Share Method.
- Press the ▲ and ▼ keys to highlight *Print* option.
- Press Select and wait a few seconds until the printer completes the job.



If no printer is connected or if the printer is offline, an error message is displayed.






See [2.10.2. HI932933 Network Box](#) section for information about printer connection methods.

See [2.3.13. Network Box Setup](#) » [Printer Setup](#) section for printer configuration.

2.4.6.2. Add to Summary

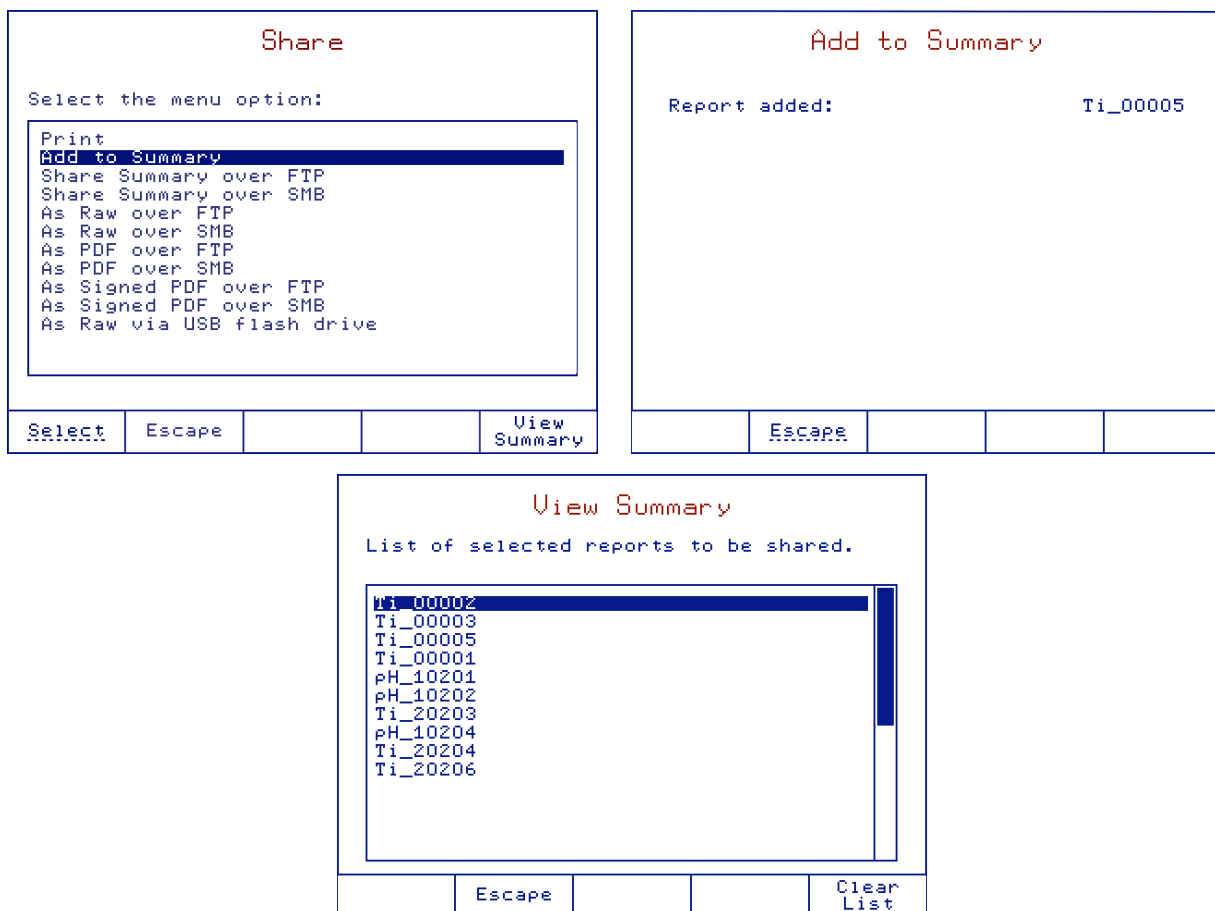
This option allows the user to consolidate data into a single report. Summary transfer is allowed via FTP/FTPS or SMB.

To share the summary over FTP/FTPS or SMB, all reports must be compiled and a Summary created.

- Press  from the main screen.
- Press .
- Press the  and  keys to highlight *Add to Summary* option.
- Press .

The report is added to the list of selected reports to be shared. See View Summary screen.

- To clear the list of items to be shared, press .





2.4.6.3. FTP & SMB Transfer

The titrator supports FTPS and SMB (Server Message Block) file protocols for secure or standard network data transfer, enabling data consolidation and sharing of raw data and (signed) PDF files.

Additionally, direct titrator output can be transferred as raw data format file via USB flash drive.

See [2.3.13. Network Box Setup](#) » [PDF Signature](#) section for importing details.

- Press  from the main screen.
- Press .

2.5. TITRATION MODE

2.5.1. RUNNING A TITRATION

Before beginning a titration, make sure that the below listed conditions are met.

- At least one pump is properly installed.
- A burette is inserted in the pump and filled with titrant.
- The aspiration tube is inserted in the titrant bottle and primed. The dispensing tube is over the titration beaker.
- The tubes are installed on the peristaltic pump and filled with reagent.
- The standard or sample has been carefully weighed / measured into the beaker.
- The electrode(s) and the temperature probe are submersed in the beaker.
- The desired method is selected and the parameters are set to the optimal values.



Starting a Titration

To start a new analysis, press  from the main screen.


When an analysis begins:

- The stirrer, if enabled, will turn on.
See [2.4.5.5. Stirrer Configuration](#) section for more information.
- The quantity of reagent will be added according to the parameters set in the method, if enabled.
See [2.4.5.7. Reagent Addition](#) section for more information.
- The pre-titration volume will be dispensed, if enabled.
See [2.4.5.12. Pre-Titration Volume](#) section for more information.
- After the pre-titration volume is added the pre-titration stir time starts, if enabled.
See [2.4.5.13. Pre-Titration Stir Time](#) section for more information.
- The titrator will start the analysis and continue to deliver titrant until the endpoint is detected or the titration is terminated.

Suspending a Titration

- While a titration or analysis is in progress, press  to temporarily stop the analysis/titration.
This will stop the dosing pump if it is running.
- To continue the titration or analysis, press .

Viewing the Titration Curve

During a titration, the potentiometric curve and the derivative curve (equivalence point only) can be displayed on the **Titration Graph** screen by pressing .

The potentiometric curve and the derivative curve are scaled to fit simultaneously inside the display.

When a titration endpoint is successfully detected, the volume is displayed on the graph and marked with an "x".

The contents of the graph as related to an endpoint type are as follows:

Equivalence endpoint (pH)

- » **Figure 1** displays the pH readings and the selected derivative vs. volume of titrant.

Equivalence endpoint (mV)

- » **Figure 2** displays the the mV readings and the selected derivative vs. volume of titrant.

Fixed endpoint (pH)

- » **Figure 3** displays the pH readings vs. volume of titrant.

Fixed endpoint (mV)

» Figure 4 displays the mV readings vs. volume of titrant.

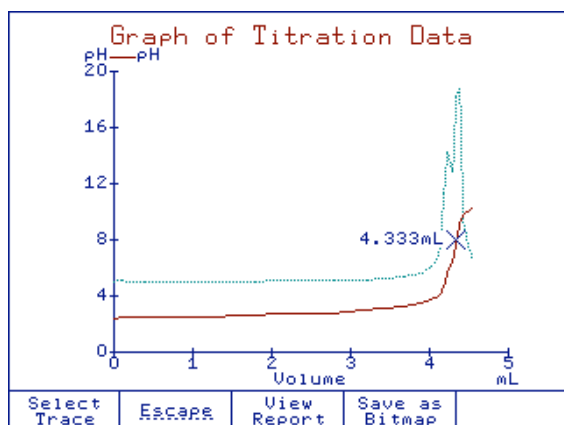


Figure 1. Equivalence endpoint (pH)

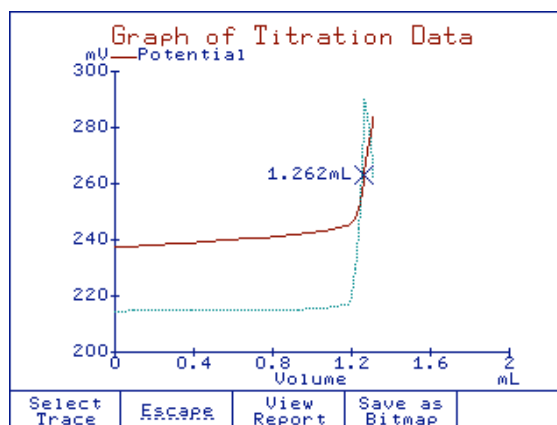


Figure 2. Equivalence endpoint (mV)

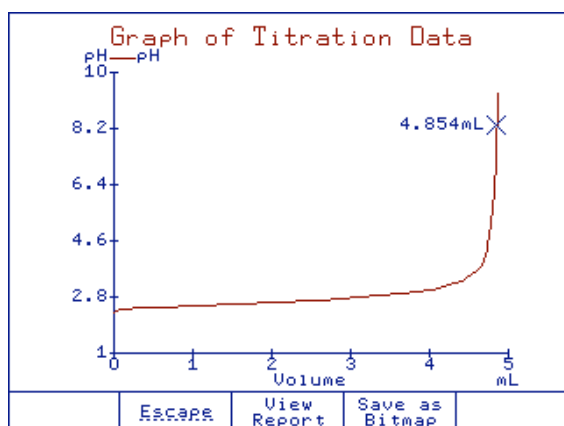


Figure 3. Fixed endpoint (pH)

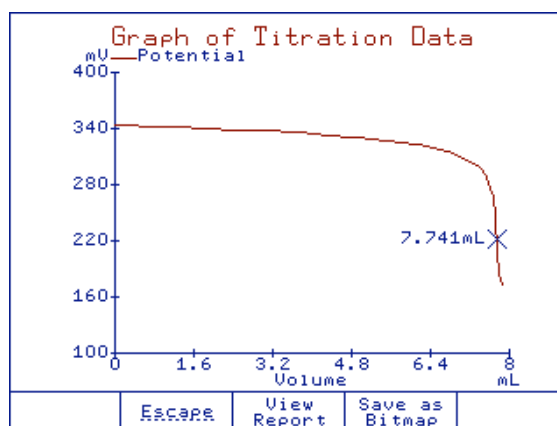




Figure 4. Fixed endpoint (mV)

- Use  to change the y-axis from the pH/mV reading to the derivate value (equivalence point titrations only).
- Use  to save the graph as a bitmap (available when titration is complete).

2.5.2. STOPPING A TITRATION / DIRECT READING

The titration or analysis is terminated when one of the below listed conditions is met.

Analysis completed

This is the only mode with valid final result values.

The endpoint or stable reading was successfully detected, the final results will be displayed.

Manually terminated

The current titration or analysis was terminated by the user before the endpoint was detected.

Limits exceeded

The maximum titrant volume was delivered without reaching the endpoint.

An error message is displayed on the screen.

Critical error

A critical error occurred and the titration was stopped. These errors are typically related to the dosing system.

An error message is displayed on the screen.

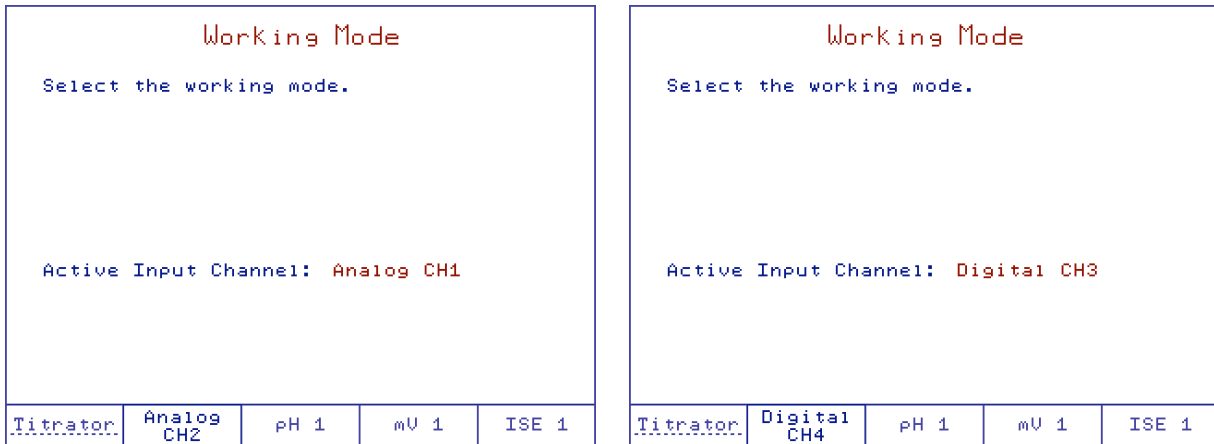
Potential out of range

The measured values from the electrode are outside the potential range.

An error message is displayed on the screen.

2.6. pH MODE

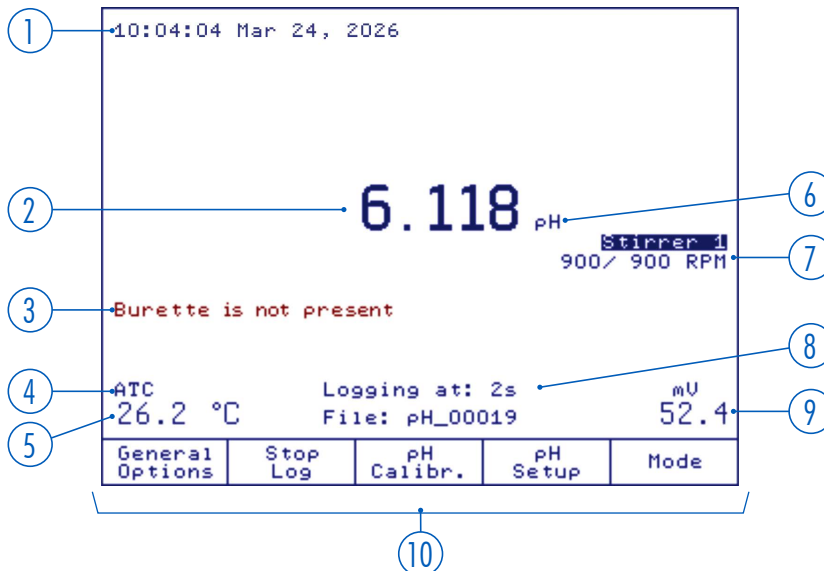
Press Mode when in main screen, to select preferred working mode i.e. **Titration**, **pH**, **mV**, **ISE**.



When one of these keys is pressed, the titrator will enter the selected mode.

- Titration » switches to **Titration** mode.
- Analog CH1 or Analog CH2 » switches the Active Input for **pH**, **mV**, and **ISE** mode.
- Analog CH1 / Analog CH2 or Digital CH3 / Digital CH4 » switches the Active Input for **pH** and **mV** mode (if installed).
- pH 1 or pH 2 » switches to **pH** mode.
- mV 1 or mV 2 » switches to **mV** mode.
- ISE 1 or ISE 2 » switches to **ISE** mode.

2.6.1. DISPLAY








- | | |
|------------------------------------|-------------------------|
| 1. Time and Date | 6. Units |
| 2. pH value | 7. Stirrer information |
| 3. Status bar | 8. Logging info |
| 4. Temperature compensation status | 9. mV reading |
| 5. Temperature reading | 10. Virtual option keys |






If one of these keys is pressed, the associated function is immediately performed.

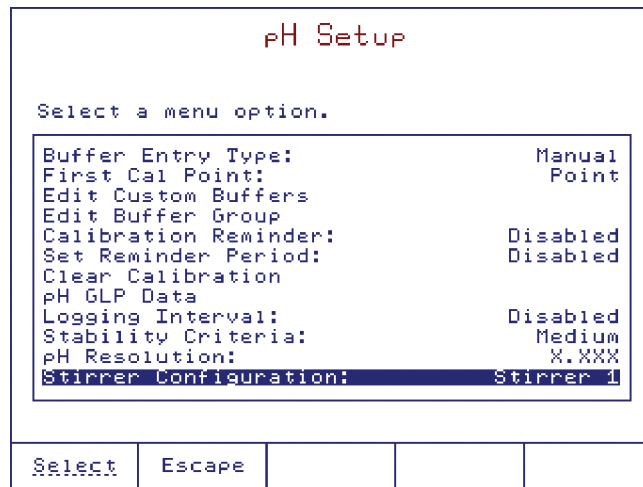
Some of the keys are active only in specific screens.

- Use General Options to access options that are not directly related to the measurement process.
See [2.3. General Options](#) section for more information.

- Use  to store the current pH reading. See [2.6.4. Logging](#) section for more information.
- or
- Use  to start the interval log. See [2.6.4. Logging](#) section for more information.
- Use  to enter the pH calibration screen. See [2.6.3. pH Calibration](#) section for more information.
- Use  to enter the pH setup screen. Parameters are associated with pH measurements and calibration. See [2.6.2. pH Setup](#) section for more information.
- Use  to switch between measurement modes: Titrator, pH, mV, or ISE.

2.6.2. pH SETUP

- To access pH Setup, press  option key while in pH mode.
- Use  and  keys to highlight the desired option.
- Press  or  to access the selected option.



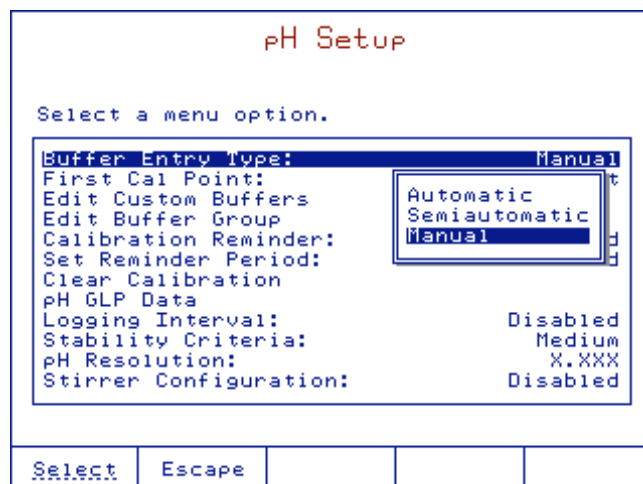
2.6.2.1. Buffer Entry Type

Option: Automatic, Semiautomatic, Manual

Automatic The instrument automatically selects the pH calibration point as the closest buffer from the predefined buffer group. See [2.6.2.4. Edit Buffer Group](#) section for more information.

Semiautomatic The instrument automatically selects the closest buffer from the available buffers (standard and custom buffers).

Manual The calibration buffer must be manually selected during calibration from the available buffer list (standard and custom buffers).

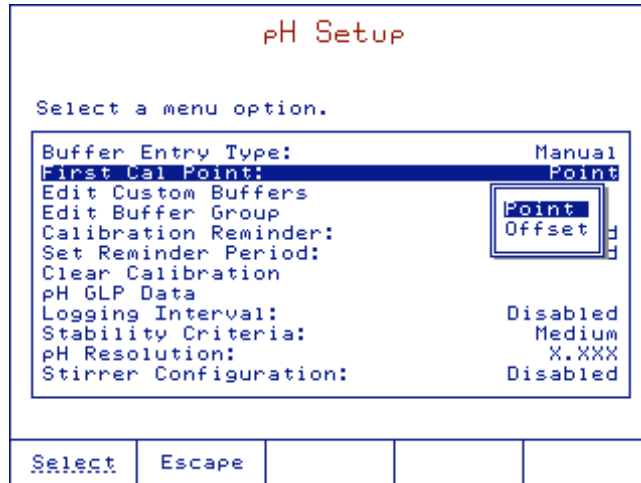


2.6.2.2. First-Calibration Point

Option: Point or Offset

Point The slope values adjacent to the calibration points will be reevaluated (normal calibration).

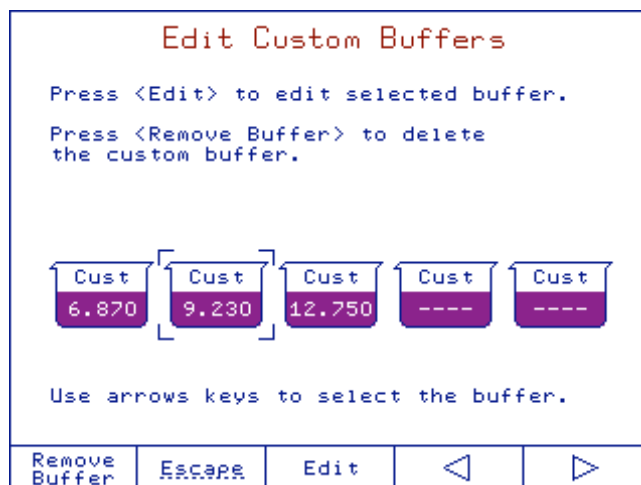
Offset The existing slope values will not be changed.







2.6.2.3. Edit Custom Buffers

When using buffers other than the standard ones, use the Edit Custom Buffers option to set the desired pH value. Up to five pH custom buffers can be set.

Note: Custom buffers are not temperature compensated. Enter the value of the buffer at the calibration temperature.



1. Use the  and  keys to select the desired buffer.
2. Press  to delete the selected buffer.
3. Press  to edit the selected buffer.

Edit Custom Buffers

Enter the custom buffer value.

9.230 pH

Cust	Cust	Cust	Cust	Cust
6.870	9.230	12.750	----	----

Low limit: -2 pH
High limit: 20 pH

ACCEPT	Escape	Delete Digit		
--------	--------	--------------	--	--

4. Use the numeric keypad to enter the pH buffer value.
5. Press Accept to save the value.
6. Press Escape to return to pH Setup menu.

2.6.2.4. Edit Buffer Group

Option: Up to five buffers

Select up to five buffers from the available buffers (Hanna[®] or custom) to be used for automatic buffer recognition.

Within the buffer group, pH values must be at least 1.5 pH apart.

If the buffer group already contains five pH buffers, at least one pH buffer has to be removed in order to add another buffer.

Edit Buffer Group

Available Buffers

Hanna	Hanna	Hanna	Hanna	Hanna
1.679	3.000	4.010	6.862	7.010
Hanna	Hanna	Hanna		
9.177	10.010	12.450		

Buffer Group

Hanna	Hanna	Hanna	Hanna	Hanna
1.679	4.010	6.862	9.177	12.450

REMOVE	Escape	▶	▲	▼
--------	--------	---	---	---

- Use the arrow keys to select the pH buffer to be included in / removed from the buffer group.
- Use Add or Remove to add or remove the selected pH buffer to / from buffer group.
- Use Escape to return to pH Setup menu.

2.6.2.5. Calibration Reminder

Option: Daily, Periodic, Disabled

Daily The calibration reminder will appear daily at a specified time.

Periodic The calibration reminder will appear after the set time since the last calibration has elapsed.

Disabled The calibration reminder will not appear.

Calibration Reminder							
Select a menu option.							
<table border="1"> <tr> <td>Daily</td> </tr> <tr> <td>Periodic</td> </tr> <tr> <td>Disabled</td> </tr> </table>					Daily	Periodic	Disabled
Daily							
Periodic							
Disabled							
Select	Escape						

2.6.2.6. Set Reminder Period

Option: Disabled to 31 days, 23 hours and 59 minutes

If *Daily* or *Periodic* option was selected for the calibration reminder, the reminder period must also be set.

For a daily reminder period, the time of day can be set.

For a periodic reminder period, the number of days, hours and minutes can be set.

Periodic Calibration Reminder				
Enter the time period that must be passed since the last calibration, whereafter the calibration reminder appears.				
10	2	30		
days	hours	minutes		
Press Next to move to the next entry.				
Accept	Escape	Delete Digit	Next	Off

- Use Next to move the cursor to the next field.
- Use Accept to save the changes or Escape to return to the previous screen.
- Use Off to disable the calibration reminder and return to pH setup.

2.6.2.7. Clear Calibration

This option clears the existing pH calibration for the selected measurement input. If the calibration is cleared, the factory calibration will be used.

- Use to clear a previous calibration.
- Alternatively, use to return to the previous screen without clearing the calibration.

Clear Calibration				
Press <Clear> to clear all calibration points.				
Press <Escape> to return without clearing the calibration points.				
Clear	Escape			

2.6.2.8. pH GLP Data

Display the pH calibration data.

pH GLP Data				
Analog CH1				
Last Calibration: 10:13 Mar 24, 2026				
Offset: -0.1 mV Average Slope: 100.7%				
1.679pH (Hanna) 316.2mV 26.3°C A				
10:10:30 Mar 24, 2026				
4.010pH (Hanna) 177.5mV 26.3°C A				
10:09:11 Mar 24, 2026				
7.010pH (Hanna) -0.6mV 26.3°C A				
10:08:40 Mar 24, 2026				
10.010pH (Hanna) -179.1mV 26.3°C A				
10:09:43 Mar 24, 2026				
12.450pH (Hanna) -325.6mV 26.3°C A				
10:13:15 Mar 24, 2026				
	Escape			

2.6.2.9. Logging Interval

Option: 2 seconds to 8 hours 59 minutes 59 seconds

Set the logging interval to be used for automatic logging.

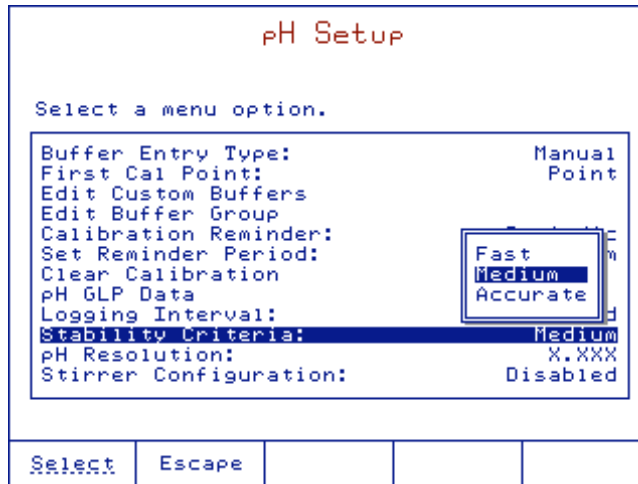
Select Off to enable manual logging.

Logging Interval				
Enter the data logging interval.				
<div style="display: flex; justify-content: space-around; align-items: center;"> <div style="text-align: center;"> <div style="border: 1px solid black; width: 20px; height: 15px; background-color: black; margin: 0 auto;"></div> <div style="margin: 2px;">0</div> <div style="margin: 2px;">hours</div> </div> <div style="text-align: center;"> <div style="margin: 2px;">0</div> <div style="margin: 2px;">minutes</div> </div> <div style="text-align: center;"> <div style="margin: 2px;">2</div> <div style="margin: 2px;">seconds</div> </div> </div>				
Press Next to move to the next entry.				
Accept	Escape	Delete Digit	Next	Off

2.6.2.10. Signal Stability Criteria

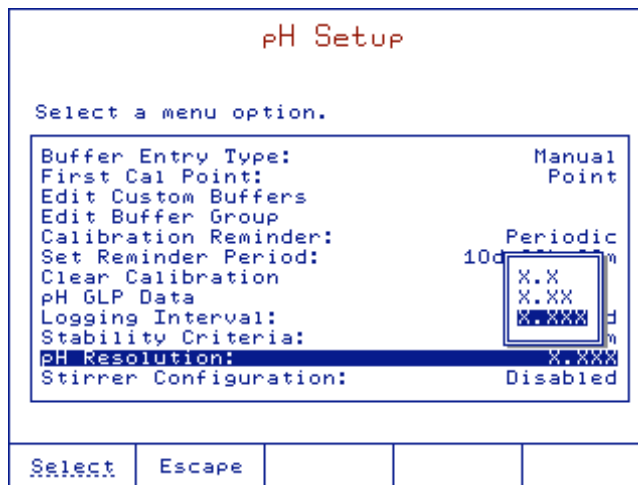
Option: Fast, Medium, Accurate

- Fast** Quicker results, less accuracy
- Medium** Medium speed results, medium accuracy
- Accurate** Slower results, high accuracy



2.6.2.11. pH Resolution

Option: X.X (one), X.XX (two), X.XXX (three) decimal places



2.6.2.12. Stirrer Configuration

pH Setup	
Select a menu option.	
Buffer Entry Type:	Manual Point
First Cal Point:	Point
Edit Custom Buffers	
Edit Buffer Group	
Calibration Reminder:	Disabled
Set Reminder Period:	Disabled
Clear Calibration	
pH GLP Data	
Logging Interval:	Disabled
Stability Criteria:	Medium
pH Resolution:	X.XXX
Stirrer Configuration:	Stirrer 1

Select	Escape			
--------	--------	--	--	--

Stirrer Option

Option: Disabled, Stirrer 1, Stirrer 2 (if installed)

Stirrer Configuration	
Select a menu option.	
Stirrer Option:	Stirrer 1
Stirring Speed:	

Disabled
Stirrer 1
Stirrer 2

Select	Escape			
--------	--------	--	--	--

Stirring Speed

Option: 200 to 2500 RPM

Stirring Speed	
Enter the speed of the stirrer within below range.	
1100 RPM	
The range is from 200 to 2500 RPM.	

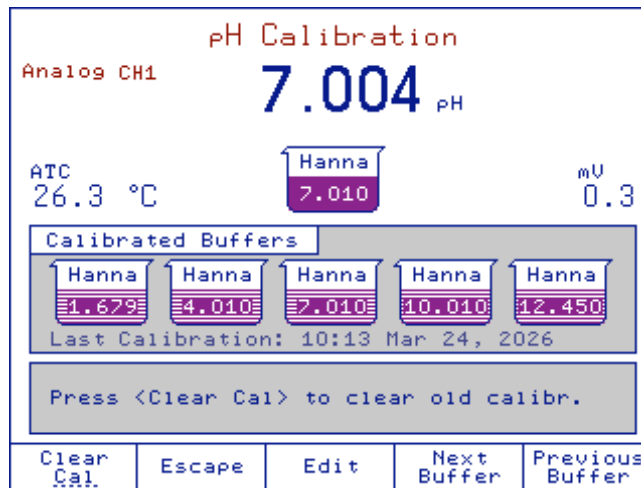
Accept	Escape	Delete Digit		
--------	--------	--------------	--	--

2.6.3. pH CALIBRATION

Calibrate the instrument often, especially if high accuracy is required.

The instrument should be recalibrated:

- Whenever the pH electrode is replaced.
- At least once a week.
- After testing aggressive chemicals.
- When “No pH Calibration” or “pH Calibration Expired” message appears on the display.



Preparation

- Pour small quantities of the buffer solutions into clean beakers.
If possible, use plastic beakers to minimize any EMC interferences.
- For accurate calibration and to minimize cross-contamination, use two beakers for each buffer solution: one for rinsing the electrode and one for calibration.
- If measuring in the acidic range, use pH 7.01 or 6.86 as the first buffer and pH 4.01 / 3.00 or 1.68 as the second buffer.
- If measuring in the alkaline range, use pH 7.01 or 6.86 as the first buffer and pH 10.01 / 9.18 or 12.45 as the second buffer.
- For extended range measurements (acidic and alkaline), perform a five-point calibration by selecting five buffers across the entire pH range.

Calibration procedure

During calibration, the user has a choice of 8 standard buffers (pH 1.68, 3.00, 4.01, 6.86, 7.01, 9.18, 10.01, 12.45) and up to 5 custom buffers.

For accurate measurements it is recommended to perform a five-point calibration. However, at least a two-point calibration is suggested.

For pH titrations, the selected buffers should bracket the endpoint; if endpoint value is at 8.5, use 7.01 or 6.86 and 9.18 or 10.01 for calibration.

To calibrate:

1. Press pH
Calibr..

If the instrument was calibrated before, press Clear
Cal to clear previous calibration.

Note: It is very important to clear calibration history when a new electrode is used.

2. Immerse the pH electrode and the temperature probe approximately 4 cm (1.5") into a buffer solution and stir gently.
3. If necessary, use Next
Buffer or Previous
Buffer to select the pH calibration buffer value.
4. Once the reading has stabilized, press Accept to update the calibration.
The calibration buffer will be added to the Calibrated Buffers section.
5. Rinse the pH electrode and the temperature probe, then immerse them into the next buffer solution and follow the above procedure.
Alternatively, press Escape to exit the calibration.

Notes:

- The new calibration points will replace old ones if the difference between them is ± 0.2 pH.
- Buffers used in previous calibrations will not have a solid background.
- If calibrating with a standard buffer in MTC mode, press Edit to modify the pH value and the temperature. The values can be adjusted using the numeric keys. Press Accept to save the new values.

Manual Edit				
Edit pH buffer and manual temperature.				
Buffer: 7.010 pH				
Temperature: 25.0 °C				
Low limit: 6.990 pH				
High limit: 7.030 pH				
Press Next to move to the next entry.				
ACCEPT	Escape	Delete Digit	Next	

- In ATC mode, the pH value for custom buffers can be modified by pressing Edit.
- If the Automatic Buffer entry type was selected for the calibration procedure, the titrator will automatically select the buffer closest to the measured pH value from the buffer group.
- If the Semiautomatic Buffer entry type was selected, use the Previous
Buffer or Next
Buffer to select the buffer. Only buffers in the buffer group will be displayed.

Calibration Messages**Wrong Buffer. Please check the buffer.**

Message displayed when the difference between the pH reading and the value of the selected calibration buffer is significant. Check if the appropriate calibration buffer has been selected.

Wrong buffer temperature.

Message displayed if the buffer temperature is out of the defined temperature range.

Clean the electrode or check the buffer. Press Accept to update calibration.

This message alerts the user that some dirt or deposits could be on the electrode, or the buffer is contaminated.

Slope too low. Please check the buffer .

This message appears if the current slope is under 80% of the default slope.

Recalibrate the instrument using fresh buffers.

**Slope too high. Press Clear
Cal to clear the old calibration.**

Message appears as a result of an erroneous slope condition.

2.6.4. LOGGING

Data logging is available in pH mode.

Available options are logging on demand (Manual Logging) and automatic logging at predefined time intervals (Interval Logging). The the logging report can be customized. See [Setting up pH / mV / ISE Report](#) section for more information.

Interval Logging

The logging interval is set in the pH Setup screen.

- Press Start
Log to start the log.
The logging interval and name of logging file will be displayed on the measure screen.
- Press Stop, to stop the automatic logging.

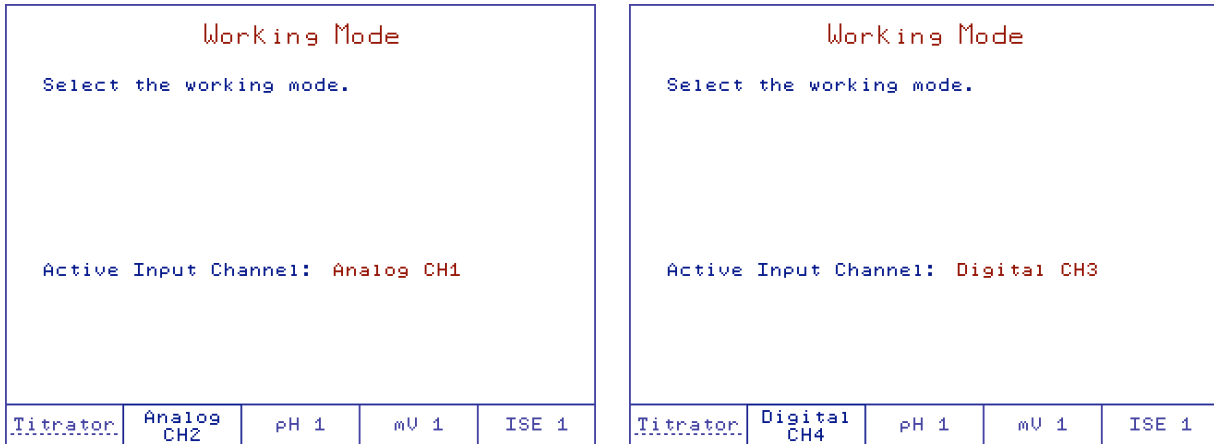
Manual Logging

Press  from the pH measurement screen, to manually log pH readings.

A new record will be added to the report every time  is pressed.

2.7. mV MODE

By pressing  from the main screen, the titrator can be switched to **Titration**, **pH**, **mV** or **ISE** modes.







When one of these keys is pressed, the titrator will enter the selected mode.



 » switches to **Titration** mode.

 or  » switches the active measurement input for **pH**, **mV**, and **ISE** mode (if installed).

 /  or  /  » switches the Active Input for **pH** and **mV** mode (if installed).

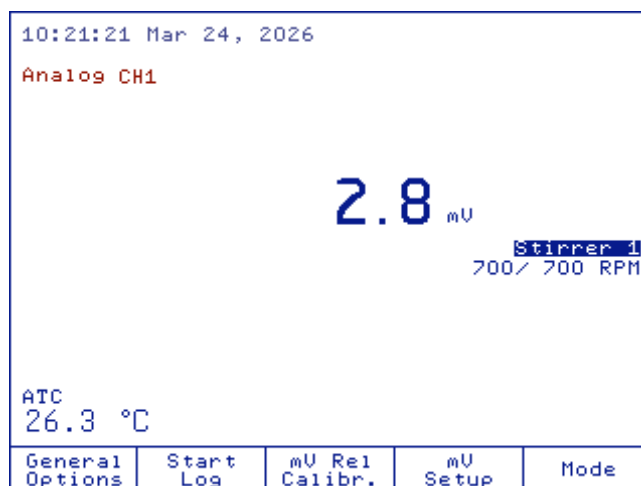
 or  » switches to **pH** mode.

 or  » switches to **mV** mode.

 or  » switches to **ISE** mode.


2.7.1. DISPLAY

The mV screen is shown here.








If one of these keys is pressed, the associated function is immediately performed.

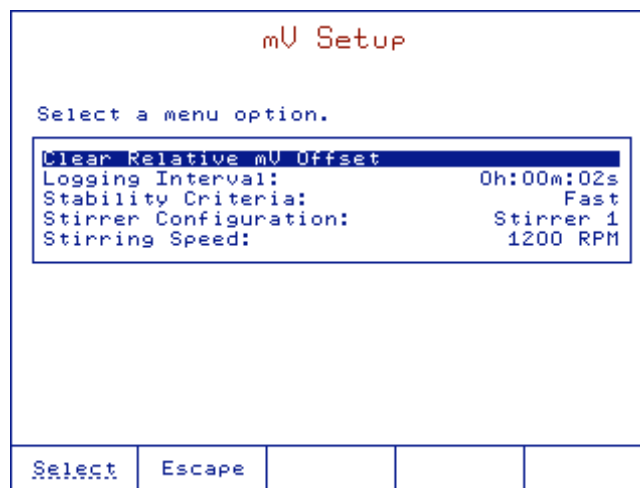
Some of the keys are active only in specific screens.

- Use  to access options that are not directly related to the measurement process.



See [2.3. General Options](#) section.

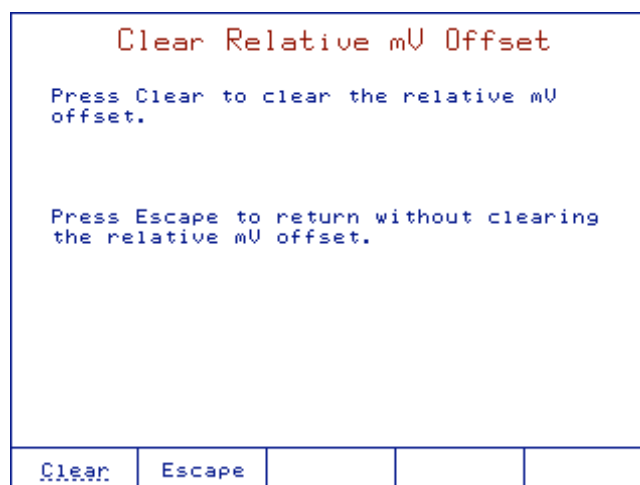
- Use  to store the current pH reading.
See [2.7.4. Logging](#) section.
- or
- Use  to start the interval log.
See [2.7.4. Logging](#) section.
- Use  to enter the pH calibration screen.
See [2.7.3. Relative mV Calibration](#) section.
- Use  to enter the pH setup screen.
Parameters are associated with pH measurements and calibration.
See [2.7.2. mV Setup](#) section.
- Use  to switch between measurement modes: Titrator, pH, mV, or ISE.

2.7.2. mV SETUP



Clear Relative mV Offset

- Press  to clear the relative mV offset.
- Alternatively, press  to return to the previous screen.



Logging Interval

Option: 2 seconds to 8 hours 59 minutes 59 seconds

Press off to enable manual logging.



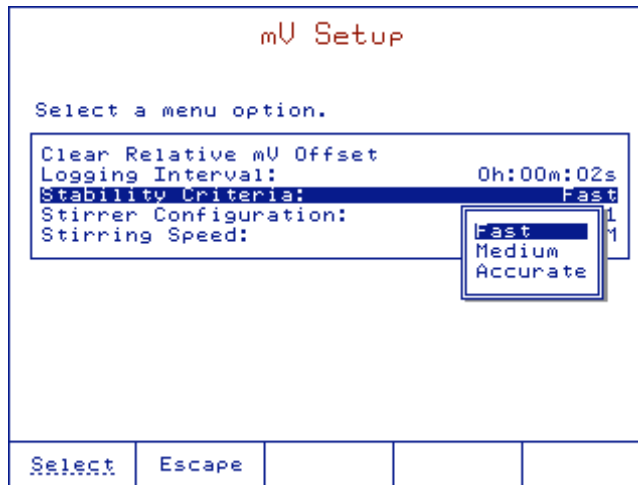
Stability Criteria

Option: Fast, Medium, Accurate

Fast Quicker results, less accuracy

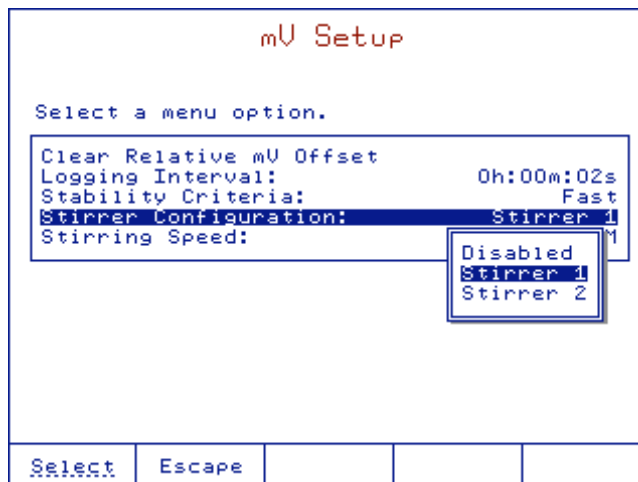
Medium Medium speed results, medium accuracy

Accurate Slower results, high accuracy



Stirrer Configuration

Option: Stirrer 1, Stirrer 2 (if available), Disabled






Stirring Speed

Option: 200 to 2500 RPM

Stirring Speed				
Enter the speed of the stirrer within below range.				
1100 RPM				
The range is from 200 to 2500 RPM.				
ACCEPT	Escape	Delete Digit		

2.7.3. RELATIVE mV CALIBRATION

- Press  to accept the value.
- Press  to cancel this operation and return to the previous screen.
- Press  to delete the last digit.

Relative mV				
Analog CH1				
Set the value for the relative mV offset.				
Absolute mV: 2.7 mV				
Stirrer #1 1100/1100 RPM				
Relative mV: 2.7 mV				
Low limit: -1997.3 mV				
High limit: 2002.7 mV				
ACCEPT	Escape	Delete Digit		

2.7.4. LOGGING

Data logging is available in mV mode.


Available options are logging on demand (Manual Logging) or automatic logging at predefined time intervals (Interval Logging).

The logging report can be customized.


See [Setting up pH / mV / ISE Report](#) section for more information.


Interval Logging

The logging interval is set in the mV Setup screen.

- Press  to start the log.
The logging interval and name of logging file will be displayed on the measure screen.
- To stop the automatic logging, press Stop.

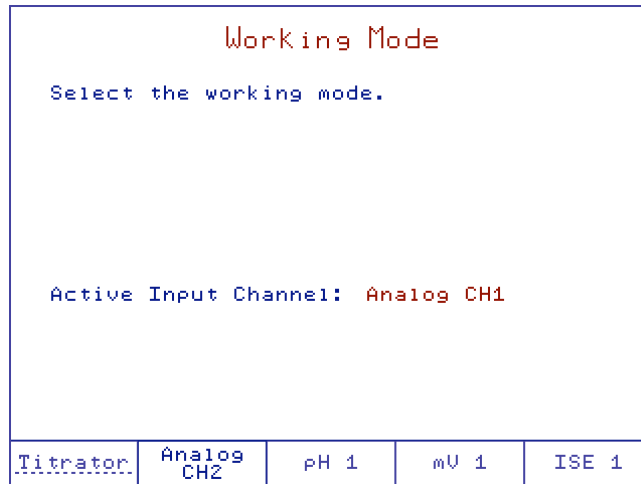
Manual Logging

To manually log mV readings, press  from the mV measurement screen.










A new record will be added to the report every time  is pressed.

2.8. ISE MODE

Press  from the main screen to switch between **Titration**, **pH**, **mV**, or **ISE** mode.

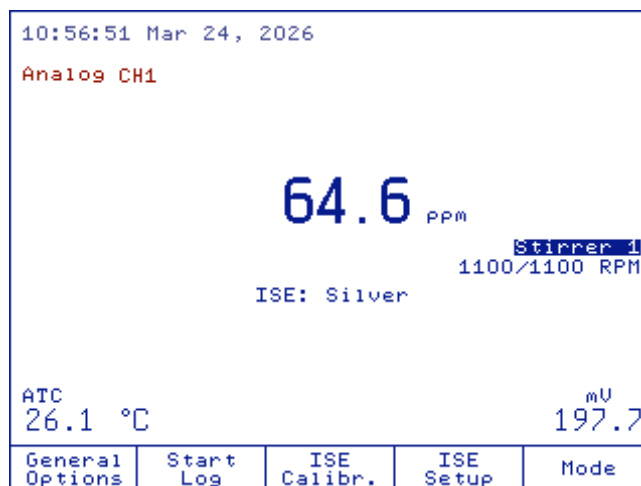


When one of these keys is pressed, the titrator will enter the selected mode.







-  » switches to **Titration** mode.
-  or  » switches the Measurement Input for **pH**, **mV**, and **ISE** mode (if installed).
-  or  » switches to **pH** mode.
-  or  » switches to **mV** mode.
-  or  » switches to **ISE** mode.

2.8.1. DISPLAY


The ISE screen is shown below.



If one of these keys is pressed, the associated function is immediately performed. Some of the keys are active only in specific screens.

- Use  to access options that are not directly related to the measurement process.
See [2.3. General Options](#) section for more information.
- Use  to store the current concentration reading. See [2.8.4. Logging](#) section for more information.
- or
- Use  to start the interval log.
See [2.8.4. Logging](#) section for more information.
- Use  to enter the ISE calibration screen.
See [2.8.3. ISE Calibration](#) section for more information.
- Use  to enter the ISE setup screen. Parameters are associated with ISE measurements and calibration.
- Use  to switch between measurement modes: **Titration**, **pH**, **mV**, or **ISE**.

2.8.2. ISE SETUP

To access the ISE Setup, press  option key in ISE mode.

ISE Setup	
Select a menu option.	
Calibration Group:	All Standards
Temperature Compensation:	Disabled
Isopotential Point:	None
Edit Custom Standards:	
Edit Standards Group:	
Calibration Reminder:	Disabled
Set Reminder Period:	Disabled
Clear Calibration	
ISE GLP Data	
Electrode Type:	Silver
Concentration Unit:	PPM
Logging Interval:	0h:00m:02s
Stability Criteria:	Medium
Select	Escape

2.8.2.1. Calibration Group

Option: All Standards or Standards Group

All Standards Includes both standard and custom solutions.

Standards Group Includes only the standards selected by the user.

ISE Setup	
Select a menu option.	
Calibration Group:	All Standards
Temperature Compensat:	Disabled
Isopotential Point:	None
Edit Custom Standards	
Edit Standards Group:	All Standards Standards Group
Calibration Reminder:	Disabled
Set Reminder Period:	Disabled
Clear Calibration	
ISE GLP Data	
Electrode Type:	Silver
Concentration Unit:	PPM
Logging Interval:	0h:00m:02s
Stability Criteria:	Medium
Select	Escape

2.8.2.2. Temperature Compensation

Option: Enabled or Disabled

Note: When Temperature compensation is enabled, the isopotential point must also be set.

ISE Setup	
Select a menu option.	
Calibration Group:	All Standards
Temperature Compensation:	Disabled
Isopotential Point:	20.0 ppm
Edit Custom Standards:	Disabled
Edit Standards Group:	Enabled
Calibration Reminder:	Disabled
Set Reminder Period:	Disabled
Clear Calibration	
ISE GLP Data	
Electrode Type:	Silver
Concentration Unit:	ppm
Logging Interval:	0h:00m:02s
Stability Criteria:	Medium
Select	Escape

2.8.2.3. Isopotential Point (Temperature Compensation)

Option: 1.00 E^{-2} to 1.00 E^{+5} ppm

This option allows the user to set an isopotential point for the selected electrode when temperature compensation is enabled.



The isopotential point is edited in ppm units only.

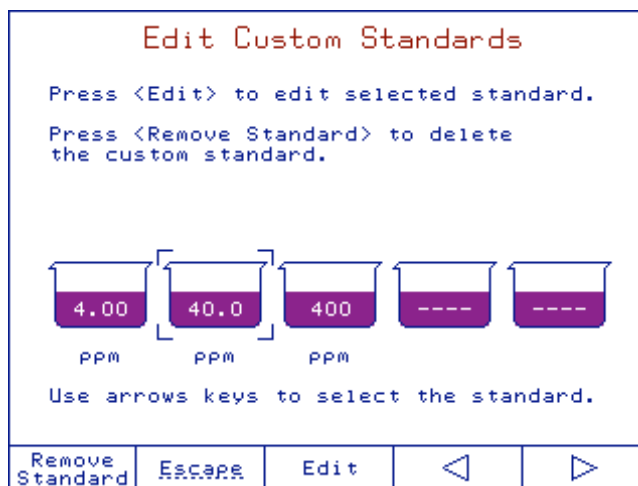
The isopotential point will vary for different electrodes, if measurements are going to be made at several temperatures, the value should be entered if it is known.

Isopotential Point	
Enter the value for isopotential point.	
20.0 ppm	
Low limit: 1.00E-2 ppm	
High limit: 1.00E+5 ppm	
Accept	Escape
Delete Digit	Exponent

2.8.2.4. Editing Custom Standards




Option: Up to five

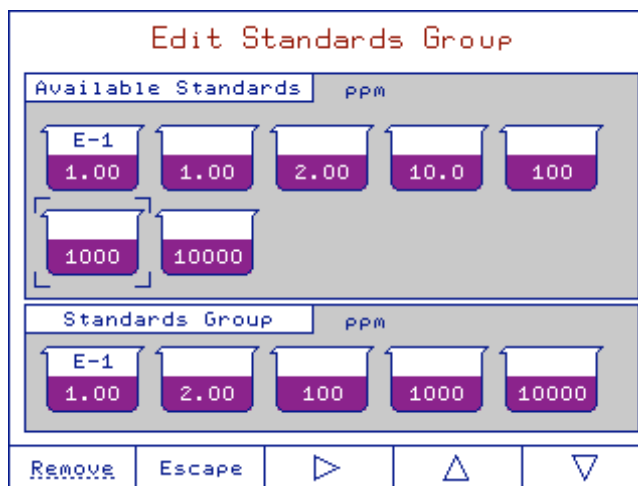
1. Use the < and > keys to select the standard.
2. Press  to delete the standard.
3. Press  to edit the selected custom standard.
Use the numeric keys to edit the standard.



2.8.2.5. Editing Standard Group

Option: Up to 5 standards

- Use the < and > keys to select the standard to be included in or removed from the standard group.
- Use the  or  keys to add or remove the selected standard to or from the standard group.
- Use the  key to return to ISE Setup menu.



2.8.2.6. Calibration Reminder

Option: Daily, Periodic, Disabled

Daily The calibration reminder will appear daily, at the specified time.

Periodic The calibration reminder will appear after the set time since the last calibration has elapsed.

Disable The calibration reminder will not appear.

Calibration Reminder							
Select a menu option.							
<table border="1"> <tr> <td>Daily</td> </tr> <tr> <td>Periodic</td> </tr> <tr> <td>Disabled</td> </tr> </table>					Daily	Periodic	Disabled
Daily							
Periodic							
Disabled							
Select	Escape						

2.8.2.7. Setting Reminder

If Daily or Periodic option was selected for the calibration reminder, the reminder period must also be set.

For a daily reminder, the time of day can be set.

For a periodic reminder, the number of days, hours, and minutes can be set.

Periodic Calibration Reminder				
Enter the time period that must be passed since the last calibration, whereafter the calibration reminder appears.				
10	2	30		
days	hours	minutes		
Press Next to move to the next entry.				
Accept	Escape	Delete Digit	Next	Off

- Use Next to move the cursor to the next field.
- Use Accept to save the changes or Escape to return to the previous screen.
- Use Off to disable the calibration reminder and return to ISE setup menu.

2.8.2.8. Clearing Calibration

This option clears the existing ISE calibration.

If the calibration is cleared, a new calibration must be done in order to take measurements.

Use Clear key to clear the previous calibration or Escape key to return to the previous screen.

Clear Calibration				
Press <Clear> to clear all calibration points. Press <Escape> to return without clearing the calibration points.				
Clear	Escape			

2.8.2.9. ISE GLP Data



Displays the ISE calibration data.

ISE GLP Data				
Analog CH1 Last Calibration: 13:42 Mar 24, 2026 Slope: 100.8% ISE: Silver Isopotential Point: 20.0 ppm 1.00E-1 ppm, 0.1mV 28.1°C A 13:39:43 Mar 24, 2026 1.00 ppm, 59.5mV 28.1°C A 13:40:39 Mar 24, 2026 2.00 ppm, 77.6mV 28.1°C A 13:41:25 Mar 24, 2026 10.0 ppm, 120.0mV 28.1°C A 13:41:45 Mar 24, 2026 100 ppm, 181.0mV 28.2°C A 13:42:17 Mar 24, 2026				
	Escape			

2.8.2.10. Electrode Type

Option: Ammonia, Bromide, Cadmium, Calcium, Carbon Dioxide, Chloride, Cupric, Cyanide, Fluoride, Iodide, Lead, Nitrate, Potassium, Silver, Sodium, Sulfate, Sulfide, five custom electrodes

Electrode Type				
Select a menu option.				
<div style="border: 1px solid black; padding: 5px;"> Ammonia Bromide Cadmium Calcium Carbon Dioxide Chloride Cupric Cyanide Fluoride Iodide Lead Nitrate Potassium Silver </div>				
Select	Escape	View	Page Up	Page Down

- Use  to display the ion constants (name, molar weight, electric charge/slope).
- Use  to return to the setup screen.

Ion Constants										
View Ion constants.										
<table border="1"> <tr> <td>Name:</td> <td>Silver</td> </tr> <tr> <td>Molar Weight:</td> <td>107.868 g/mol</td> </tr> <tr> <td>Electric Charge / Slope:</td> <td>1 / 59.16</td> </tr> </table>					Name:	Silver	Molar Weight:	107.868 g/mol	Electric Charge / Slope:	1 / 59.16
Name:	Silver									
Molar Weight:	107.868 g/mol									
Electric Charge / Slope:	1 / 59.16									
<table border="1"> <tr> <td>Escape</td> <td></td> <td></td> <td></td> <td></td> </tr> </table>					Escape					
Escape										

The ion constants for custom electrodes can be modified.

Name

Option: up to 10 characters

Electrode Name																																																																																																																																																																				
Select the highlighted letter by using the arrow keys then press "Enter". Select the empty field for a space. Press Accept to save the entire name.																																																																																																																																																																				
<table border="1"> <tr><td>A</td><td>B</td><td>C</td><td>D</td><td>E</td><td>F</td><td>G</td><td>H</td><td>I</td><td>J</td><td>K</td><td>L</td></tr> <tr><td>M</td><td>N</td><td>O</td><td>P</td><td>Q</td><td>R</td><td>S</td><td>T</td><td>U</td><td>V</td><td>W</td><td>X</td><td>Y</td></tr> <tr><td>Z</td><td>a</td><td>b</td><td>c</td><td>d</td><td>e</td><td>f</td><td>g</td><td>h</td><td>i</td><td>j</td><td>k</td><td>l</td></tr> <tr><td>m</td><td>n</td><td>o</td><td>p</td><td>q</td><td>r</td><td>s</td><td>t</td><td>u</td><td>v</td><td>w</td><td>x</td><td>y</td></tr> <tr><td>z</td><td>À</td><td>Á</td><td>Â</td><td>Ã</td><td>Ä</td><td>Å</td><td>Æ</td><td>Ç</td><td>È</td><td>É</td><td>Ê</td><td>Ë</td><td>Ì</td><td>Í</td><td>Î</td><td>Ï</td></tr> <tr><td>Ò</td><td>Ó</td><td>Ô</td><td>Õ</td><td>Ö</td><td>Ù</td><td>Ú</td><td>Û</td><td>Ü</td><td>Ý</td><td>à</td><td>á</td><td>â</td><td>ã</td><td>ä</td><td>å</td></tr> <tr><td>æ</td><td>ç</td><td>è</td><td>é</td><td>ê</td><td>ë</td><td>ì</td><td>í</td><td>î</td><td>ï</td><td>ò</td><td>ó</td><td>ô</td><td>õ</td><td>ö</td><td>ø</td></tr> <tr><td>ù</td><td>ú</td><td>û</td><td>ü</td><td>¿</td><td>¡</td><td>*</td><td>/</td><td>_</td><td>\$</td><td>'</td><td>^</td><td>#</td><td>:</td><td>.</td><td>,</td></tr> <tr><td>0</td><td>1</td><td>2</td><td>3</td><td>4</td><td>5</td><td>6</td><td>7</td><td>8</td><td>9</td><td>%</td><td>&</td><td>.</td><td>,</td><td></td><td></td></tr> <tr><td>?</td><td>!</td><td>(</td><td>)</td><td>[</td><td>]</td><td><</td><td>></td><td>=</td><td>/</td><td>+</td><td>-</td><td></td><td></td><td></td><td></td></tr> <tr><td colspan="12" style="text-align: center;">Custom: </td></tr> </table>					A	B	C	D	E	F	G	H	I	J	K	L	M	N	O	P	Q	R	S	T	U	V	W	X	Y	Z	a	b	c	d	e	f	g	h	i	j	k	l	m	n	o	p	q	r	s	t	u	v	w	x	y	z	À	Á	Â	Ã	Ä	Å	Æ	Ç	È	É	Ê	Ë	Ì	Í	Î	Ï	Ò	Ó	Ô	Õ	Ö	Ù	Ú	Û	Ü	Ý	à	á	â	ã	ä	å	æ	ç	è	é	ê	ë	ì	í	î	ï	ò	ó	ô	õ	ö	ø	ù	ú	û	ü	¿	¡	*	/	_	\$	'	^	#	:	.	,	0	1	2	3	4	5	6	7	8	9	%	&	.	,			?	!	()	[]	<	>	=	/	+	-					Custom: 											
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Accept	Escape	Delete Letter	Cursor Left	Cursor Right																																																																																																																																																																

Molar Weight

Option: 0.001 g/mol to 1000.000 g/mol

Ion Molar Weight									
Set the value for Ion molar weight.									
<table border="1"> <tr> <td>107.868 g/mol</td> </tr> </table>					107.868 g/mol				
107.868 g/mol									
Low limit: 0.001 g/mol									
High limit: 1000.000 g/mol									
<table border="1"> <tr> <td>Accept</td> <td>Escape</td> <td>Delete Digit</td> <td></td> <td></td> </tr> </table>					Accept	Escape	Delete Digit		
Accept	Escape	Delete Digit							

Electric Charge / Slope

Option: 2/29.58; 1/59.16; -1/-59.16; -2/-29.58 or None/-59.16

Electric Charge / Slope				
Select the option.				
<div style="border: 1px solid black; padding: 5px;"> 2 / 29.58 1 / 59.16 -1 / -59.16 -2 / -29.58 None / -59.16 </div>				
Select	Escape			

2.8.2.11. Concentration UnitOptions: ppt (g/L), ppm (mg/L), ppb ($\mu\text{g/L}$), mg/mL, M (mol/L), mmol/L, %w/v, user defined

ISE Setup				
Select a menu option.				
<div style="border: 1px solid black; padding: 5px;"> Calibration Group: All Standards Temperature Compensation: Disabled Isopotential Point: None Edit Custom Standards: Edit Standards Group: Calibration Reminder: Set Reminder Period: Clear Calibration ISE GLP Data Electrode Type: Concentration Unit: ppm Logging Interval: 0h:00m:02s Stability Criteria: Medium </div>				
Select	Escape			

2.8.2.12. Logging Interval

Option: 2 seconds to 8 hours 59 minutes 59 seconds

Logging Interval				
Enter the data logging interval.				
<div style="display: flex; justify-content: space-around; align-items: center;"> <div style="text-align: center;"> 0 hours </div> <div style="text-align: center;"> 0 minutes </div> <div style="text-align: center;"> 2 seconds </div> </div>				
Press Next to move to the next entry.				
Accept	Escape	Delete Digit	Next	Off

2.8.2.13. Stability Criteria

Option: Fast, Medium, Accurate

Fast Quicker results, less accuracy

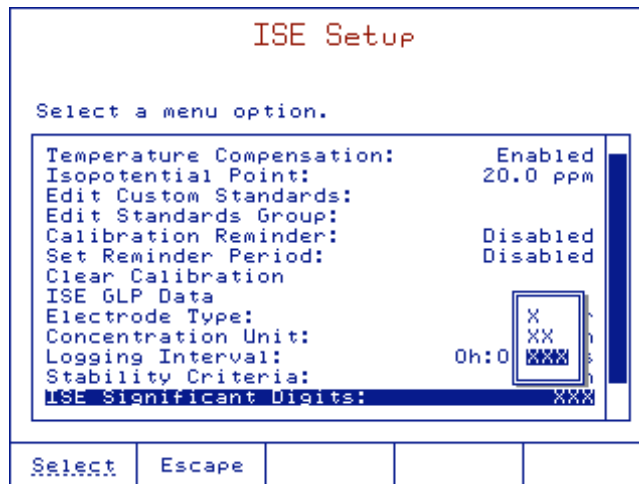
Medium Medium speed results, medium accuracy

Accurate Slower results, high accuracy

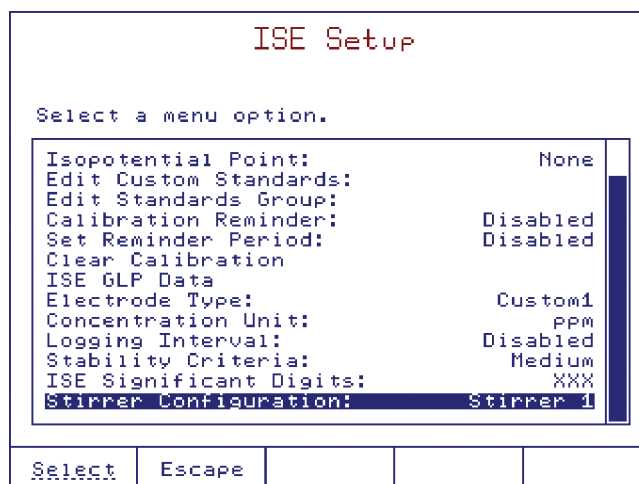


2.8.2.14. ISE Significant Digits

Option: X (one), XX (two), XXX (three)

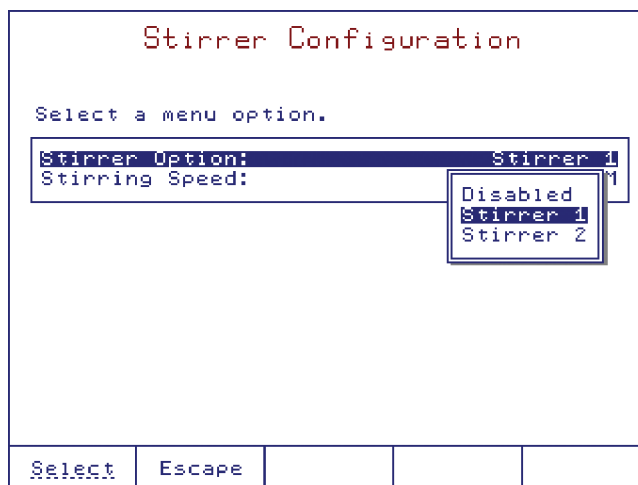


2.8.2.15. Stirrer Configuration



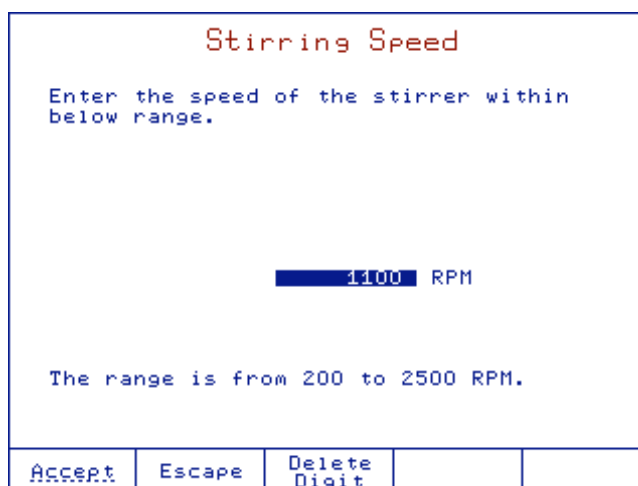
Stirrer Option

Option: Disabled, Stirrer 1, Stirrer 2 (if installed)



Stirring Speed

Option: 200 to 2500 RPM



2.8.3. ISE CALIBRATION

It is recommended to calibrate the instrument frequently if high accuracy is required.

Additionally, the instrument should be recalibrated whenever the “Calibrate Electrode” message appears on the LCD.

Due to electrode conditioning time, the electrode must be immersed for several seconds to stabilize.

The user will be guided step by step during calibration with easy-to-follow messages on the display. This makes the calibration a simple and error-free procedure.

Preparation

- Pour small quantities of the standard solution into clean beakers. If possible, use plastic beakers to minimize any EMC interferences.
- For accurate calibration and to minimize cross-contamination, use two beakers for each standard solution: one for rinsing the electrode and one for calibration.

Note: For accurate measurements, add the appropriate ISA (Ionic Strength Adjustment) to the calibration standards.

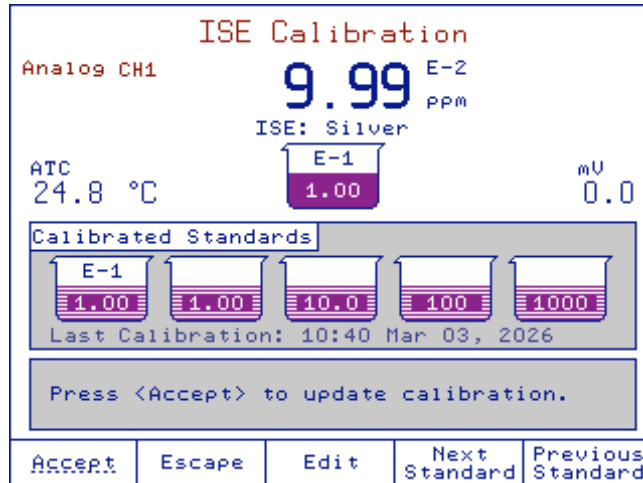
Calibration procedure

Before calibrating, make sure that the electrode type and concentration unit has been selected in ISE Setup.

Up to a five-points calibration is possible using any combination of five standard solutions and five custom solutions.

The ISE calibration and measurement can be performed with or without temperature compensation.

If the temperature compensation option is enabled, the isopotential point of the electrode must be set in ISE Setup.



1. Press ISE Calibr. from the main screen.
2. If the instrument was calibrated before and the calibration was not cleared, press Clear Cal. to clear the old calibration.
3. Immerse the ISE and the temperature probe approximately 2 cm into the standard with the lowest concentration.
4. Select the standard concentration with Next Standard or Previous Standard.
5. When the reading has stabilized, press Accept to update the calibration.
The calibration point value will be added to the Calibrated Standard list.
6. Select Next Standard and repeat the procedure with all of the available standards.
7. Press Escape to exit the calibration.

2.8.4. LOGGING

Data logging is available in ISE mode.

Available options are logging on demand (Manual Logging) or automatic logging at predefined time intervals (Interval Logging).

The logging report can be customized. See [Setting up pH / mV / ISE Report](#) section for more information.

Interval Logging

The logging interval is set in the ISE Setup screen.

- Press Start Log to start the log.
The logging interval and name of logging file will be displayed on the measure screen.
- To stop the automatic logging, press Stop.

Manual Logging

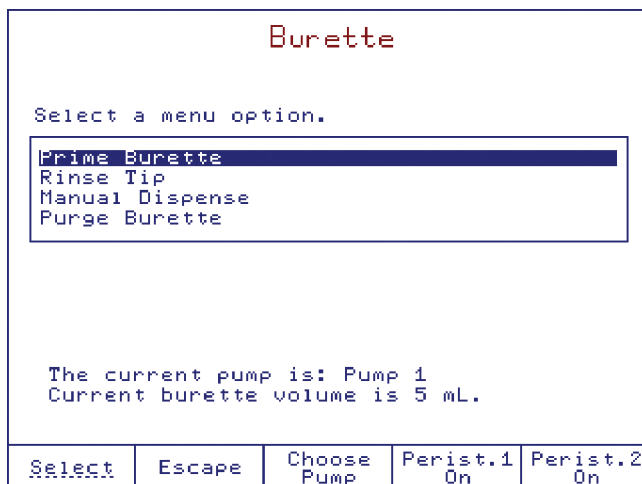
To manually log pH readings, press Save Reading from the ISE measurement screen.

A new record will be added to the report every time Save Reading is pressed.

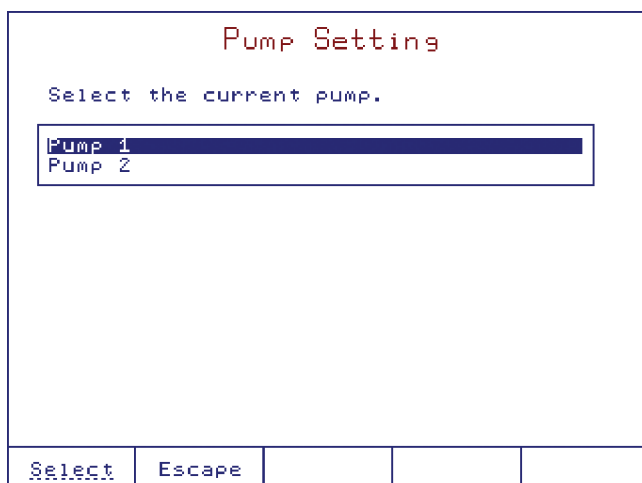
2.9. AUXILIARY FUNCTIONS

2.9.1. BURETTE

- To access the **Burette** screen, press Burette from the main titration screen.
- Highlight the desired option then press Select.



- Use Choose Pump to select the desired pump for burette operations (active only if two pumps are connected).



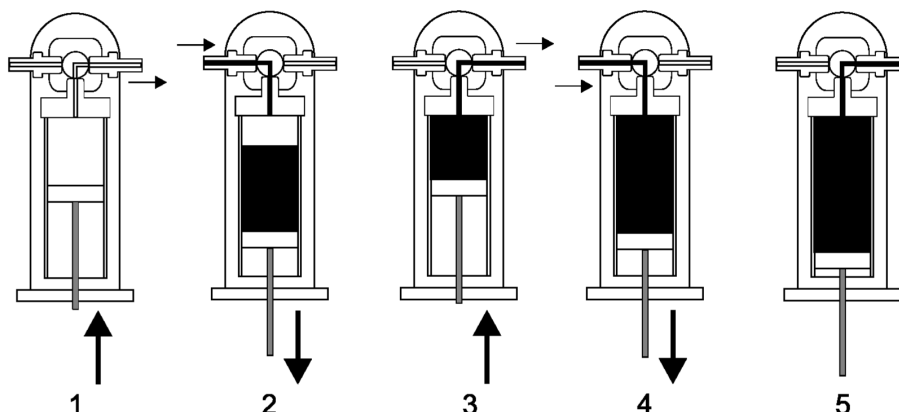
Priming the Burette

Option: Up to 5

The *Prime Burette* option is used to fill the burette with titrant or reagent before starting a titration.

The priming process consists of several cycles of filling and emptying the burette with titrant.

Two rinse cycles are shown in the figure below. The dispensing tube is connected on the right side and the aspiration tube on the left side.



Note: Before starting this operation, the aspiration tube must be inserted in the titrant bottle. A waste container should be placed under the dispensing tip to collect the waste solution.

- To prime the burette, select *Prime Burette*.
- Next, enter the number of rinses and press Accept.

We recommend at least three rinses to assure that the air bubbles are completely removed.

Total Burette Rinses				
Enter the total number of burette rinses.				
<div style="background-color: black; color: white; display: inline-block; padding: 5px 15px; border: 1px solid black;">3</div>				
A minimum of three rinses is recommended.				
Accept	Escape	Delete Digit		

Rinsing Burette Tip

A 2 mL dose of titrant will be dispensed from the burette when this operation is selected. This will eliminate any air in the dispensing tip.

Manual Dispense

This option allows a defined titrant volume to be dosed.

- Select the *Manual Dispense* option and press Select.
- Use the numeric keypad to enter the volume to be dispensed.

The manual dispense volume must be between the limits shown here.

5 mL burette	0.001 to 4.750 mL
10 mL burette	0.001 to 9.500 mL
25 mL burette	0.005 to 23.750 mL
50 mL burette	0.005 to 47.500 mL

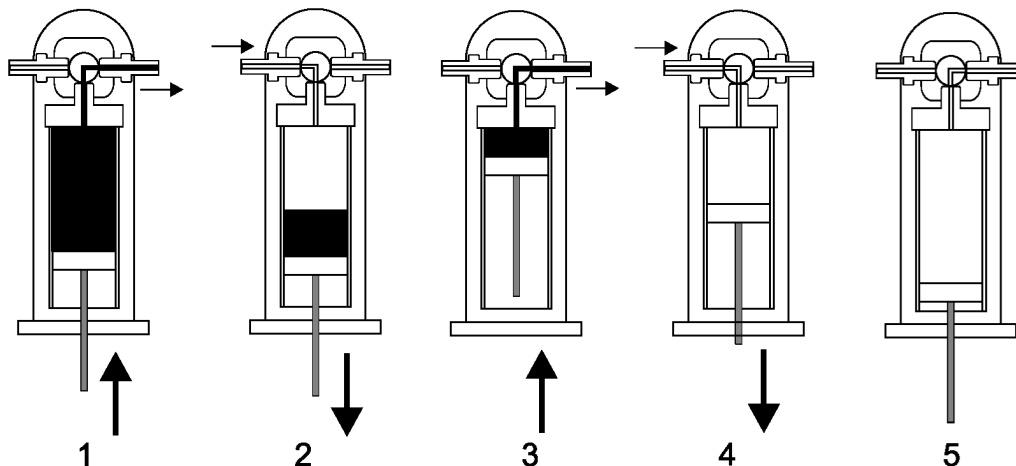
Manual Volume Dispense				
Enter the amount of volume to be dispensed.				
<div style="background-color: black; color: white; display: inline-block; padding: 5px 15px; border: 1px solid black;">1.000 mL</div>				
Current burette volume is 25 mL.				
Accept	Escape	Delete Digit		

Purging the Burette

This option allows the burette to be emptied before cleaning or storing the burette. The burette is flushed twice.

Note: Before starting this operation, remove the aspiration tube from the titrant bottle.

The figures below show the steps in a purge burette operation.

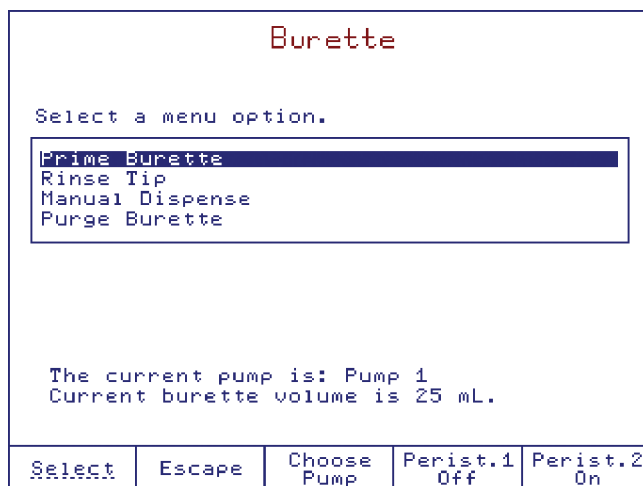


Peristaltic Pump

To manually control the peristaltic pump, press Burette from the main titration screen.

Perist.1
On or Perist.2
On Turns on the selected peristaltic pump.

Perist.1
Off or Perist.2
Off Turns off the selected peristaltic pump.



Note: If the peristaltic pump is not turned off, it will turn off automatically after 10 minutes.

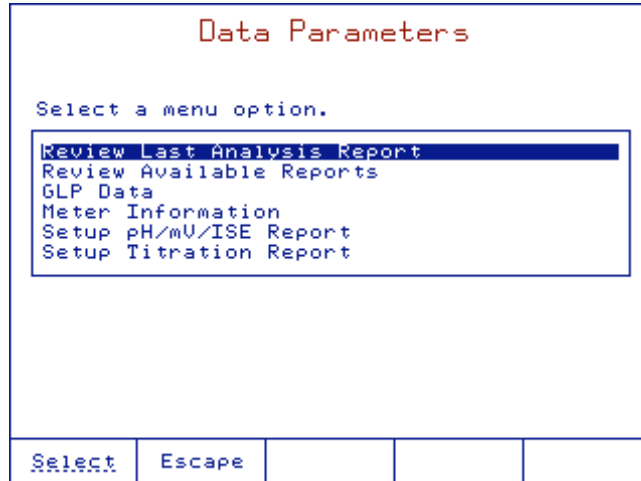
2.9.2. STIRRER

The stirrer can be turned on and off by pressing stir.

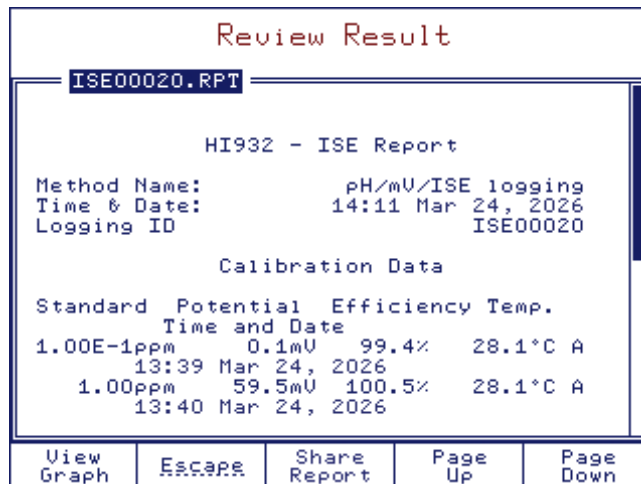
During the titration process, the stirring speed can be manually adjusted using the ▲ and ▼ keys.

2.9.3. RESULTS

From the **Data Parameters** screen, you can access the following options:



Reviewing Last Analysis Report



The report contains information based on the selections made in the **Setup Titration Report** and **Setup ISE / pH / mV Report** screen.


Use View Graph to review the graph.

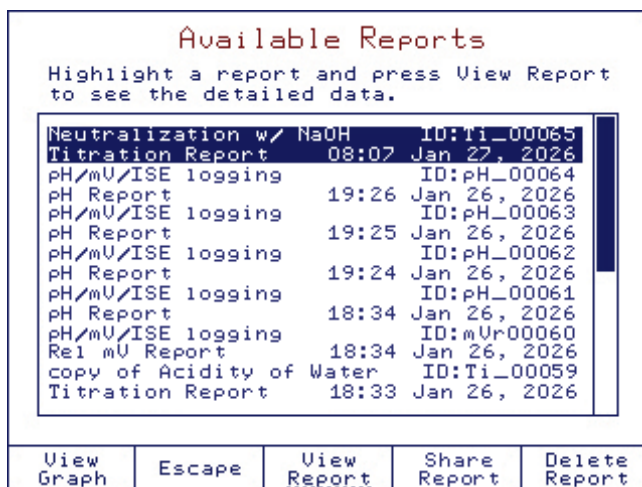
Use Escape to return to the previous screen.

Use Share Report to share the titration report.

Use Page Up and Page Down to scroll through the pages.

Reviewing Available Reports

Up to 100 reports can be saved on the titrator. To view one of the saved reports, highlight a report and then press .



The report contains only the information selected in the **Setup Titration Report** and **Setup pH / mV / ISE Report** screens during report configuration.

Use  to review the selected graph.

Use  to review the selected report.

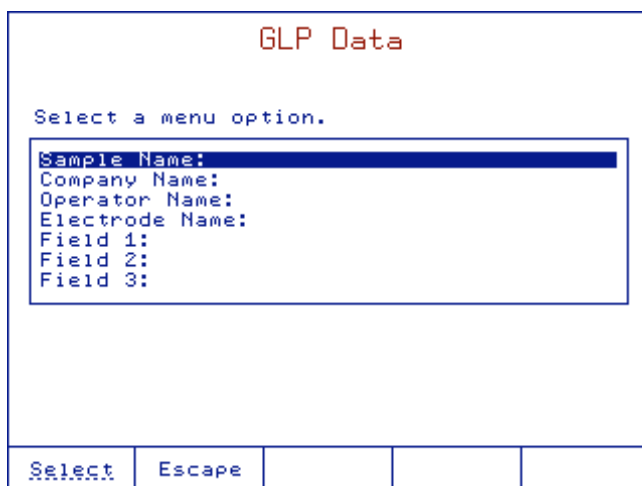
Use  to share the selected report.

Use  to delete the selected report.

Use  to return to the previous screen.

GLP Data

Option: Up to 20 characters



Sample Name Allows the sample name to be recorded in each report.

The sample name will increase by one, with each new titration or logging report, if the last character is a number.

Company Name Allows the company name to be recorded in each report.

Operator Name Allows the operator name to be recorded in each report.

Electrode Name Allows the electrode name to be recorded in each report.

Fields 1, 2, 3 Allows any additional information to be recorded in each report.

The fields must be selected from **Setup Titration Report** screen in order to be displayed in the titration report. See [Setting Up Titration Report](#) section for more information.

Meter Information

Displays titrator configuration data.

Meter Information				
METER DATA				
Meter Code: HI932				
Digital CH3 Probe Code: HI10480				
Digital CH4 Probe Code: HI36180				
Digital CH3 Probe Type: pH				
Digital CH4 Probe Type: ORP				
Stirrer Type: Overhead				
CALIBRATION DATA				
Analog CH1 Calibr. Date: Mar 22, 2026				
Analog CH2 Calibr. Date: Mar 10, 2026				
Digital CH3 Calibr. Date: Mar 22, 2026				
Digital CH4 Calibr. Date: Mar 10, 2026				
Escape	Share	Meter Calibr.	Serial Numbers	Software Versions

Meter Data

- Titrator
- Digital probes type and ordering code
- Stirrer type

Calibration dates

- Analog CH 1 and CH 2
- Digital CH 3 and CH 4

Meter Information				
SERIAL NUMBER				
Titrator Serial Number: 12133404404				
Analog CH1 Serial Number: 30134202202				
Analog CH2 Serial Number: 30000000000				
Digital CH3 Serial Number: 40023513513				
Digital CH4 Serial Number: 40024513513				
Pump 1 Serial Number: 70094513513				
Stirrer 1 Serial Number: 70091703703				
Escape	Share	Meter Calibr.	Serial Numbers	Software Versions

Serial numbers

- Titrator
- Analog CH 1 and CH 2
- Digital CH 3 and CH 4
- Pump 1
- Stirrer 1

Meter Information				
SOFTWARE VERSION				
Titrator Software Version: v2.2.2				
Base Board Software Version: v1.00				
Pump 1 Software Version: v1.00				
Stirrer 1 Software Version: v1.00				
Digital CH3 Software Version: v1.00				
Digital CH4 Software Version: v1.00				
Escape	Share	Meter Calibr.	Serial Numbers	Software Versions

Software version installed on:

- Titrator
- Base board
- Pump
- Stirrer
- Digital CH 3 and CH 4

Note: If more than one year elapsed from the calibration date of the Analog CH 1 or 2/ Digital CH 3 or 4 measurement inputs, the "Calibration Due" message will appear on the main screen. The measurement input needs to be recalibrated.

Use Share to share Meter Information.

Setting up pH / mV / ISE Report

Customize a unique report to record the pH, mV, and ISE measurements.

An asterisk means that it will be included in the report.

- Select key adds the highlighted information to the report.
- Unselect key removes the highlighted information from the report.
- Escape key returns to the Data Parameter Screen. Report is not updated.
- Save Report key updates the report with the selected items. Report previously saved will not be updated.
- Page Up Page Down keys scroll through the options.

Setting Up Titration Report

Customize a unique report to record the titration results. An asterisk means that it will be included in the titration report.

- Select key adds the highlighted information to the report.
- Unselect key removes the highlighted information from the report.
- Escape key returns to the Data Parameter Screen. Report is not updated.
- Save Report key updates the report with the selected items. Report previously saved will not be updated.
- Page Up Page Down keys scroll through the options.

2.10. MAINTENANCE & PERIPHERALS

The 25-mL burette included with the titrator exceeds ISO 8655 standard for accurate delivery of liquids by a motor-driven piston burette.

2.10.1. BURETTE MAINTENANCE

Burette Assembly

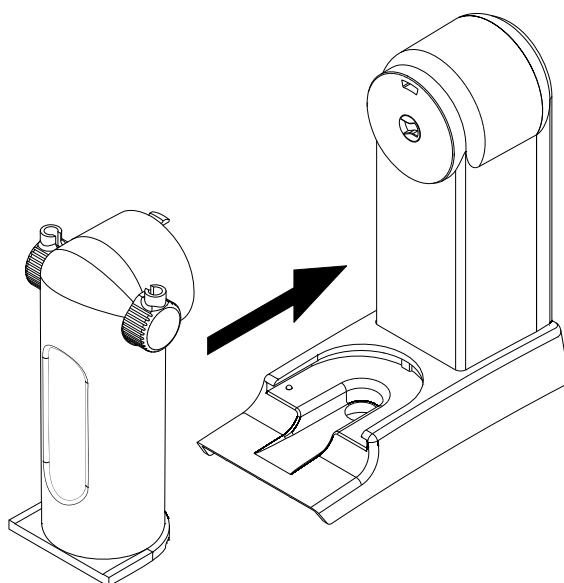
The burette is delivered with a 25-mL syringe inside and with all of the accessories mounted.

See [2.1. Setup](#) section for more information.

The burette assembly consists of a rigid housing which holds the glass syringe, a three-way valve, and titrant tubing.

Changing the Burette

- Slide the burette forward to remove.
- Slide the new burette into place.



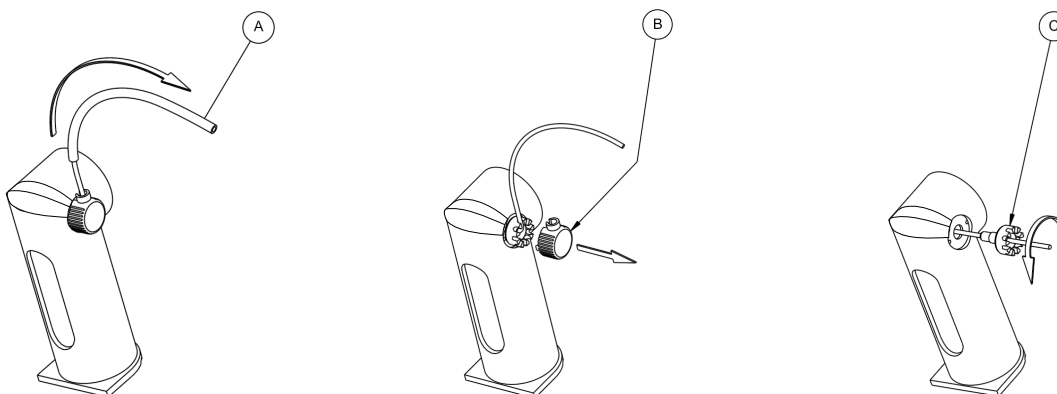
Disassembling the Burette

The aspiration and the dispensing tubes have fittings and tube protectors.

The aspiration tube is mounted on the left side and the dispensing tube is mounted on the right side of the burette.

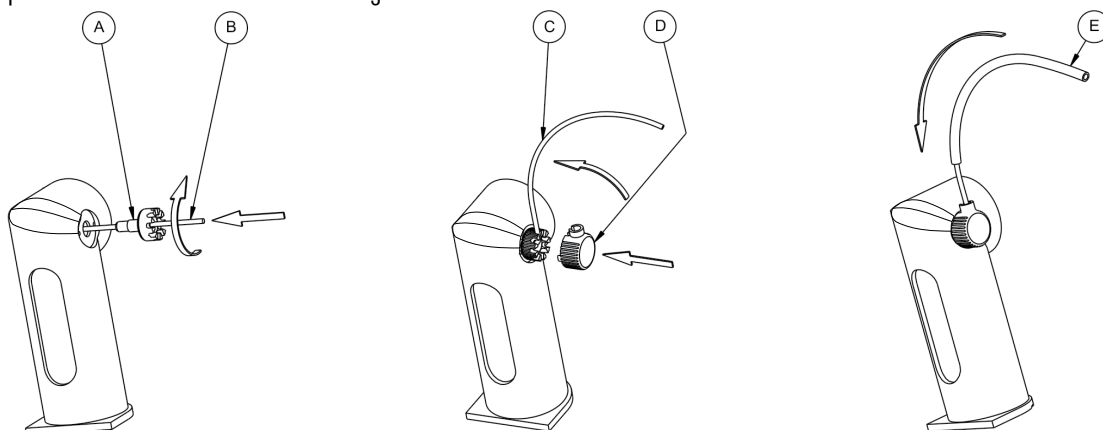
To remove the tubing:

1. Remove the blue tube protector (A) by sliding it off the clear titrant tubing.
2. Remove the tube lock (B) from the burette holder.
3. Turn the fitting (C) counterclockwise to remove it from the burette holder.
4. Slide the clear titrant tubing through the fitting.



Assembling the Burette

1. Insert the flat-shaped end of the titrant tubing (B) into the valve outlet (A) and screw the fitting clockwise to tighten. The highest of the nine cuts should be vertical in the final position.
2. Bend the tube up into the vertical position to enter the highest cut of the fitting (C).
3. Replace the tube lock fitting (D).
4. Replace the blue tube protector (E) by sliding it over the clear titrant tubing. The protector will sit in the tube lock fitting.



Cleaning the Burette

To clean the burette, follow these steps:

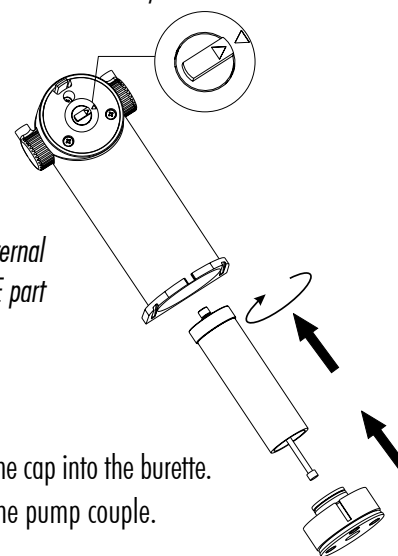
1. If the burette is filled with titrant, remove the aspiration tube from the titrant bottle and purge burette. See [Purging the Burette](#) section for more information.
2. Insert the aspiration tube into cleaning solution, deionized water or titrant solvent.
3. Go through two cycles of filling and emptying the burette. See [Purging the Burette](#) section for more information.
4. During second cycle, remove the aspiration tube from the cleaning solution, deionized water or solvent and allow the air to replace the liquid in the burette. This will clean the aspiration tube.

If this simple cleaning procedure is not adequate, continue with these steps:

5. Remove the burette assembly from the pump.
6. Remove the dispensing and aspiration tubes.
7. Clean them separately or insert new ones.
8. Use the burette removal tool to remove the protective cap from the bottom of the burette assembly.
9. Remove the syringe from the burette assembly by unscrewing it with your fingers.
10. Extract the piston from the syringe.
11. Clean both the piston and the syringe with appropriate cleaning solution. Rinse with deionized water.
12. Remove the excess liquid.

Warning: Avoid contacting the titrant with bare hands. Avoid spilling titrant. Clean the external side of the syringe and piston to remove aggressive chemicals. Do not touch the white PTFE part of the piston or internal walls of the burette with bare hands or greasy materials.

13. Reinsert the piston into the syringe.
14. Reinsert the syringe by screwing it in the valve with your fingers.
15. Reinsert the protective cap to the bottom of the burette assembly. Carefully position the cap into the burette.
16. Slide the burette into the burette stand. Notice the position of the piston shaft to the pump couple.
17. Priming the burette three times with new titrant is recommended.



Burette Preparation (Titrant Filling)

Before starting a titration, the burette must be properly filled with titrant in order to obtain an accurate and repeatable result.

To fill the burette:

1. If necessary, clean the burette and make sure it is empty.
2. From the main screen press .
3. Highlight *Prime Burette* option and press .
4. Enter the number of times the burette needs to be rinsed.

Note: *Multiple rinses (minimum three) are recommend to allow air bubbles to be evacuated.*

5. Press .

To avoid the presence of the air bubbles inside the burette, make sure to have a continuous liquid flow inside the burette.

A little air just above the liquid level at the first filling is normal. The next filling will evacuate all of the air; no air will be left in the valve.

Sometimes during this process, slight finger tapping on the tubes helps remove any residual air bubbles.

If air bubbles are still present:

1. Remove the aspiration tube from the titrant bottle.
2. Repeat burette preparation procedure.
3. If this is not successful, clean the burette again.

2.10.2. HI932933 NETWORK BOX

The HI932933 is a secure network device designed for the Hanna Instruments® HI932 Automatic Potentiometric Titrator running software version 2.2.2 and higher.

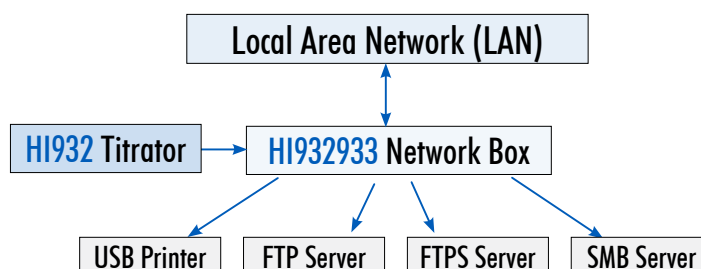
The HI932933 enables secure network (Ethernet) connection for data exchange and adds USB printer support, allowing direct printing of reports, methods, and data logs.

The device supports software updates for security patches and new features and uses SMB and FTP/FTPS protocols for secure data transfer to PC and network servers.

The HI932 Automatic Titrator allows network box configuration directly through its interface, without additional software.

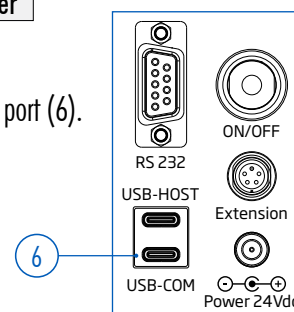
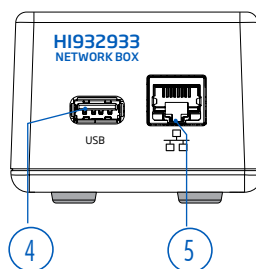
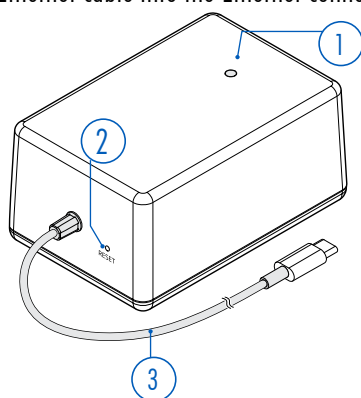
Note: For details see [2.3.13. Network Box Setup](#) » [Update Network Box Software](#).

Printing and File Sharing Environment Workflow



Network Box Connections

- Locate the rear panel of HI932 Titrator and plug the USB-C end (3) into the titrator's USB-COM port (6).
- Plug the printer directly into the USB port (4) on the network box.
- Plug the Ethernet cable into the Ethernet connection port (5).



1. LED
2. Reset button
3. Attached USB-C cable
4. USB port for printer
5. Ethernet connection port

Network Box LED Notifications

When the titrator and the plugged-in network box are connected to power, the titrator enters system configuration mode.

The network box is automatically detected and recognized.

The LED on the network box (1) turns on, indicating active network communication.

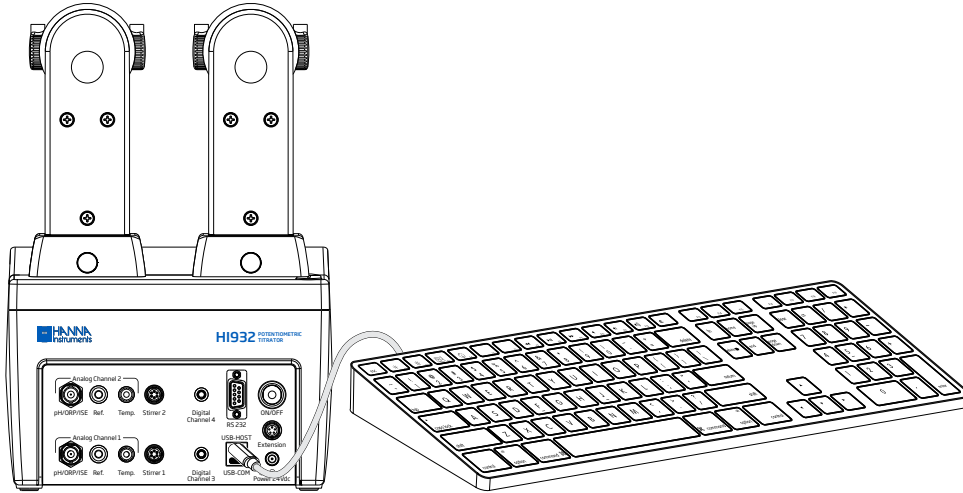
LED	Behavior	Operational Status	Description
○	Static	Not connected	The network box is booting up.
●	Blinking	Active	Configuration synchronization between titrator and network box.
●	Blinking	Active	Once the file is loaded or a job sent, the LED turns blue, indicating active processing.
●	Static	Ready	The box is initialized and waiting for data.
●	Static	Active	Firmware update in progress.
●	Static	Error/Timeout/Blocked	Error

2.10.3. OTHER PERIPHERALS

Warning: Connection or disconnection of POWER, PUMP ASSEMBLY, NETWORK BOX, RS232 INTERFACE, or AUTOSAMPLER must only be done when titrator and external devices are turned off.

Connecting an External PC Keyboard

This connection allows the use of an external keyboard with USB C connection in addition to the titrator's keypad. Connect to the USB-C port labelled **USB HOST**.



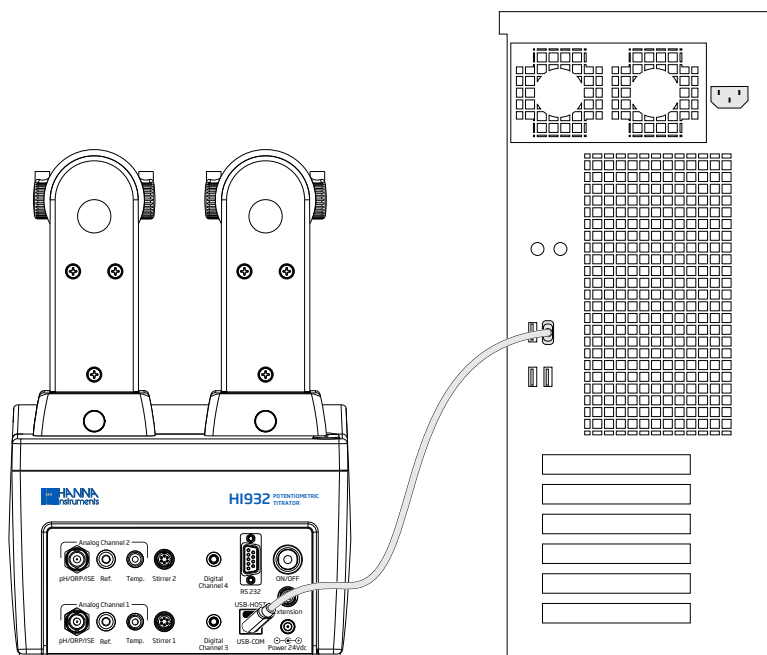
The correspondence between the titrator's keypad and the United States 101-type external keyboard is detailed below.

External PC Keyboard (United States 101)	Titrator Keypad
Function key F-1	
Function key F-2	
Function key F-3	
Function key F-4	
Function key F-5	Option key 1 (from left to right)
Function key F-6	Option key 2 (from left to right)
Function key F-7	Option key 3 (from left to right)
Function key F-8	Option key 4 (from left to right)
Function key F-9	Option key 5 (from left to right)
Function key F-10	
Arrow key: Up	
Arrow key: Down	
Arrow key: Left	
Arrow key: Right	
Page Up	
Page Down	
Numeric keys: 0 to 9	
Enter	
Alphanumeric keys	Allow alphanumeric entries

Connecting to a Computer

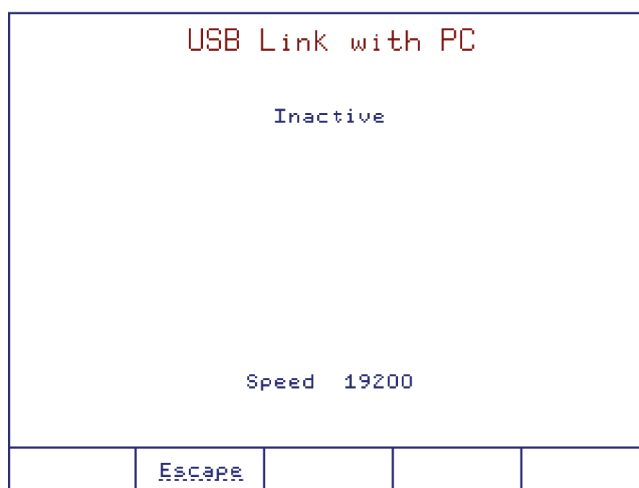
The titrator can be connected to a computer using a USB cable.

HI900 PC application needs to be installed on the PC.



To connect the PC to the titrator:

1. Connect the cable to the USB-C port labelled **USB COM** on the rear panel of the titrator.
2. Connect the cable to the USB port on the PC.



The **HI900** PC application allows the transfer of methods and reports between the titrator and PC. See [2.3.12. USB Link with PC](#) section for more information.

2.11. AUTOSAMPLER

2.11.1. START UP

Once the instrument is assembled and installed, follow the steps below to start the titrator and access the autosampler.


1. Use supplied power adapter to connect the titrator to a power outlet.
2. Use [HI920-933](#) communication cable to connect the autosampler to the titrator.
3. Press the power switch located on the back of the Autosampler to turn it on.
4. Press the power switch located on the back of the titrator to turn it on.
5. Wait for the titrator initialization process to complete.
6. When prompted, press device to enter the autosampler interface.

The autosampler information screen will be displayed.

If the autosampler has not been detected, an **X** will appear over the autosampler symbol located in the top right corner.

Note: The autosampler can be accessed at any time from the titrator's main screen by pressing **DEVICE**.



HI 922 Autosampler 

Autosampler


Serial Number: 21142001

Software Version: v1.0

Tray Type: 16 beakers


Status: Not Connected

Sequence Name: Default Sequence

Please press Enter ... 

Select Sequence		
-----------------	--	--

The sample table screen will be displayed.

18:40:32 Mar 06, 2026 


Default Sample Sequence

Last Seq.: TRAY0045, 17:29 Feb 30, 2026

#	Name	Size[g]	Result
1	----		
2	----		
3	----		
4	----		
5	----		
6	----		
7	----		
8	----		
9	----		
10	----		
11	----		
12	----		

Add Sample		AutoSmp. Setup
------------	--	----------------

Press  to view the autosampler's main screen.

18:40:17 Mar 06, 2026 

Default Sample Sequence




Method: 0.1N Sodium Hydroxide

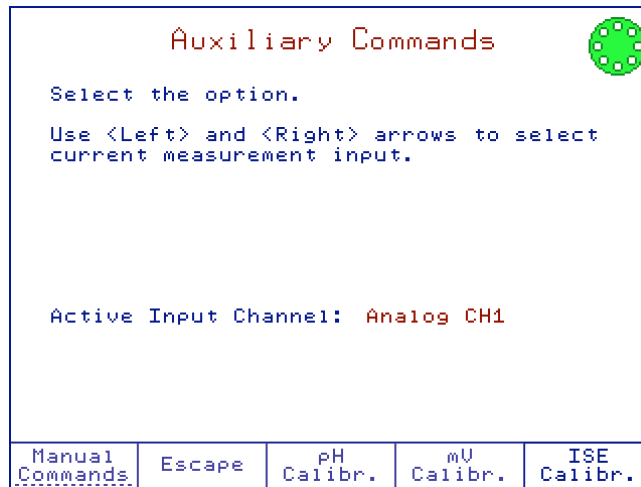
Analog CH1

#	Name	Size[g]	Result
1	----		
2	----		
3	----		
4	----		
5	----		
6	----		

AutoSmp. Options	Select Sequence	Sequence Options	Aux Commands	Sample Table
------------------	-----------------	------------------	--------------	--------------

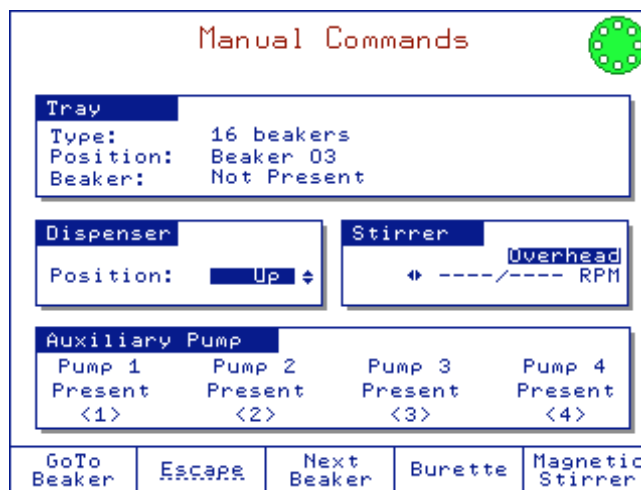
2.11.2. AUXILIARY COMMANDS

- Press  to access the auxiliary commands menu from the main screen.
The auxiliary commands menu allows electrode calibration, running the pumps, or moving the tray.
- Use the  and  keys to select the measurement input to be used for calibration.



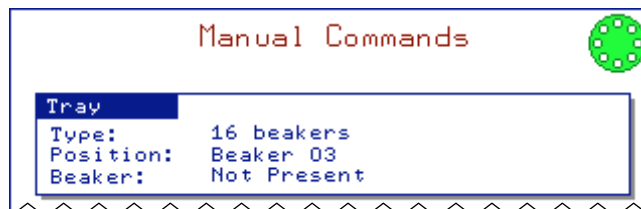
2.11.2.1. Manual Commands

The manual commands screen is used to manually operate the autosampler dispenser, beaker position, auxiliary pumps, burette and stirrers.



Tray

To move the tray, use   or the  and  arrow keys on the autosampler keypad.



Type Is the tray size currently detected by the autosampler. The tray size can be manually selected, if necessary.

See [2.11.3.6. Tray Type](#) section for more information.

Position Refers to the beaker located under the dispenser.

Beaker Indicates beaker's presence (present or not) under the dispenser.

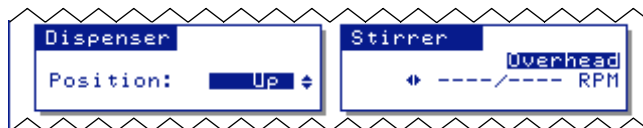
Beaker detection can be disabled, if necessary.

See [2.11.3.7. Beaker Detection](#) section for more information.

Dispenser

The position (up or down) of the dispensing head will be displayed.

To move the dispenser use the \triangle and ∇ keys on the autosampler or titrator keypad.



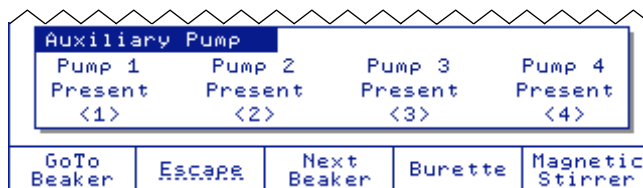
Stirrer

- Use $\left\{ \begin{array}{c} \text{Magnetic} \\ \text{Stirrer} \end{array} \right\}$ or $\left\{ \begin{array}{c} \text{Overhead} \\ \text{Stirrer} \end{array} \right\}$ to toggle between the stirrer type.
- Press $\left\{ \begin{array}{c} \text{stir} \end{array} \right\}$ on the titrator keypad to turn on the stirrer.
- When active, use the $\left\{ \begin{array}{c} < & > \end{array} \right\}$ keys on the titrator keypad to change the stir speed.



Auxiliary Pumps

To run an active pump, press and hold the corresponding number key on the autosampler keypad or titrator keypad (e.g. press numeric key 1 for auxiliary pump 1, numeric key 2 for auxiliary pump 2).

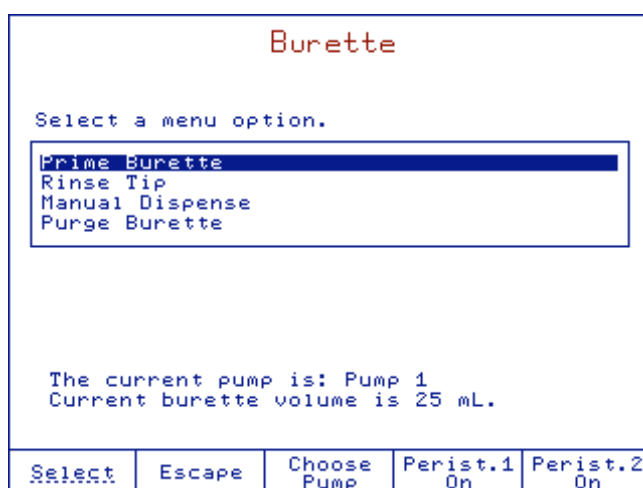


Burette


To access the Burette screen, press $\left\{ \begin{array}{c} \text{Burette} \end{array} \right\}$ from the manual commands screen.

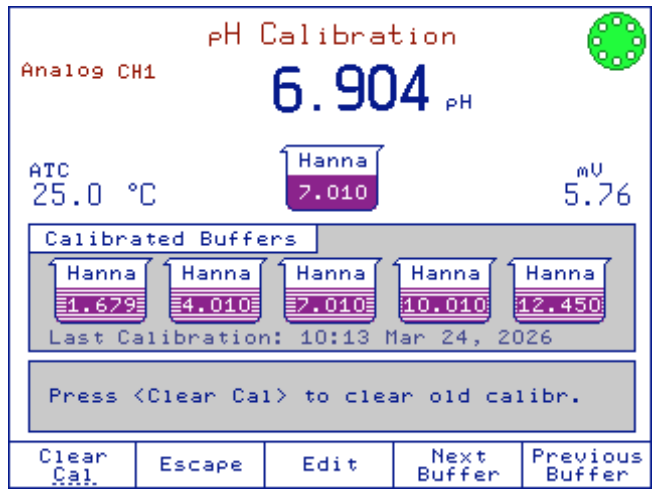
Highlight the desired option then press $\left\{ \begin{array}{c} \text{Select} \end{array} \right\}$.

See 2.9.1. [Burette](#) section for more information.




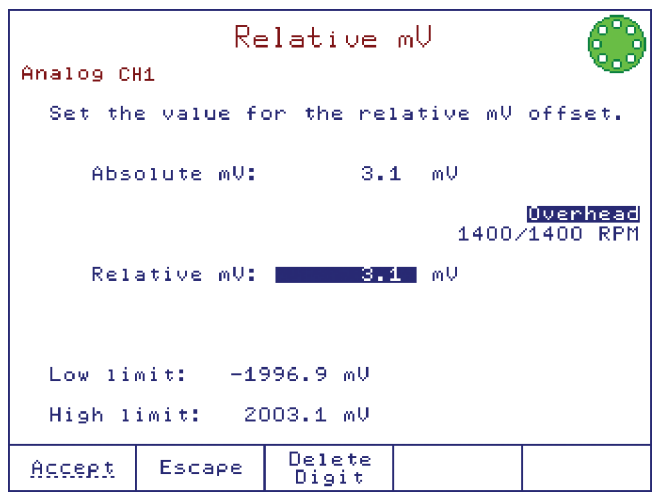
2.11.2.2. pH Calibration

From the Auxiliary Commands screen, press  to view pH calibration screen.
See [2.6.3. pH Calibration](#) section for more information.



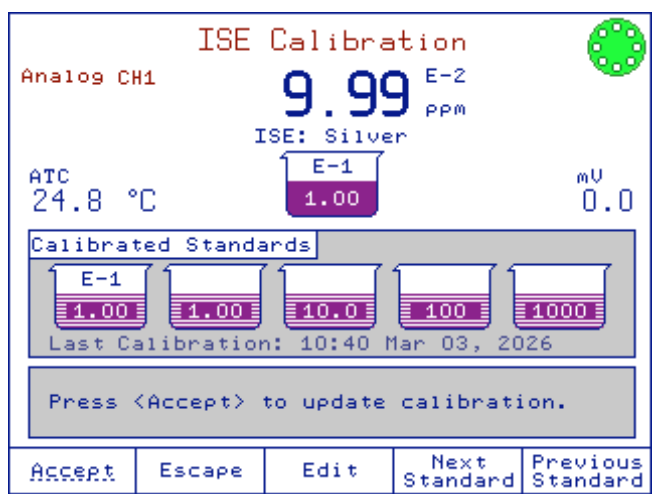
2.11.2.3. Relative mV Calibration

Press  to view mV calibration screen.
See [2.7.3. Relative mV Calibration](#) section for more information.




2.11.2.4. ISE Calibration

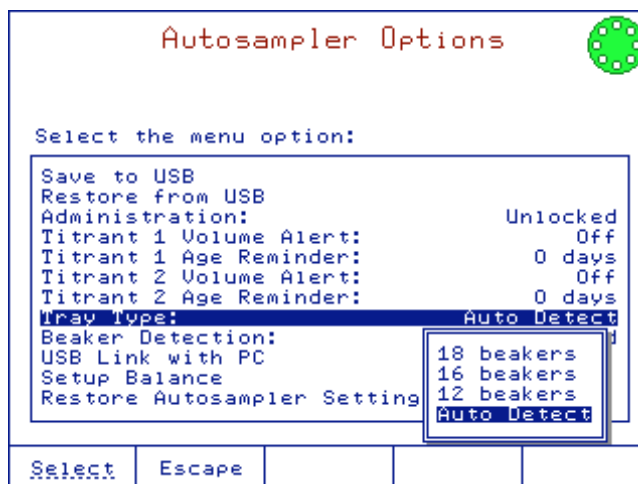
Press  to view ISE calibration screen.
See [2.8.3. ISE Calibration](#) section for more information.



2.11.3. AUTOSAMPLER OPTIONS

The Autosampler Options screen gives access to options that are not directly related to the autosampler sequences.

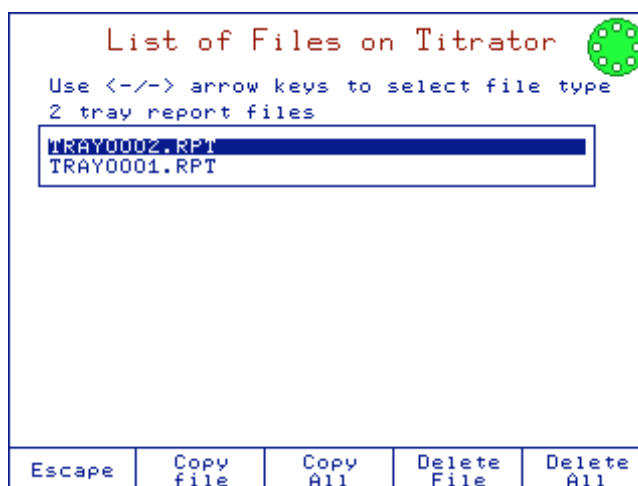
To access this screen, press  from the autosampler main screen.



2.11.3.1. Save to USB





This option allows the user to save files from titrator to a USB storage device.

Note: Autosampler report files contain the individual titration reports for all samples run on that tray.








From the autosampler options, the available file types are given here.

Standard method HIXXXYY.MTD (e.g. H11001EN.MTD, H11004EN.MTD)
User-defined method USERXXX.MTD (e.g. USER0001.MTD)
Sequence SEQXXX.MTD (e.g. SEQ0001.MTD)
Autosampler report TRAYXXX.RPT (e.g. TRAY0001.RPT)

- Use the  and  keys to select the file type.
The number of files and the file name on the titrator will be displayed.
- Use the  and  keys to scroll through the list.

The option keys allow the following operations:

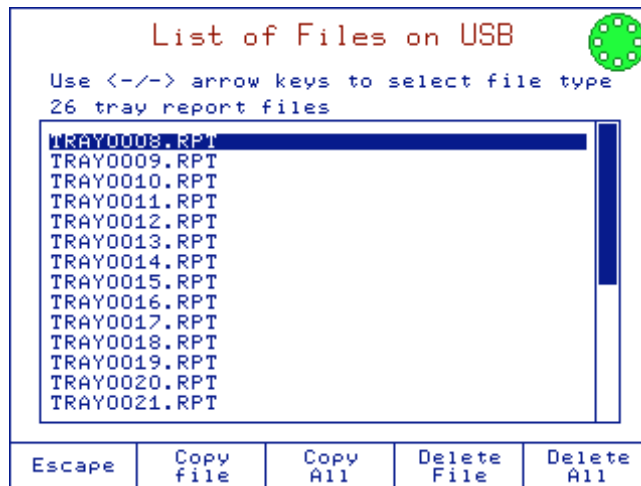
-  Deletes the highlighted file.
-  Deletes all currently displayed files.
-  Copies the highlighted file from the titrator to a USB storage device.
-  Copies all currently displayed files from the titrator to a USB storage device.
-  Returns to the **Autosampler Options** screen.

The status of the transfer (“Successful” or “Unsuccessful”) and the file name of the currently processed file are displayed during copying or deleting.

Note: The saved files will be stored on the USB storage device in the **HI932** Folder, as follows:

Methods USB Drive:\HI932R\Methods*.mtd
Sequences USB Drive:\HI932R\Sequence*.mtd
Reports USB Drive:\HI932R\ASReport*.rpt

2.11.3.2. Restore From USB



This screen allows the user to transfer files from a USB storage device to the titrator. The file types that can be transferred are:






Standard Method HIXXXYY.MTD (e.g. HI1001EN.MTD, HI1004EN.MTD)
User-defined Method USERXXX.MTD (e.g. USER0001.MTD)
Sequence SEQXXX.MTD (e.g. SEQ0001.MTD)
Autosampler Report TRAYXXX.RPT (e.g. TRAY0001.RPT)

Note: Autosampler report files contain the individual titration reports for all samples run on that tray.

Use the < and > keys to select the file type. The number of files and the file name on the titrator will be displayed.

Use the ^ and v keys to scroll through the list.

The option keys allow the following operations:

-  Deletes the highlighted file.
-  Deletes all currently displayed files.
-  Copies the highlighted file from the titrator to a USB storage device.
-  Copies all currently displayed files from the titrator to a USB storage device.
-  Returns to the **Autosampler Options** screen.

Note: The saved files will be stored on the USB storage device in the **HI932R** Folder, as follows:

Methods USB Drive:\HI932R\Methods*.mtd
Sequences USB Drive:\HI932R\Sequence*.mtd
Reports USB Drive:\HI932R\ASReport*.rpt

2.11.3.3. Administration

A 4-digit numeric PIN can be set to prevent unauthorized changes from being made.

See [2.3.3. Administration](#) section for more information.

Titrator Administration				
Titrator is LOCKED.				
<div style="border: 1px solid black; padding: 5px; width: fit-content; margin: auto;"> Unlock Titrator Enter PIN: ***- </div>				
ACCEPT	Escape	Delete Digit		

2.11.3.4. Titrant Volume Alert

This screen allows a programmable reminder to appear when the titrant reservoir is below 100 mL. The titrant volume will decrease as the titrant is used.

See [2.3.10. Total Volume Alert](#) section for more information.

Titrant 1 Volume Alert				
Enter the amount of titrant available to the titration/reagent system from its reservoir. The mLs will decrease as the titrant/reagent is depleted.				
<div style="border: 1px solid black; padding: 2px; width: fit-content; margin: auto;"> 0.0 mL </div>				
A reminder will appear when less than 100 mLs of titrant volume is left.				
ACCEPT	Escape	Delete Digit		Off

2.11.3.5. Titrant Age Reminder

A programmable reminder will appear when it is time to verify the titrant concentration or to change the titrant.

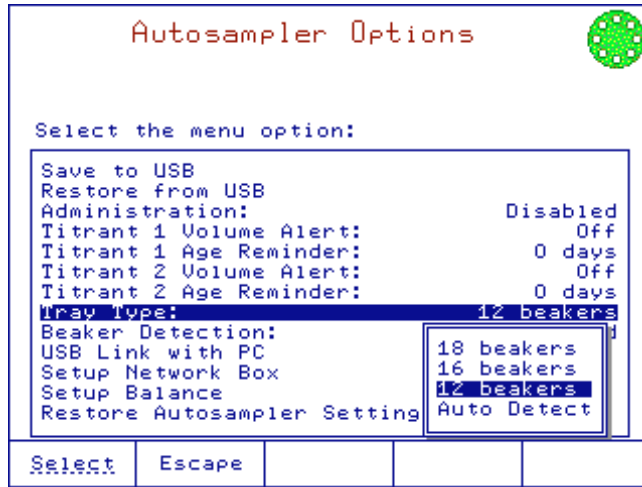
See [2.3.11. Titrant Age Reminder](#) section for more information.

Titrant 1 Age Reminder				
Enter the number of days to pass since the last Titr. Vol. updating or the last Start pressing, whereafter the reminder appears.				
<div style="border: 1px solid black; padding: 2px; width: fit-content; margin: auto;"> 5 days </div>				
The range is from 0 to 31 days.				
Start	Escape	Delete Digit		Off

2.11.3.6. Tray Type

Option: Auto Detect, 18 beakers, 16 beakers, 12 beakers

The autosampler trays have a built in RFID tag that transmits the tray type and serial number directly to the titrator. The tray type can also be selected manually.

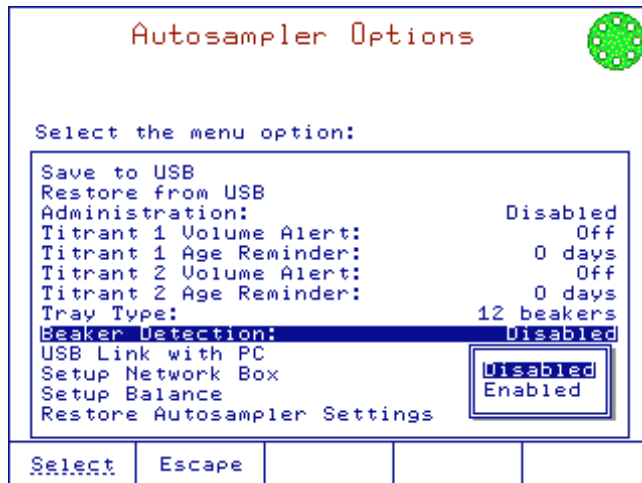


2.11.3.7. Beaker Detection

Option: Enabled or Disabled

The autosampler detects the presence of a beaker when it is under the dispenser. This prevents titrations from occurring when no beaker is present.

WARNING: If this feature is disabled, the autosampler will titrate in any spot on the tray a sample has been entered. Please ensure all beakers are correctly positioned before starting the sequence. Otherwise, serious injury could result.

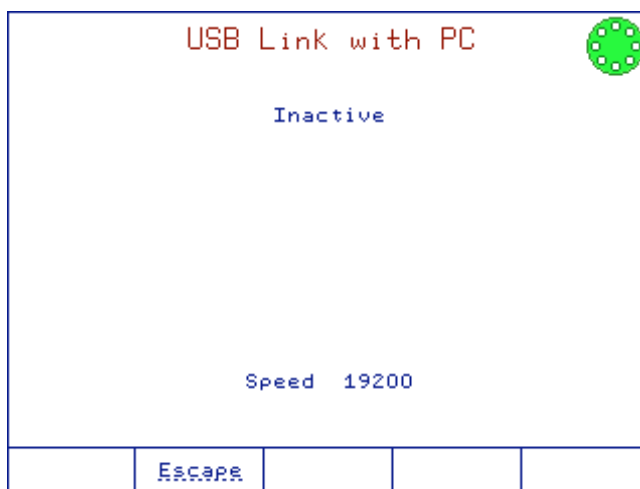


2.11.3.8. USB Link with PC

In order to use this feature, the USB cable needs to connect the titrator to the PC.

Make sure that the **HI900** (or **HI900900**) PC application (version number 4.00) is running on the PC.

See [2.3.12. USB Link with PC](#) section for more information.



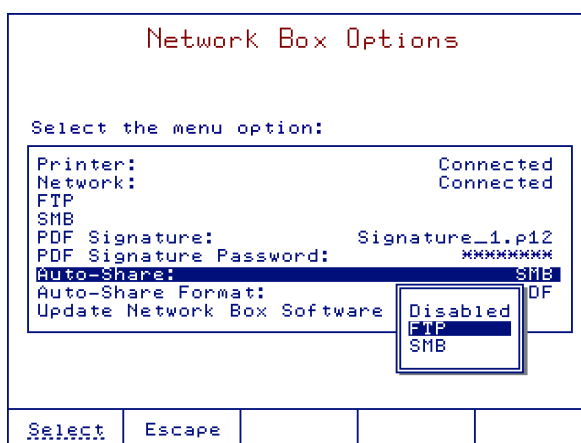
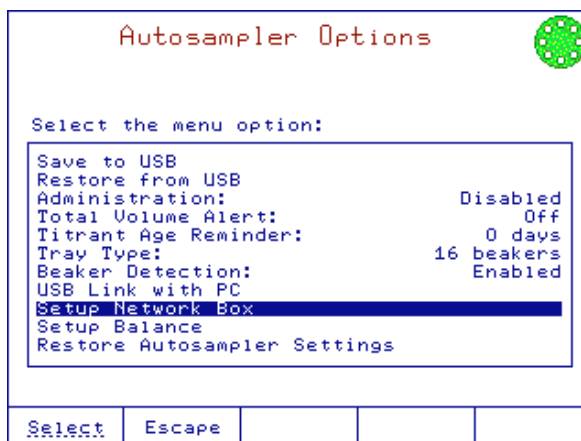
2.11.3.9. Network Box Setup

Option: Printer, Network, FTP, SMB, PDF Signature, PDF Signature Password, Auto-Share, Auto-Share Format, Update Network Box Software

- Connect **HI932933** Network Box to the **HI932** Titrator.
- Use the \triangle and ∇ keys to highlight selected item and press Setup.

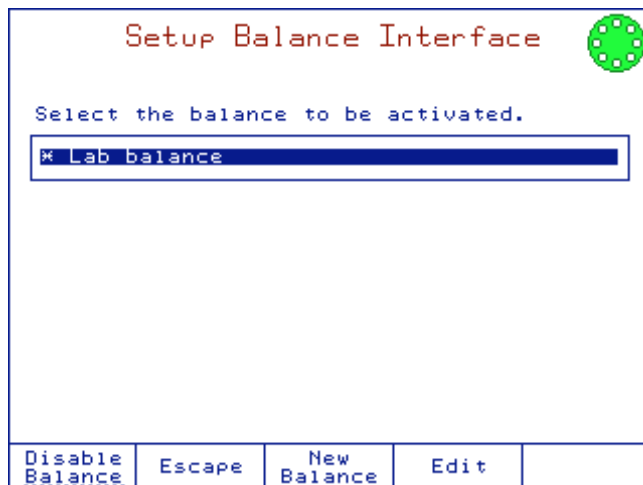
See [2.3.13. Network Box Setup](#) section for detailed information:

- » [Update Network Box Software](#) for steps on software updates, security patches or new features.
- » [Auto-Share](#) section for detailed information.



2.11.3.10. Setting up the Analytical Balance

This screen allows the users to connect an analytical balance for automatic acquisition of sample mass prior to titration or standardization. See [2.3.13. Network Box Setup » Auto-Share](#) section for more information.



2.11.3.11. Restore Autosampler Settings

This option restores the manufacturer settings for the autosampler interface only!

Note: This will delete all user-created sequences, tray reports, etc.



2.11.4. AUTOSAMPLER SEQUENCES


All of the parameters required to complete an analysis on the autosampler are grouped into a sequence.

A default sequence is provided. The default sequence is used as a starting point for creating user-defined sequences.

Up to 30 sequences can be created and stored on the titrator.

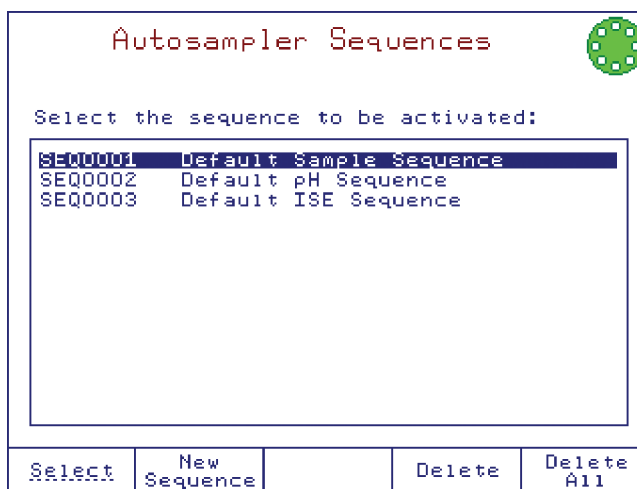
User-defined sequences allow the user to customize reagent additions and rinsing cycles to suit specific applications. New sequences are created in the select sequence screen.


Selecting a Sequence

To select a sequence, press  from the main screen.

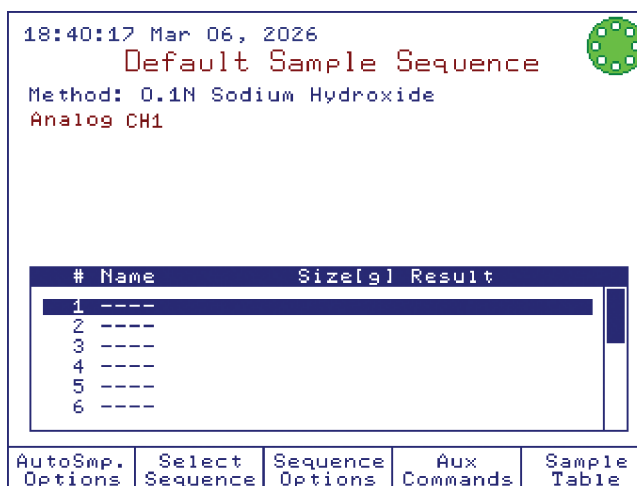
A list of available sequences will be displayed.

In the **Autosampler Sequence** screen, the list of all available sequences is displayed.



To select a sequence, highlight the sequence and press .

The name of the selected sequence will be displayed on the main screen.








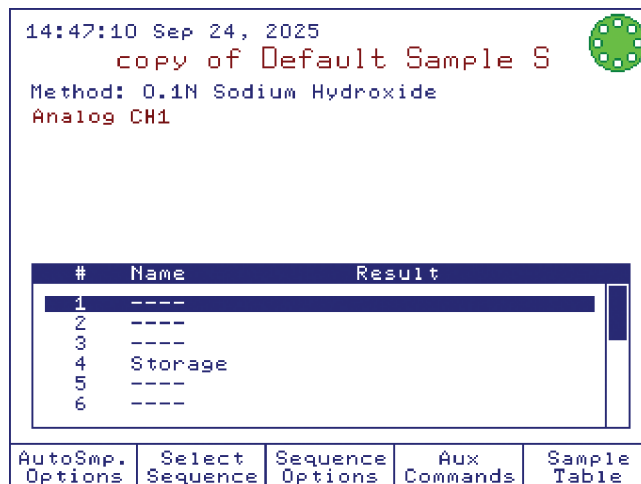
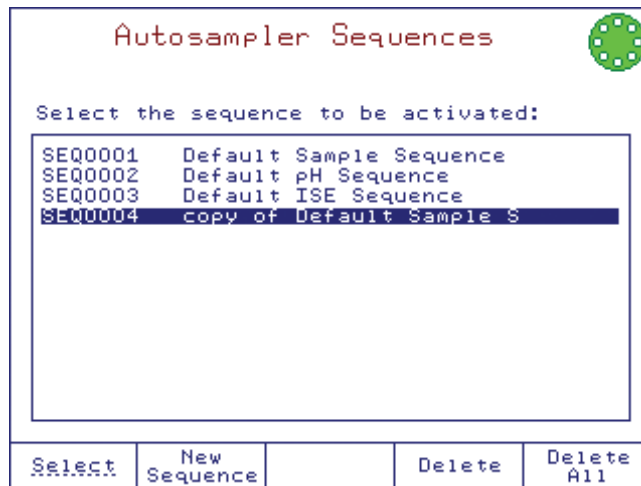
Creating a Sequence

Sequences are developed by the users in accordance with the analysis requirements.


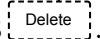


All sequence parameters can be modified by the user.

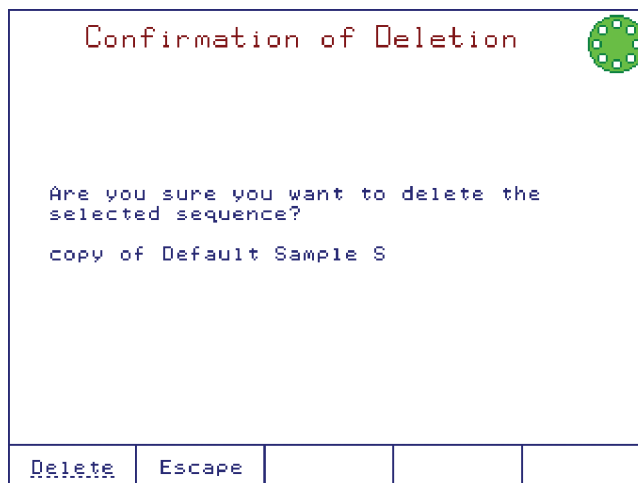
To create a new sequence, start from the default sequence or a previously created sequence and follow these steps given here.

1. Press  from the main screen.
2. Use the  and  keys to highlight an existing sequence from the list.
3. Press . A new sequence will be generated.
4. Press  to activate the new sequence.



Deleting a Sequence

- To remove a sequence, press  from the main screen.
 - Highlight the sequence to be deleted.
 - Next, press .
- A screen will appear in order to confirm the deletion.
- Press  again to confirm or press  to cancel the operation.







Sequence Types

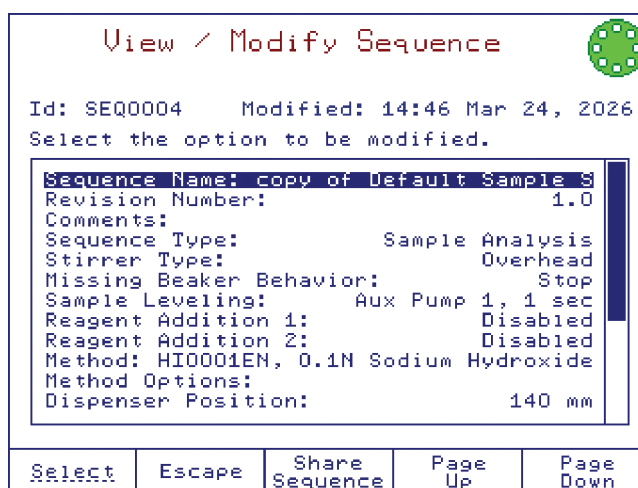
The autosampler can run three sequence types:

- Sample analysis
- pH calibration
- ISE calibration

2.11.5. SAMPLE ANALYSIS SEQUENCE

2.11.5.1. View/Modify Sequence

- To modify the sequence options, press  from the main screen.
A list of all the parameters for the selected sequence will be displayed.
- Use the  and  keys to highlight the option to be modified.
- Next, press .



- To exit the **View/Modify Sequence** screen, press .

Opt to save the modifications or to discard them.

Saving Sequence

Select a menu option.

Save Sequence
Exit Without Saving Sequence

"Escape" - exit without saving sequence.

Select	Escape			
--------	--------	--	--	--

Sequence Name

Option: Up to 24 characters

Sequence Name

Select the highlighted letter by using the arrow keys then press "Enter".
Select the empty field for a space.
Press Accept to save the entered text.

█	A	B	C	D	E	F	G	H	I	J	K	L
M	N	O	P	Q	R	S	T	U	V	W	X	Y
Z	a	b	c	d	e	f	g	h	i	j	k	l
m	n	o	p	q	r	s	t	u	v	w	x	y
z	À	Á	Â	Ã	Ä	Å	Æ	Ç	È	É	Ê	Ë
Ì	Í	Î	Ï	Ñ	Ò	Ó	Ô	Õ	Ö	×	÷	¸
Ù	Ú	Û	Ü	Ý	Þ	ß	à	á	â	ã	ä	å
æ	ç	è	é	ê	ë	ì	í	î	ï	ñ	ò	ó
ô	õ	ö	÷	¸	¸	¸	¸	¸	¸	¸	¸	¸
0	1	2	3	4	5	6	7	8	9	×	÷	,
?	!	()	[]	<	>	=	/	+	-	

█ Salt Analysis █

Accept	Escape	Delete Letter	Cursor Left	Cursor Right
--------	--------	---------------	-------------	--------------

Revision Number

Option: Up to 3 characters

Revision Number

Select the highlighted letter by using the arrow keys then press "Enter".
Select the empty field for a space.
Press Accept to save the entered text.

█	A	B	C	D	E	F	G	H	I	J	K	L
M	N	O	P	Q	R	S	T	U	V	W	X	Y
Z	a	b	c	d	e	f	g	h	i	j	k	l
m	n	o	p	q	r	s	t	u	v	w	x	y
z	À	Á	Â	Ã	Ä	Å	Æ	Ç	È	É	Ê	Ë
Ì	Í	Î	Ï	Ñ	Ò	Ó	Ô	Õ	Ö	×	÷	¸
Ù	Ú	Û	Ü	Ý	Þ	ß	à	á	â	ã	ä	å
æ	ç	è	é	ê	ë	ì	í	î	ï	ñ	ò	ó
ô	õ	ö	÷	¸	¸	¸	¸	¸	¸	¸	¸	¸
0	1	2	3	4	5	6	7	8	9	×	÷	,
?	!	()	[]	<	>	=	/	+	-	

█ 1.0 █

Accept	Escape	Delete Letter	Cursor Left	Cursor Right
--------	--------	---------------	-------------	--------------

Comments

Option: Up to 20 characters

Comments				
Select the highlighted letter by using the arrow keys then press "Enter". Select the empty field for a space. Press Accept to save the entered text.				
<input checked="" type="checkbox"/>	A	B	C	D
	M	N	O	P
	Z	a	b	c
	m	n	o	p
	z	À	Á	Â
	ò	ó	ô	õ
	ä	ç	è	é
	ù	ú	û	ü
	0	1	2	3
	?	!	()

Accept	Escape	Delete Letter	Cursor Left	Cursor Right
--------	--------	---------------	-------------	--------------

Sequence Type

Option: Sample Analysis, pH Calibration, ISE Calibration

View / Modify Sequence				
Id: SEQ0004 Modified: 14:46 Mar 24, 2026				
Select the option to be modified.				
Sequence Name: copy of Default Sample S				
Revision Number: 1.0				
Comments:				
Sequence Type: Sample Analysis				
Stirrer Type: d				
Missing Beaker Behavior: Sample Analysis p				
Sample Leveling: pH Calibration d				
Reagent Addition 1: ISE Calibration d				
Reagent Addition 2: d				
Method: HI0001EN, 0.1N Sodium Hydroxide				
Method Options:				
Dispenser Position: 140 mm				

Select	Escape			
--------	--------	--	--	--

Stirrer Type

Option: Overhead or Magnetic

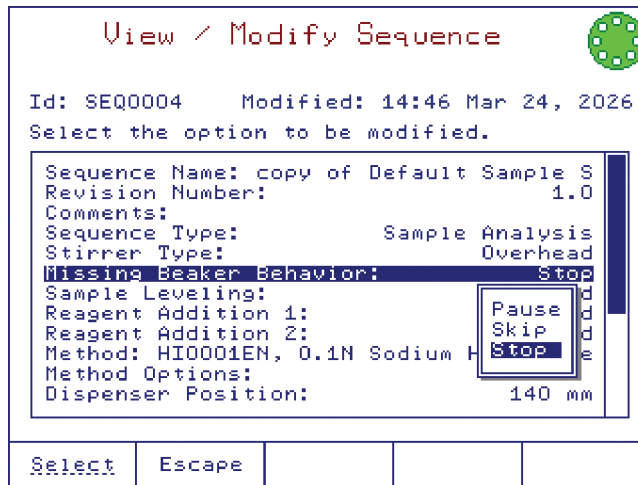
View / Modify Sequence				
Id: SEQ0004 Modified: 14:46 Mar 24, 2026				
Select the option to be modified.				
Sequence Name: copy of Default Sample S				
Revision Number: 1.0				
Comments:				
Sequence Type: Sample Analysis				
Stirrer Type: Overhead				
Missing Beaker Behavior: p				
Sample Leveling: Overhead d				
Reagent Addition 1: Magnetic d				
Reagent Addition 2: d				
Method: HI0001EN, 0.1N Sodium Hydroxide				
Method Options:				
Dispenser Position: 140 mm				

Select	Escape			
--------	--------	--	--	--

Missing Beaker Behavior

Option: Pause, Skip, Stop

Select type of behaviour when beaker detection is enabled and no beaker is detected.



Pause The autosampler will pause the sequence at the current beaker and wait for the user before continuing the analysis.

Skip The autosampler will automatically move to the next available sample.

Stop All operations will stop and the analysis will be stopped.

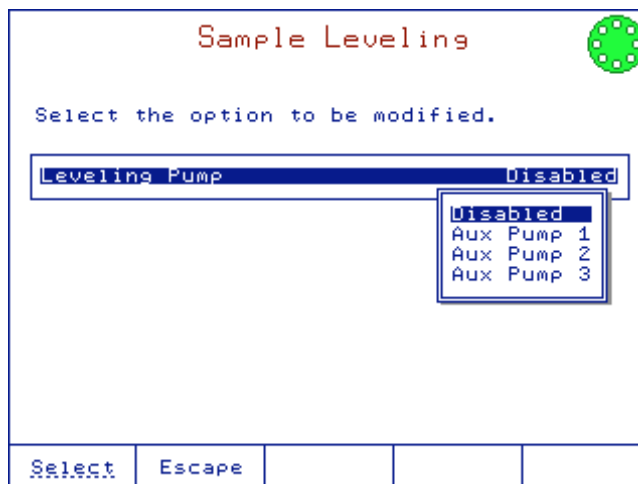
Sample Leveling

Option: Disabled or Enabled

Volumetric samples that do not require high accuracy can be leveled to the correct volume rather than manually dispensed with a pipette.

During sample leveling, excess sample is added to each beaker and then excess is being removed by the autosampler through the aspiration tube. This allows samples to be quickly poured into each beaker by the user while the autosampler accurately removes excess.

Note: Sample leveling requires one peristaltic pump configured for aspiration using HI920-203 tubing set for aspiration.



Leveling Pump

Select the peristaltic pump that is connected to the aspiration tube.

Sample Leveling	
Select the option to be modified.	
Leveling Pump	Aux Pump 1
Leveling Time	10 sec
Head Height	120 mm
Select	Escape

Leveling Time

Option: 1 second to 300 seconds

Set the duration that the peristaltic pump will run for.

Leveling Time	
Enter the period of time for running auxiliary pump.	
10 sec	
Low limit:	1 second
High limit:	300 seconds
Accept	Escape
Delete Digit	

Dispenser Head Height

Option: 10 to 150 mm

The height for the dispenser head should be set to produce the sample volume that is defined within method options.

The correct height must be determined experimentally by the user and will depend on the sample size, beaker shape and size, and the aspiration tube position.

The easiest way to determine the volume of a particular height setting is to manually aspirate water from a pre-weighed beaker and weigh the remaining water in the beaker.

Preset Head Height

Press <UP> or <DOWN> keys to position the head to appropriate position, or Use numeric keys to manually enter the head position.

0 mm

The range is from 10 to 150 mm. press <Accept> to save the head position.

Accept	Escape		△	▽
--------	--------	--	---	---

Reagent Addition 1 and 2

Option: Disabled or Enabled

Reagents and/or deionized water can be automatically added to each sample using the reagent addition feature.

Note: Reagent addition requires separate peristaltic pumps for each reagent.

The pumps should be configured for dispensing, using HI920-208 tubing set for dispensing.

The HI922 can perform up to two reagent additions.

Reagent Pump

Select the peristaltic pump that is connected to the reagent container.

Reagent Addition 1

Select the option to be modified.

Reagent Pump: Disabled

- Disabled
- Aux Pump 1
- Aux Pump 2
- Aux Pump 3

Select	Escape				
--------	--------	--	--	--	--

Reagent Addition 1	
Select the option to be modified.	
Reagent Pump:	Aux Pump 1
Dispenser Position:	130 mm
Dispensing Time:	1 sec
Stirring Time:	0 sec
Dispenser Waiting Position:	Down
Wait Time:	0 sec
Select	Escape

Dispenser Position

Option: 10 to 150 mm

Enter the position of the dispenser during reagent addition and stir time.

Preset Head Height			
Press <UP> or <DOWN> keys to position the head to appropriate position, or Use numeric keys to manually enter the head position.			
130 mm			
The range is from 10 to 150 mm. press <Accept> to save the head position.			
Accept	Escape	Delete Digit	Go to Position

Dispensing Time

Option: 1 to 300 seconds

Enter the dispensing time required to add the desired amount of reagent.

Note: This time should be determined experimentally. The flow rate is approximately 200 mL per minute.

Dispensing Time			
Enter the period of time for running auxiliary pump.			
5 sec			
Low limit:	1 second		
High limit:	300 seconds		
Accept	Escape	Delete Digit	

Stirring Time

Option: 0 to 1800 seconds

Set the amount of desired time to stir the sample after adding Reagent Addition 1.

Stirring Time				
Please enter the stirring time in seconds.				
<div style="background-color: black; color: white; display: inline-block; padding: 2px 10px;">2</div> sec				
Low limit: 0 second High limit: 1800 seconds				
Accept	Escape	Delete Digit		

Dispenser Waiting Position

Option: Up or Down

The user can set the position of the dispenser during wait time.

This is useful if it is undesirable for the electrode(s) to be immersed in the solution for extended periods of time.

Reagent Addition 1																
Select the option to be modified.																
<table border="1" style="width: 100%;"> <tbody> <tr> <td>Reagent Pump:</td> <td>Aux Pump 1</td> </tr> <tr> <td>Dispenser Position:</td> <td>130 mm</td> </tr> <tr> <td>Dispensing Time:</td> <td>1 sec</td> </tr> <tr> <td>Stirring Time:</td> <td>0 sec</td> </tr> <tr style="background-color: #e0e0e0;"> <td>Dispenser Waiting Position:</td> <td>Down</td> </tr> <tr> <td>Wait Time:</td> <td>C</td> </tr> </tbody> </table>					Reagent Pump:	Aux Pump 1	Dispenser Position:	130 mm	Dispensing Time:	1 sec	Stirring Time:	0 sec	Dispenser Waiting Position:	Down	Wait Time:	C
Reagent Pump:	Aux Pump 1															
Dispenser Position:	130 mm															
Dispensing Time:	1 sec															
Stirring Time:	0 sec															
Dispenser Waiting Position:	Down															
Wait Time:	C															
<table border="1" style="margin-left: auto;"> <tbody> <tr> <td>Up</td> </tr> <tr> <td>Down</td> </tr> </tbody> </table>					Up	Down										
Up																
Down																
Select	Escape															

Wait Time

Option: 0 to 1800 seconds

Set the reaction time.

This is the amount of time, after the stirring is completed, that the autosampler will wait before performing any other actions.

Wait Time				
Please enter the wait time in seconds.				
<div style="background-color: black; color: white; display: inline-block; padding: 2px 10px;">2</div> sec				
Low limit: 0 second High limit: 1800 seconds				
Accept	Escape	Delete Digit		

Addition Phase (linked methods only)**Option: First Titration or Second Titration**

Set the addition phase for the reagent addition.

Reagent addition can be done before the first titration or before the second titration.

The screenshot shows a terminal-style interface titled "Reagent Addition 1" with a green status indicator in the top right. The main instruction is "Select the option to be modified." Below this is a list of parameters with their current values and a character to the right of each value:

Reagent Pump:	Aux Pump 1	
Dispenser Position:	130 mm	
Dispensing Time:		c
Stirring Time:	First Titration	c
Dispenser Waiting Pos:	Second Titration	n
Wait Time:		c
Addition Phase:	First Titration	

At the bottom of the screen, there are five buttons: "Select", "Escape", and three empty buttons.

Analysis Method

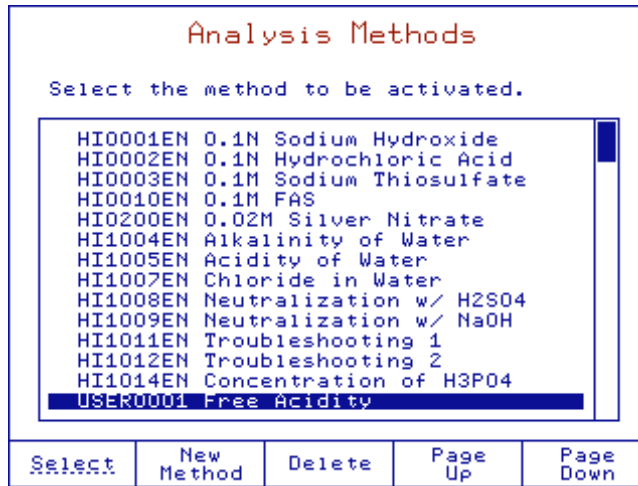
The below listed types of methods can be run on the autosampler.

Non-linked methods

- Sample titration, single endpoint (fixed or equivalence point)
- Sample titration, multiple equivalence point
- Direct reading
- Titration standardization
- Back titration

Linked methods

- Sample titration, single endpoint (fixed or equivalence point), linked to:
 - Sample titration, single endpoint (fixed or equivalence point)
 - Sample titration, multiple equivalence point
 - Direct reading
- Sample titration, multiple equivalence point, linked to:
 - Sample titration, single endpoint (fixed or equivalence point)
 - Sample titration, multiple equivalence point
 - Direct reading
- Direct reading, linked to:
 - Sample titration, single endpoint (fixed or equivalence point)
 - Sample titration, multiple equivalence point
 - Direct reading
 - Back titration
- Back titration, linked to:
 - Direct reading

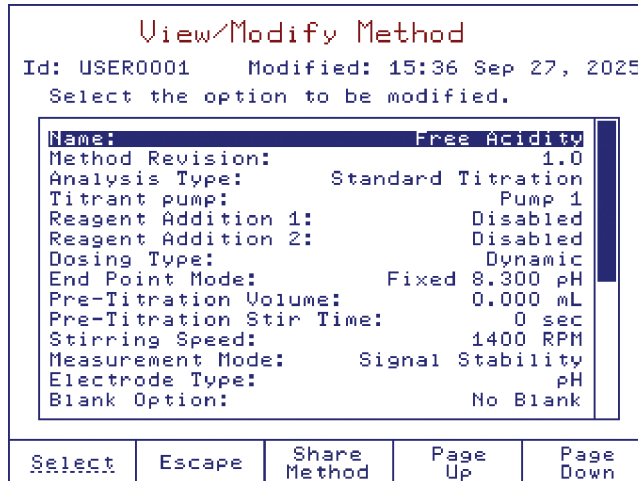


Method Options

The analysis method options can be accessed directly from the autosampler interface.

Analysis method options can be reviewed and/or modified if necessary.

See [2.4.5. Method Options](#) section for more information.

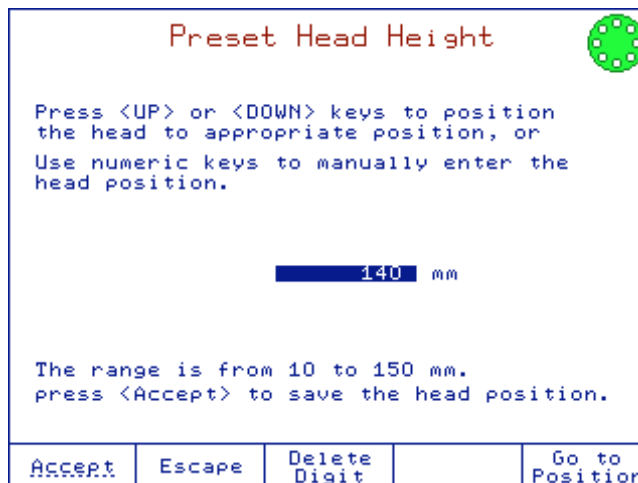


Dispenser Position

Option: 10 to 150 mm

User can select the height for the dispenser head to be positioned at during titration.

The default option is 140 mm.



Head Up Wait Time

Option: 1 to 30 seconds

Set the duration that the autosampler will wait with the dispenser in the up position for any drops of solution to fall off of the electrodes or stirrer before moving to another sample or rinse beaker.

Head Up Wait Time

Please enter the wait time in seconds.

2 sec

Low limit: 1 second
High limit: 30 seconds

Accept Escape Delete Digit

Sample Aspiration

Option: Disabled, Aspirate Only, Aspirate / Spray Rinse

Reacted samples may be aspirated into a waste container after each titration.

Note: Sample aspiration requires one peristaltic pump to be configured for aspiration, using HI920-203 Tubing Set for Aspiration.

Aspirate Sample

Select the option to be modified.

Aspiration Option: Disabled

- Disabled
- Aspirate Only
- Aspirate/Spray Rinse

Select Escape

Aspirate Only The existing waste from the sample beaker will be removed according to the parameters defined in this menu.

Aspirate / Spray Reserved for future

Aspiration Pump

Option: Disabled, Auxiliary Pump 1, Auxiliary Pump 2, Auxiliary Pump 3

Select the peristaltic pump that is connected to the aspiration tube.

Aspirate Sample

Select the option to be modified.

Aspiration Option:	Aspirate Only
Aspiration Pump:	Disabled
Aspiration Time:	
Head Height:	

Disabled

Aux Pump 1

Aux Pump 2

Aux Pump 3

Select	Escape		
--------	--------	--	--

Aspiration Time

Option: 1 to 300 seconds

Set the duration that the peristaltic pump will run.

Aspiration Time

Enter the period of time for running auxiliary pump.

15 sec

Low limit: 1 second
High limit: 300 seconds

Accept	Escape	Delete Digit	
--------	--------	--------------	--

Dispenser Head Height

Option: 10 to 150 mm

Set the height for the dispenser head.

The aspiration tube should be positioned such that it reaches the bottom of the sample beaker when the dispenser head is positioned in the range of 10 to 150 mm.

Preset Head Height

Press <UP> or <DOWN> keys to position the head to appropriate position, or Use numeric keys to manually enter the head position.

140 mm

The range is from 10 to 150 mm.
press <Accept> to save the head position.

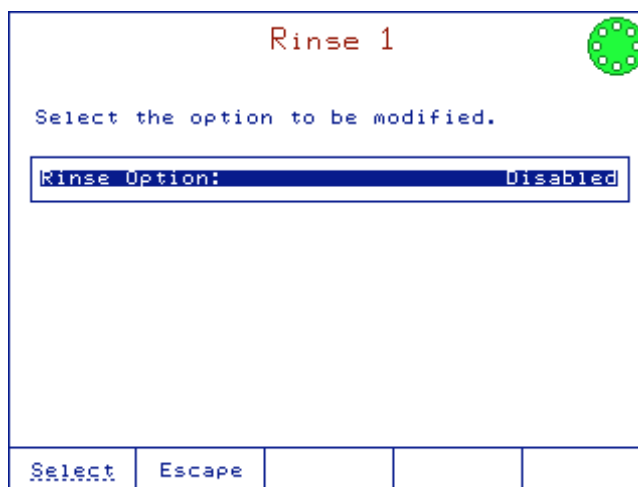
Accept	Escape	Delete Digit	Go to Position
--------	--------	--------------	----------------

Rinse

Option: Disabled, Dip Rinse, Spray Rinse

The autosampler can perform a dip rinse function after each analysis.

Up to three dip rinses can be performed in a dedicated rinse beaker dip-rinse function.

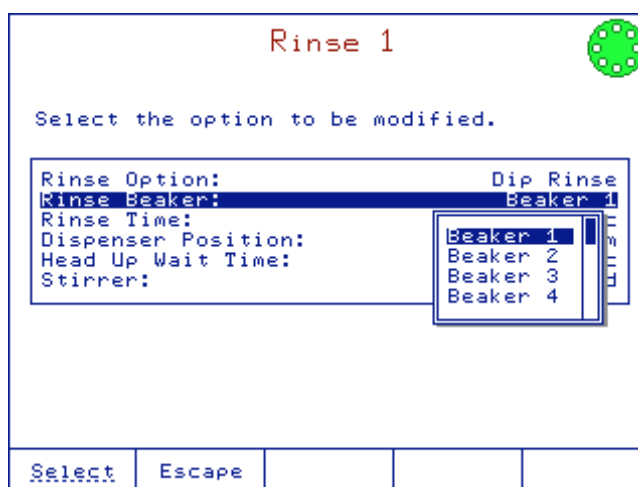


Dip Rinse Dip rinse option can be used after each analysis to clean the electrodes and stirrer of contaminants, using dedicated rinsing beakers.

Spray Rinse Reserved for future

Rinse Beaker

Select the tray position for the dedicated rinse beaker.



Rinse Time

Option: 1 to 300 seconds

<h3 style="color: red;">Rinse Time</h3> <p>Enter the period of time you would like to remain in the rinse beaker.</p> <p style="text-align: center;">10 sec</p> <p>Low limit: 1 second High limit: 300 seconds</p>				
Accept	Escape	Delete Digit		

Dispenser Position

Option: 10 to 150 mm

Set the height for the dispenser head during rinsing.

<h3 style="color: red;">Preset Head Height</h3> <p>Press <UP> or <DOWN> keys to position the head to appropriate position, or Use numeric keys to manually enter the head position.</p> <p style="text-align: center;">140 mm</p> <p>The range is from 10 to 150 mm. press <Accept> to save the head position.</p>				
Accept	Escape	Delete Digit		Go to Position

Head Up Wait Time

Option: 1 to 300 seconds

Set the duration that the autosampler will wait with the dispenser in the up position for any drops of solution to fall off of the electrodes or stirrer before moving to another sample or rinse beaker.

<h3 style="color: red;">Head Up Wait Time</h3> <p>Please enter the stirring time in seconds.</p> <p style="text-align: center;">1 sec</p> <p>Low limit: 1 second High limit: 300 seconds</p>				
Accept	Escape	Delete Digit		

Stirrer

Option: Enabled or Disabled

Select if the stirrer will run during rinsing.

Rinse 1	
Select the option to be modified.	
Rinse Option:	Dip Rinse
Rinse Beaker:	Beaker 1
Rinse Time:	10 sec
Dispenser Position:	140 mm
Head Up Wait Time:	1 sec
Stirrer:	Enabled
<div style="border: 1px solid black; padding: 2px; display: inline-block;"> Disabled Enabled </div>	
Select	Escape

Beaker Height

Option: 30 to 120 mm

Set the height of the beaker being used on the autosampler.

Beaker Height	
Enter the beaker height in mm.	
<div style="border: 1px solid black; display: inline-block; padding: 2px;">100 mm</div>	
Low limit:	30 mm
High limit:	120 mm
Accept	Escape
Delete Digit	

Position when Finished

Option: Home, Sample, Storage

View / Modify Sequence	
Id: SEQ0002 Modified: 11:48 Mar 24, 2026	
Select the option to be modified.	
Reagent Addition 2:	Disabled
Method:	USER0003, Free Acidity
Linked To:	USER0004, Direct pH reading
Method Options:	
Dispenser Position:	140 mm
Head Up Wait Time:	1 sec
Aspirate Sample:	Asp
Rinse 1:	d
Rinse 2:	d
Rinse 3:	d
Beaker Height:	m
Position when finished:	Storage
<div style="border: 1px solid black; padding: 2px; display: inline-block;"> Home Sample Storage </div>	
Select	Escape

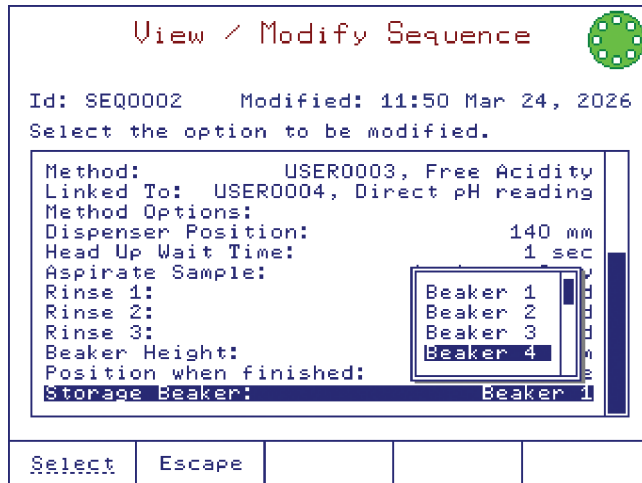
Home The dispenser head will be in the up positioned above beaker one.

Sample The dispenser head will remain down in the last sample that was analyzed / titrated.

Storage The dispenser head will be down in a preset beaker containing storage solution.

Storage Beaker (position when finished, storage only)

User can select the beaker to be placed in storage at the end of a sequence analysis. After the sequence has been completed, the autosampler will automatically move the beaker to storage and lower the dispenser head.

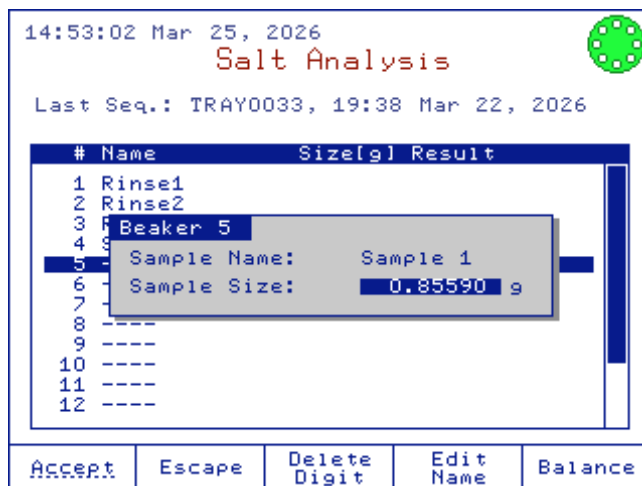


2.11.5.2. Sample Table

All sample information is entered into the sample table according to the tray position. The sample table screen is the default screen when entering the autosampler interface while the autosampler is idle. The sample table is automatically formatted with the appropriate beaker number, with rinse / storage beaker positions reserved.

To add a sample to the sample table follow the steps detailed below.

- Use the and keys to highlight an empty beaker position.
- Use to opens the sample dialog box. Edit the sample name and size.



- Use to enter the current sample name and size into the sample table.
- Use to return to the sample table.
- Use to modify the sample size.
- Use to modify the sample name.
- Use to access the balance interface for direct entry of sample weight (if available).

Several features have been added to to speed up sample entry, depending on your peripheral connections and analysis method. With an empty table position highlighted, below detailed features are available.

Name entry	If a non-numeric character is entered using an external keyboard, the sample name will be updated.
Size entry	If a numeric character is entered on the keypad or external keyboard, the sample size will be updated. The sample name will be auto-incremented.
Barcode reader	Scanning a barcode automatically enters the barcode into the sample name field.
Fixed sample size	The sample dialog box is omitted if the sample size entry is set to Fixed. Any character entered from the keypad or external keyboard will go directly to the Edit Name screen.
Autofill (Fixed sample size)	All empty sample table positions are filled automatically. The sample name will be auto-incremented.

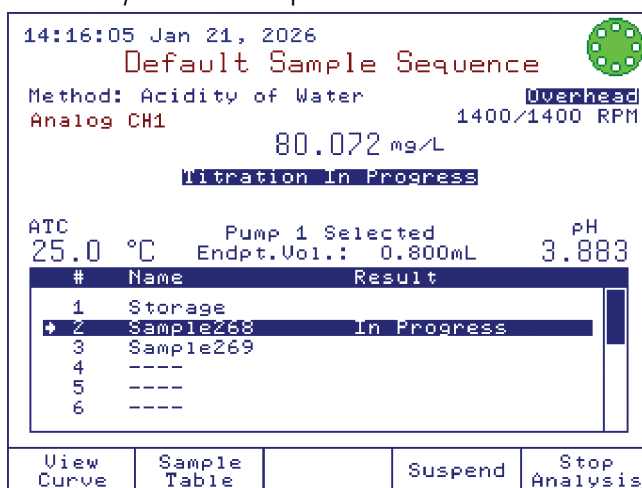
2.11.5.3. Running Sample Analysis Sequence

The sequence can be started by pressing the  key.

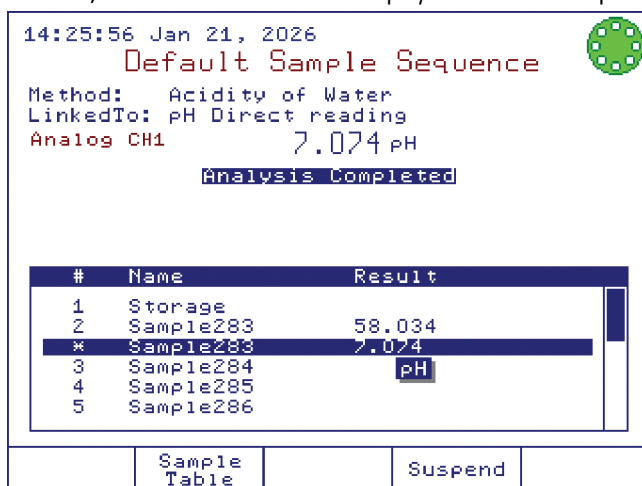
The autosampler will process each sample according to the settings in sequence options.







While the autosampler is running, the top part of the screen shows titration information for the current titration, and the bottom part of the screen shows a portion of the sample table.

The sample in progress is marked with  symbol in the sample table.



If the selected titration method is linked, the linked method will be displayed below the sample when it is in progress.



- Use the  and  keys to scroll the sample table.
- Use  to view the graph of the current titration.
- Use  to view or modify the sample table entries. Sample can be added to the sample table while the autosampler is running.
- Use  to pause the current titration.
- Use  to end the current titration immediately and proceed to the next sample.

2.11.5.4. Results

Press View GLP to view GLP Data or the results key for more information.

14:30:52 Jan 21, 2026 🟢

Default Sample Sequence

Last Seq.: TRAY0013, 14:28 Jan 21, 2026

#	Name	Result
1	Storage	
2	Sample298	7.083
3	Sample299	3.877
4	----	pH
5	----	
6	----	
7	----	
8	----	
9	----	
10	----	
11	----	
12	----	

View Result	Auto Fill	Reset Table	AutoSmp. Setup
-------------	-----------	-------------	----------------

Analysis Reports 🟢

Sequence Name: Default Sample Sequence
 Tray ID: TRAY0013
 Date & Time: 14:28 Jan 21, 2026

#	Sample Name	Result	Report Id
2	Sample298	7.083	pH_11301
3	Sample299	3.877	pH_11302
		pH	

Escape	View Report	Share Report	Delete Report
--------	-------------	--------------	---------------

2.11.6. pH CALIBRATION

2.11.6.1. View / Modify Sequence

- To modify the sequence options, press {Sequence} Options from the main screen.
A list of all the parameters for the selected sequence will be displayed.
- Use the ▲ and ▼ keys to highlight the option to be modified.
- Next, press Select.
- To exit the **View / Modify Sequence** screen, press Escape.

View / Modify Sequence 🟢

Id: SEQ0002 Modified: 18:41 Mar 06, 2026

Select the option to be modified.

Sequence Name:	Default pH Sequence
Revision Number:	1.0
Comments:	
Sequence Type:	pH Calibration
Measurement Input:	Analog CH1
Stirrer Type:	Overhead
Missing Beaker Behavior:	Stop
pH Setup:	
Edit Sample Table:	
Dispenser Position:	140 mm
Head Up Wait Time:	1 sec
Aspirate Sample:	Disabled

Select	Escape	Share Sequence	Page Up	Page Down
--------	--------	----------------	---------	-----------

You can choose to save the modifications or to discard them.

Saving Sequence				
Select a menu option.				
<div style="border: 1px solid black; padding: 2px;"> Save Sequence Exit Without Saving Sequence </div>				
"Escape" - exit without saving sequence.				
Select	Escape			

Sequence Name

Option: Up to 24 characters

Sequence Name				
Select the highlighted letter by using the arrow keys then press "Enter". Select the empty field for a space. Press Accept to save the entered text.				
<pre> A B C D E F G H I J K L M N O P Q R S T U V W X Y Z a b c d e f g h i j k l m n o p q r s t u v w x y z A A A A A C E E E I I N 0 0 0 0 0 0 0 0 0 0 0 0 a c e e e i i n o o o o o 0 0 0 0 0 0 0 0 0 0 0 0 ? ! () [] < > = / + - </pre> <p style="text-align: center;">pH Calibration</p>				
Accept	Escape	Delete Letter	Cursor Left	Cursor Right

Revision Number

Option: Up to 3 characters

Revision Number				
Select the highlighted letter by using the arrow keys then press "Enter". Select the empty field for a space. Press Accept to save the entered text.				
<pre> A B C D E F G H I J K L M N O P Q R S T U V W X Y Z a b c d e f g h i j k l m n o p q r s t u v w x y z A A A A A C E E E I I N 0 0 0 0 0 0 0 0 0 0 0 0 a c e e e i i n o o o o o 0 0 0 0 0 0 0 0 0 0 0 0 ? ! () [] < > = / + - </pre> <p style="text-align: center;">1.0</p>				
Accept	Escape	Delete Letter	Cursor Left	Cursor Right

Comments

Option: Up to 20 characters

Comments

Select the highlighted letter by using the arrow keys then press "Enter". Select the empty field for a space. Press Accept to save the entered text.

<input checked="" type="checkbox"/>	A	B	C	D	E	F	G	H	I	J	K	L
M	N	O	P	Q	R	S	T	U	V	W	X	Y
Z	a	b	c	d	e	f	g	h	i	j	k	l
m	n	o	p	q	r	s	t	u	v	w	x	y
z	À	Á	Â	Ã	Ä	Å	Æ	Ç	È	É	Ê	Ë
Ì	Í	Î	Ï	Ñ	Ò	Ó	Ô	Õ	Ö	×	Ù	Ú
Û	Ü	Ý	ÿ	ı	*	\	_	^	#	:		
0	1	2	3	4	5	6	7	8	9	×	.	,
?	!	()	[]	<	>	=	/	+	-	

█

Accept	Escape	Delete Letter	Cursor Left	Cursor Right
--------	--------	---------------	-------------	--------------

Sequence Type

Option: Sample Analysis, pH Calibration, ISE Calibration

View / Modify Sequence

Id: SEQ0002 Modified: 18:41 Mar 06, 2026

Select the option to be modified.

Sequence Name:	Default pH Sequence
Revision Number:	1.0
Comments:	
Sequence Type:	pH Calibration
Measurement Input:	1
Stirrer Type:	d
Missing Beaker Behavior:	p
pH Setup:	
Edit Sample Table:	
Dispenser Position:	140 mm
Head Up Wait Time:	1 sec
Aspirate Sample:	Disabled

Sample Analysis	d
pH Calibration	p
ISE Calibration	

Select	Escape			
--------	--------	--	--	--

Measurement Input

Option: Analog CH1, Analog CH2, Digital CH3, or Digital CH4

View / Modify Sequence

Id: SEQ0002 Modified: 18:41 Mar 06, 2026

Select the option to be modified.


Sequence Name:	Default pH Sequence
Revision Number:	1.0
Comments:	
Sequence Type:	pH Calibration
Measurement Input:	Analog CH1
Stirrer Type:	d
Missing Beaker Behavior:	p
pH Setup:	
Edit Sample Table:	
Dispenser Position:	
Head Up Wait Time:	
Aspirate Sample:	Disabled

Analog CH1	p
Analog CH2	
Digital CH3	
Digital CH4	m

Select	Escape			
--------	--------	--	--	--

Stirrer Type

Option: Overhead or Magnetic

View / Modify Sequence 

Id: SEQ0002 Modified: 18:41 Mar 06, 2026
Select the option to be modified.

Sequence Name:	Default pH Sequence
Revision Number:	1.0
Comments:	
Sequence Type:	pH Calibration
Measurement Input:	Analog CH1
Stirrer Type:	Overhead
Missing Beaker Behavior:	Overhead
pH Setup:	
Edit Sample Table:	
Dispenser Position:	
Head Up Wait Time:	1 sec
Aspirate Sample:	Disabled


Overhead
Magnetic

Select	Escape		
--------	--------	--	--

Missing Beaker Behavior

Option: Pause, Stop

Select type of behaviour when beaker detection is enabled and no beaker is detected.

View / Modify Sequence 

Id: SEQ0002 Modified: 18:41 Mar 06, 2026
Select the option to be modified.

Sequence Name:	Default pH Sequence
Revision Number:	1.0
Comments:	
Sequence Type:	pH Calibration
Measurement Input:	Analog CH1
Stirrer Type:	Overhead
Missing Beaker Behavior:	Stop
pH Setup:	
Edit Sample Table:	
Dispenser Position:	
Head Up Wait Time:	
Aspirate Sample:	Disabled

Pause
Stop

Select	Escape		
--------	--------	--	--


Pause The autosampler will pause the sequence at the current beaker and wait for the user before continuing the analysis.

Stop All operations will stop and the analysis will be stopped.

pH Setup

This option allows the user to select pH Setup menu.

See [2.6.2. pH Setup](#) section for more information.

View / Modify Sequence 

Id: SEQ0002 Modified: 18:41 Mar 06, 2026
 Select the option to be modified.

Sequence Name:	Default pH Sequence
Revision Number:	1.0
Comments:	
Sequence Type:	pH Calibration
Measurement Input:	Analog CH1
Stirrer Type:	Overhead
Missing Beaker Behavior:	Stop
pH Setup:	
Edit Sample Table:	
Dispenser Position:	140 mm
Head Up Wait Time:	1 sec
Aspirate Sample:	Disabled

Select	Escape	Share Sequence	Page Up	Page Down
--------	--------	----------------	---------	-----------

pH Setup


Select a menu option.

Buffer Entry Type:	Manual
First Cal Point:	Point
Edit Custom Buffers	
Edit Buffer Group	
Calibration Reminder:	Disabled
Set Reminder Period:	Disabled
Clear Calibration	
pH GLP Data	
Logging Interval:	Disabled
Stability Criteria:	Medium
pH Resolution:	X.XXX
Stirrer Configuration:	Always Stirring

Select	Escape			
--------	--------	--	--	--

Edit Sample Table

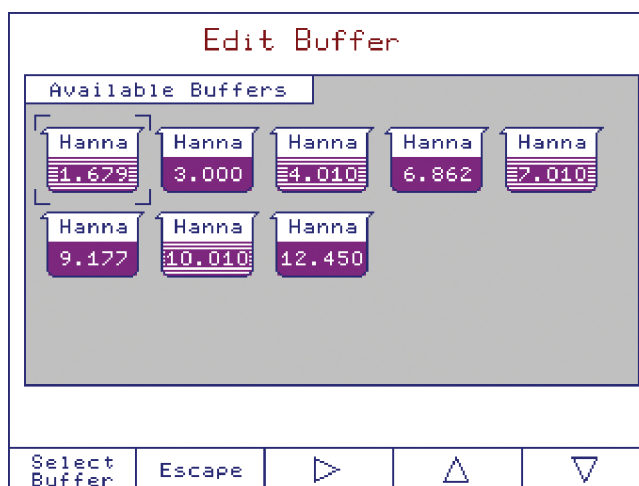
This option allows to add, remove and edit the calibration beakers in the tray.

Edit Sample Table 

#	Name
1	Storage
2	Rinse01.679
3	pH01.679Hanna
4	Rinse04.010
5	pH04.010Hanna
6	Rinse10.010
7	pH10.010Hanna
8	Rinse07.010
9	pH07.010Hanna
10	----
11	----
12	----

Edit Buffer	Escape	Delete Buffer		
-------------	--------	---------------	--	--

Note: Up to five calibration beakers can be added. Highlighted beakers have already been added to the calibration sequence.

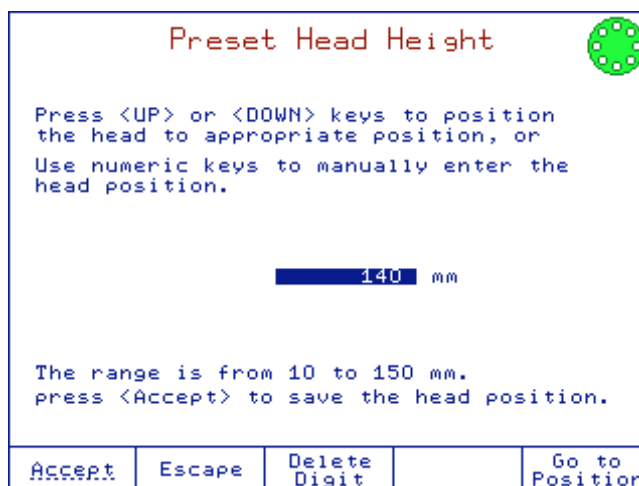


Dispenser Position

Option: 10 to 150 mm

User can select the height for the dispenser head to be positioned at during titration.

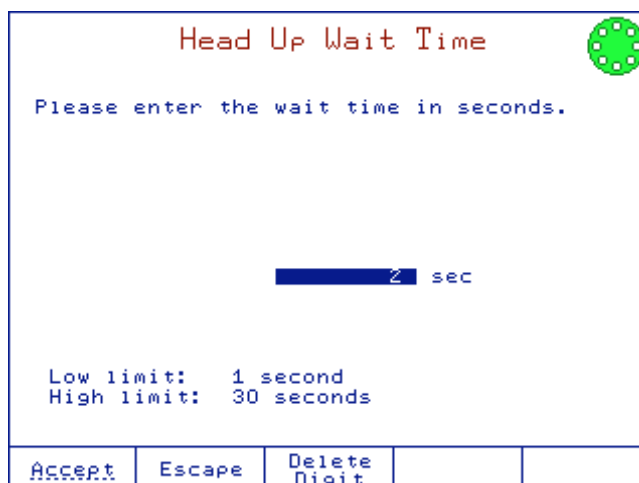
The default option is 140 mm.



Head Up Wait Time

Option: 1 to 30 seconds

User can set the time for the autosampler to wait, with the dispenser head in the up position to catch any residual droplets, before moving on to the next sample or rinse beaker.



Sample Aspiration

Option: Disabled, Aspirate Only, Aspirate / Spray Rinse

Reacted samples may be aspirated into a waste container after each titration.

Note: Sample aspiration requires one peristaltic pump to be configured for aspiration, using [HI920-203 Tubing Set for Aspiration](#).

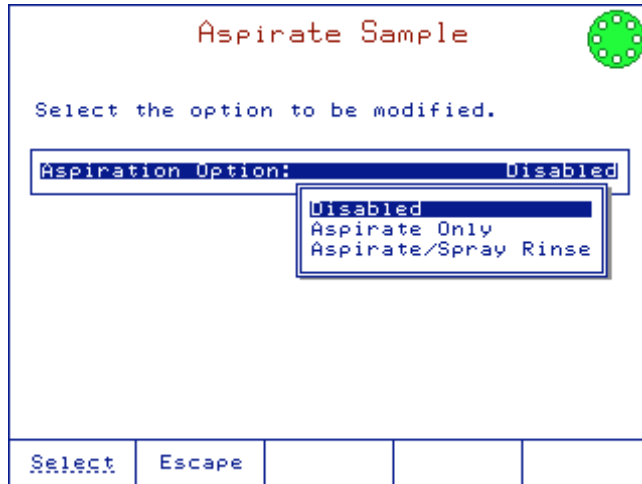
Select the aspiration mode.

Aspirate Only

The existing waste from the sample beaker will be removed according to the parameters defined in this menu.

Aspirate / Spray rinse

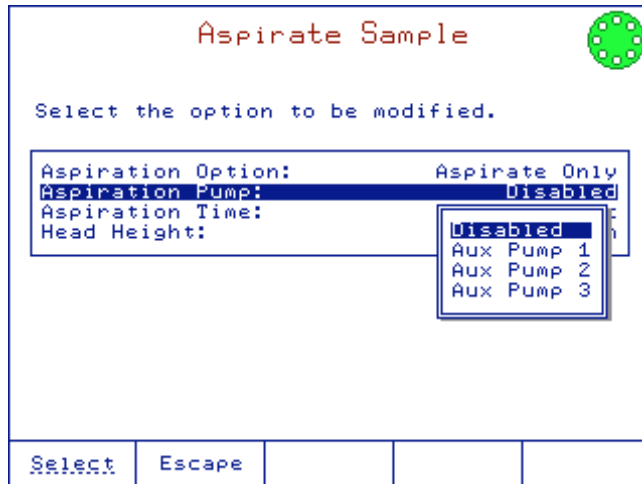
Reserved for future



Aspiration Pump

Option: Disabled, Auxiliary Pump 1, Auxiliary Pump 2, Auxiliary Pump 3

Select the peristaltic pump that is connected to the aspiration tube.



Aspiration Time

Option: 1 to 300 seconds

Set the duration that the peristaltic pump will run.


Aspiration Time 				
Enter the period of time for running auxiliary pump.				
15 sec				
Low limit: 1 second High limit: 300 seconds				
ACCEPT	Escape	Delete Digit		

Dispenser Head Height

Option: 10 to 150 mm

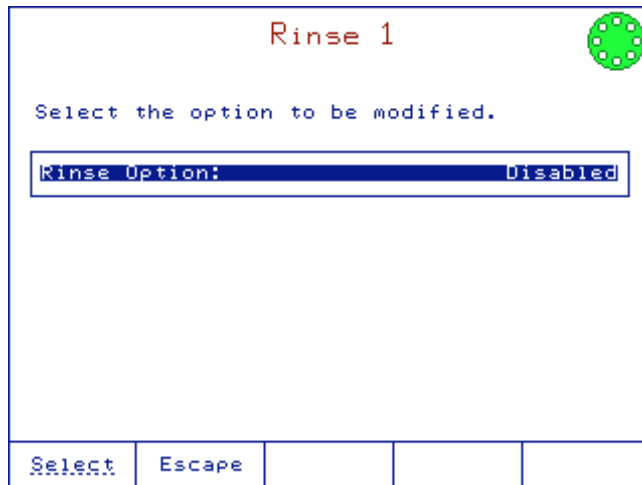
Set the height for the dispenser head.

The aspiration tube should be positioned such that it reaches the bottom of the sample beaker when the dispenser head is positioned in the range of 10 to 150 mm.

Preset Head Height 				
Press <UP> or <DOWN> keys to position the head to appropriate position, or Use numeric keys to manually enter the head position.				
140 mm				
The range is from 10 to 150 mm. press <Accept> to save the head position.				
ACCEPT	Escape	Delete Digit		Go to Position

Rinse

The autosampler can perform a dip rinse function after each analysis.
 Up to three dip rinses can be performed in a dedicated rinse beaker dip-rinse function.

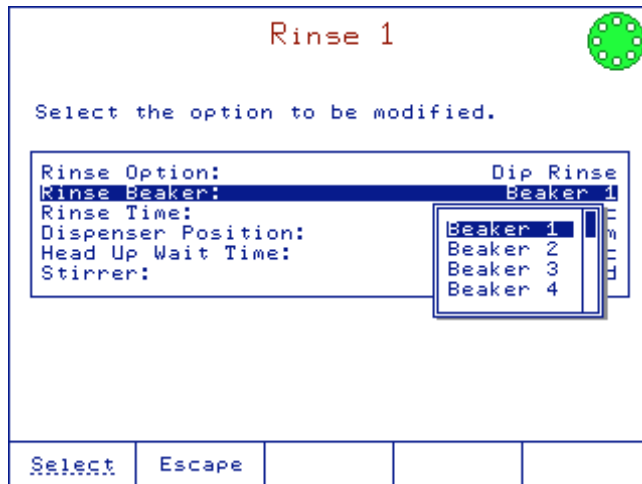


Dip rinse Dip rinse option can be used after each analysis to clean the electrodes and stirrer of contaminants, using dedicated rinsing beakers.

Spray Rinse Reserved for future

Rinse Beaker

Select the tray position for the dedicated rinse beaker.



Note: If the selected beaker is already in use for buffer, standard or calibration rinse, it will be removed from the sample table.

Rinse Time

Option: 1 to 300 seconds

Rinse Time				
Enter the period of time you would like to remain in the rinse beaker.				
10 sec				
Low limit: 1 second High limit: 300 seconds				
ACCEPT	Escape	Delete Digit		

Dispenser Position

Option: 10 to 150 mm

Set the height for the dispenser head during rinsing.

Preset Head Height				
Press <UP> or <DOWN> keys to position the head to appropriate position, or Use numeric keys to manually enter the head position.				
140 mm				
The range is from 10 to 150 mm. press <Accept> to save the head position.				
ACCEPT	Escape	Delete Digit		Go to Position

Head Up Wait Time

Option: 1 to 300 seconds

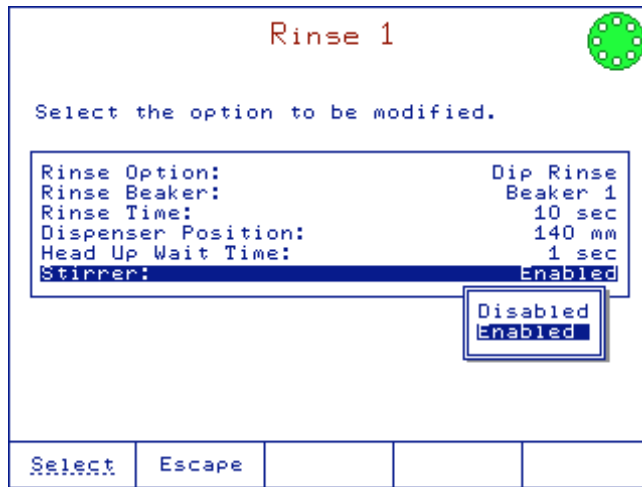
Set the duration that the autosampler will wait with the dispenser in the up position for any drops of solution to fall off of the electrodes or stirrer before moving to another sample or rinse beaker.

Head Up Wait Time				
Please enter the stirring time in seconds.				
1 sec				
Low limit: 1 second High limit: 300 seconds				
ACCEPT	Escape	Delete Digit		

Stirrer

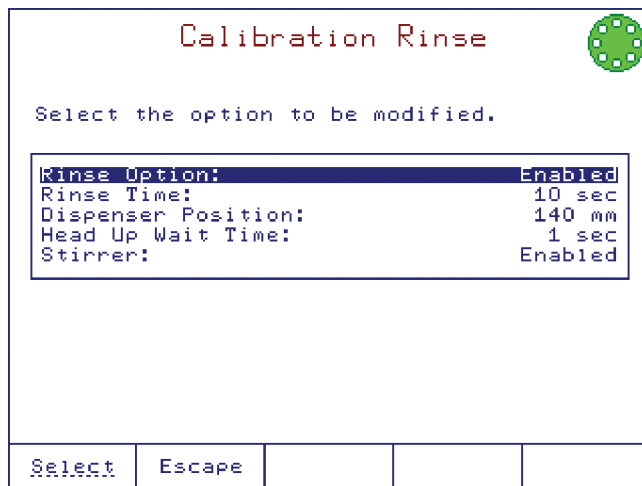
Option: Enabled or Disabled

Select if the stirrer will run during rinsing.



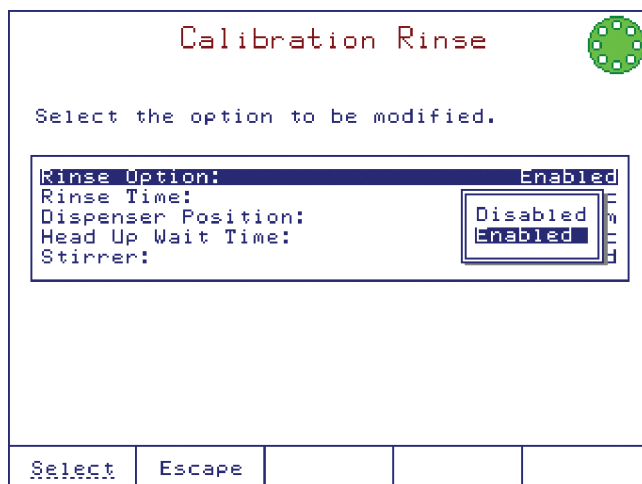
Calibration Rinse

The user can select to enable the rinse option during pH calibrations.



Rinse Option

Option: Enabled or Disabled



Rinse Time


Option: 1 to 300 seconds

Rinse Time 				
Enter the period of time you would like to remain in the rinse beaker.				
10 sec				
Low limit: 1 second High limit: 300 seconds				
ACCEPT	Escape	Delete Digit		

Dispenser Position

Option: 10 to 150 mm


Set the height for the dispenser head during rinsing.

Preset Head Height 				
Press <UP> or <DOWN> keys to position the head to appropriate position, or Use numeric keys to manually enter the head position.				
140 mm				
The range is from 10 to 150 mm. press <Accept> to save the head position.				
ACCEPT	Escape	Delete Digit		Go to Position

Head Up Wait Time

Option: 1 to 300 seconds

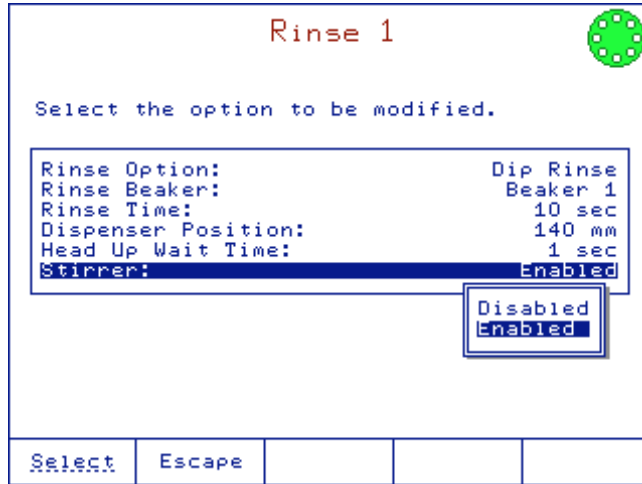
Set the duration that the autosampler will wait with the dispenser in the up position for any drops of solution to fall off of the electrodes or stirrer before moving to another sample or rinse beaker.

Head Up Wait Time 				
Please enter the stirring time in seconds.				
1 sec				
Low limit: 1 second High limit: 300 seconds				
ACCEPT	Escape	Delete Digit		

Stirrer

Option: Enabled or Disabled

Select if the stirrer will run during rinsing.



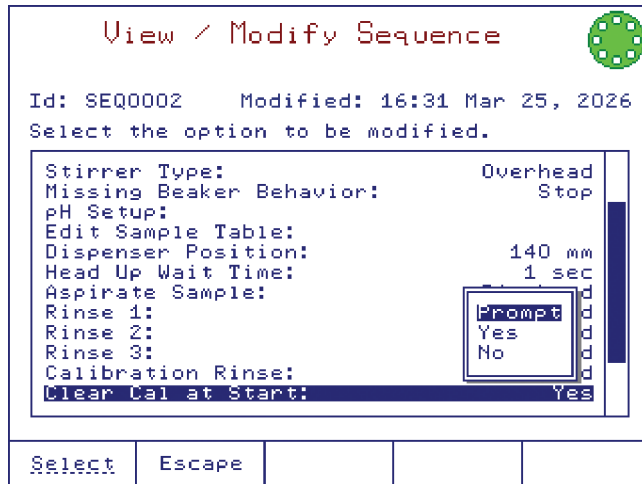
Clear Calibration at Start

Option: Prompt, Yes, No

Prompt Prompts the user to clear the calibration when the sequence starts

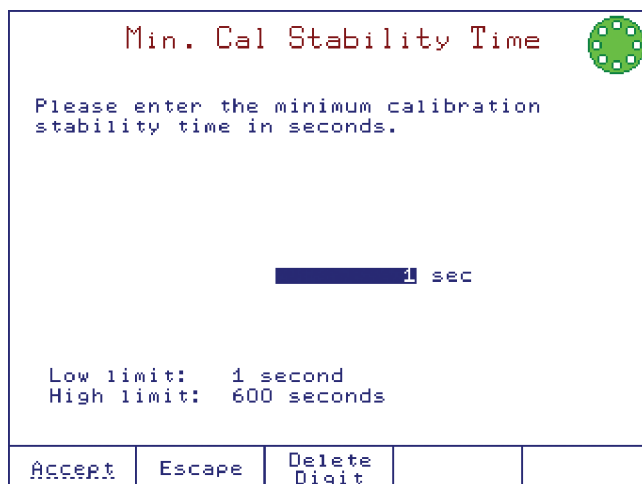
Yes Always clear the old calibration

No The new calibration data will be merged with the old data



Minimum Calibration Stability Time

Option: 1 to 600 seconds



Beaker Height

Option: 30 to 120 mm

Set the height of the beaker being used on the autosampler.

Beaker Height				
Enter the beaker height in mm.				
100 mm				
Low limit: 30 mm High limit: 120 mm				
ACCEPT	Escape	Delete Digit		

Position when Finished

Option: Home, Buffer, Storage

Home The dispenser head will be in the up positioned above beaker one.

Buffer The dispenser head will remain down in the last buffer that was analyzed / titrated.

Storage The dispenser head will be down in a preset beaker containing storage solution.


View / Modify Sequence																												
Id: SEQ0002 Modified: 16:31 Mar 25, 2026																												
Select the option to be modified.																												
<table border="1"> <tr><td>Edit Sample Table:</td><td></td></tr> <tr><td>Dispenser Position:</td><td>140 mm</td></tr> <tr><td>Head Up Wait Time:</td><td>1 sec</td></tr> <tr><td>Aspirate Sample:</td><td>Disabled</td></tr> <tr><td>Rinse 1:</td><td>Disabled</td></tr> <tr><td>Rinse 2:</td><td>Disabled</td></tr> <tr><td>Rinse 3:</td><td>Disabled</td></tr> <tr><td>Calibration Rinse:</td><td>Home</td></tr> <tr><td>Clear Cal at Start:</td><td>Buffer</td></tr> <tr><td>Min. Cal Stability Time:</td><td>Storage</td></tr> <tr><td>Beaker Height:</td><td></td></tr> <tr><td>Position when finished:</td><td>Storage</td></tr> </table>					Edit Sample Table:		Dispenser Position:	140 mm	Head Up Wait Time:	1 sec	Aspirate Sample:	Disabled	Rinse 1:	Disabled	Rinse 2:	Disabled	Rinse 3:	Disabled	Calibration Rinse:	Home	Clear Cal at Start:	Buffer	Min. Cal Stability Time:	Storage	Beaker Height:		Position when finished:	Storage
Edit Sample Table:																												
Dispenser Position:	140 mm																											
Head Up Wait Time:	1 sec																											
Aspirate Sample:	Disabled																											
Rinse 1:	Disabled																											
Rinse 2:	Disabled																											
Rinse 3:	Disabled																											
Calibration Rinse:	Home																											
Clear Cal at Start:	Buffer																											
Min. Cal Stability Time:	Storage																											
Beaker Height:																												
Position when finished:	Storage																											
Select	Escape																											

Storage Beaker (Position when finished, storage only)

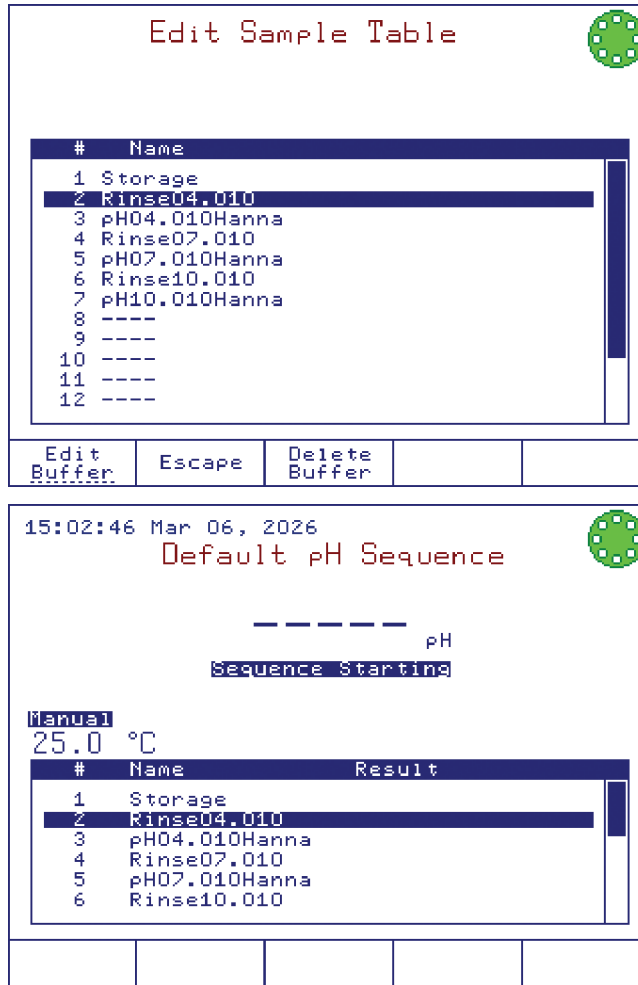
After the sequence has been completed, the autosampler will move to this position automatically and lower the dispenser head.

View / Modify Sequence																												
Id: SEQ0002 Modified: 16:31 Mar 25, 2026																												
Select the option to be modified.																												
<table border="1"> <tr><td>Dispenser Position:</td><td>140 mm</td></tr> <tr><td>Head Up Wait Time:</td><td>1 sec</td></tr> <tr><td>Aspirate Sample:</td><td>Disabled</td></tr> <tr><td>Rinse 1:</td><td>Disabled</td></tr> <tr><td>Rinse 2:</td><td>Disabled</td></tr> <tr><td>Rinse 3:</td><td>Disabled</td></tr> <tr><td>Calibration Rinse:</td><td>Disabled</td></tr> <tr><td>Clear Cal at Start:</td><td>Beaker 1</td></tr> <tr><td>Min. Cal Stability Time:</td><td>Beaker 2</td></tr> <tr><td>Beaker Height:</td><td>Beaker 3</td></tr> <tr><td>Position when finished:</td><td></td></tr> <tr><td>Storage Beaker:</td><td>Beaker 1</td></tr> </table>					Dispenser Position:	140 mm	Head Up Wait Time:	1 sec	Aspirate Sample:	Disabled	Rinse 1:	Disabled	Rinse 2:	Disabled	Rinse 3:	Disabled	Calibration Rinse:	Disabled	Clear Cal at Start:	Beaker 1	Min. Cal Stability Time:	Beaker 2	Beaker Height:	Beaker 3	Position when finished:		Storage Beaker:	Beaker 1
Dispenser Position:	140 mm																											
Head Up Wait Time:	1 sec																											
Aspirate Sample:	Disabled																											
Rinse 1:	Disabled																											
Rinse 2:	Disabled																											
Rinse 3:	Disabled																											
Calibration Rinse:	Disabled																											
Clear Cal at Start:	Beaker 1																											
Min. Cal Stability Time:	Beaker 2																											
Beaker Height:	Beaker 3																											
Position when finished:																												
Storage Beaker:	Beaker 1																											
Select	Escape																											





2.11.6.2. Running pH Calibration Sequence

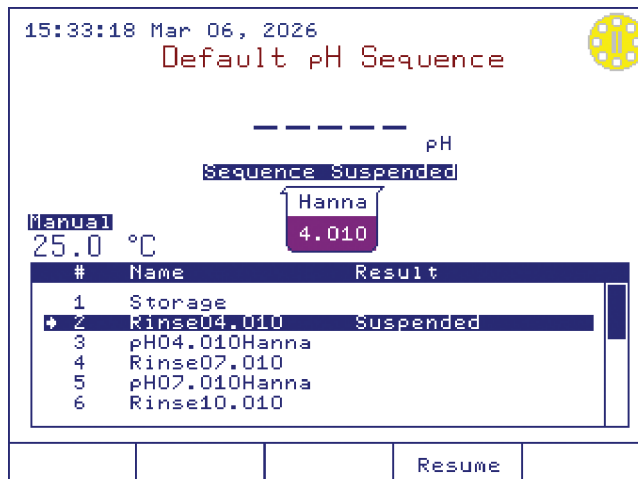
Press the  key to start the sequence.

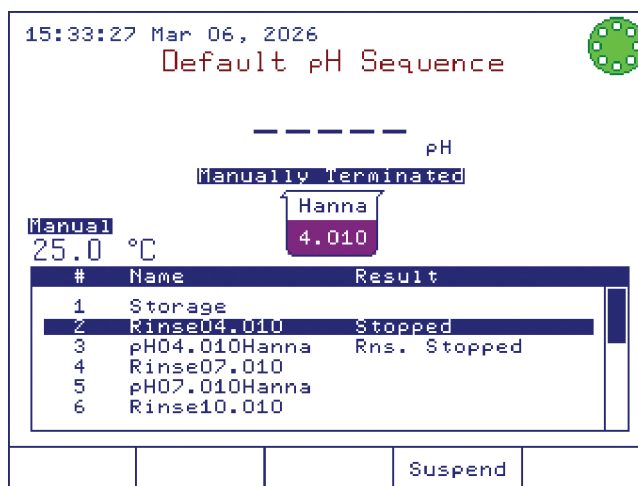
Note: If the Calibration Rinse option is enabled, the beaker used for rinsing are positioned before beaker used for buffer calibration.



The autosampler will process each calibration point according to the settings in sequence options. While the autosampler is running, the top part of the screen shows the calibration information, and the bottom part of the screen shows a portion of the sample table. The sample in progress is marked with symbol in the sample table.

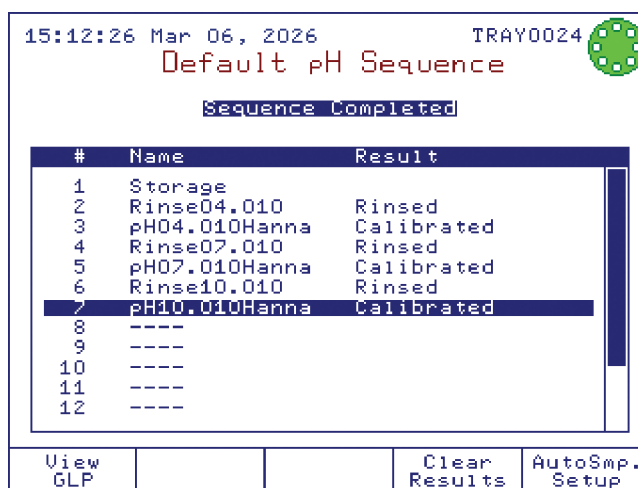
- Use the  and  keys to scroll the sample table.
- Press  to pause the current calibration.
- Press  to stop the calibration.





At the end of the sequence "Sequence Completed" will be displayed.

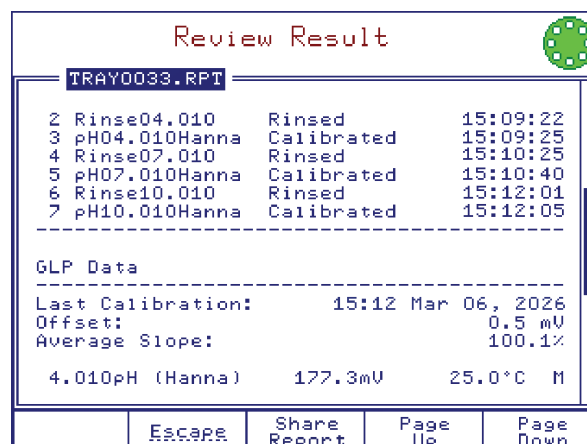
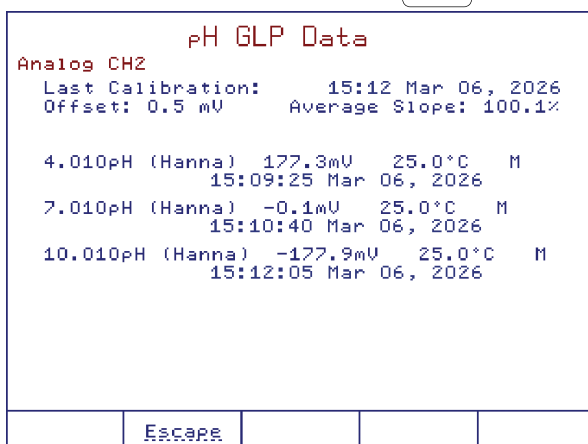
The "Result" column will display the status for each beaker.



Note: If errors will appear during calibration the sequence will stop.

2.11.6.3. Results

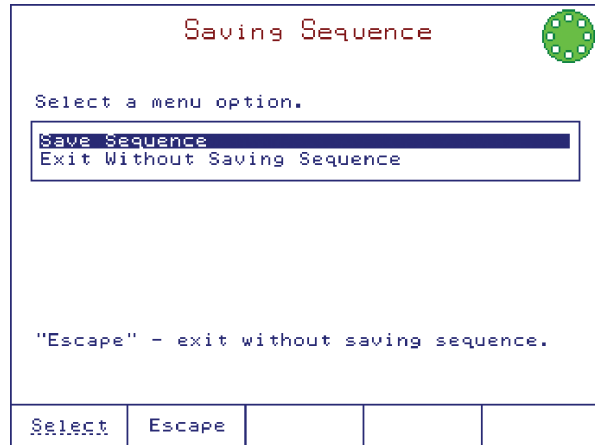
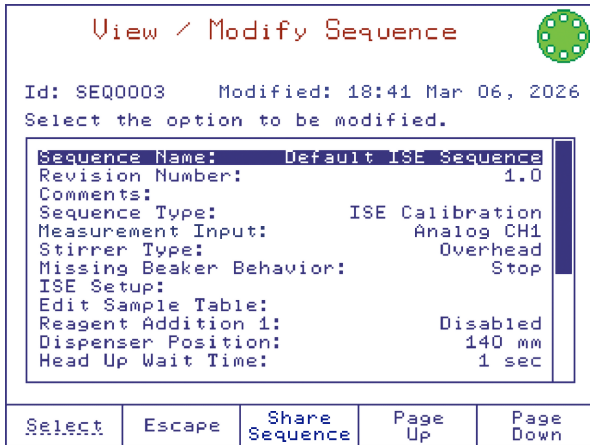
Press to view pH GLP Data or the key for more information.



2.11.7. ISE CALIBRATION

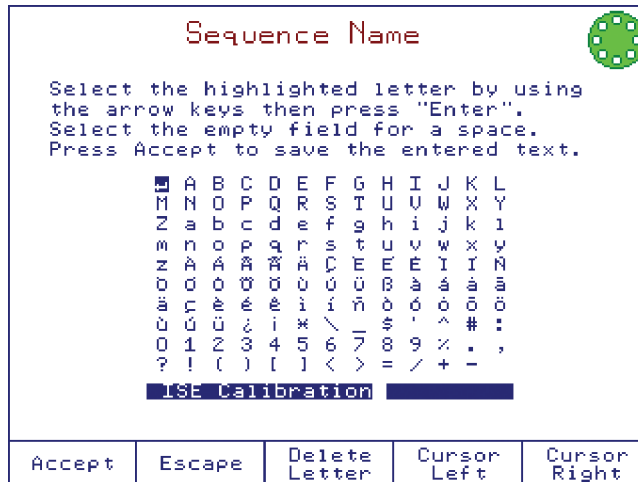
2.11.7.1. View / Modify Sequence

- To modify the sequence options, press **[Sequence Options]** from the main screen.
- A list of all the parameters for the selected sequence will be displayed.
- Use the **[Up]** and **[Down]** keys to highlight the option needed to be modified. Press **[Select]**.
- To exit the **View / Modify Sequence** screen, press **[Escape]**.
- Select to save the modifications or to discard them.



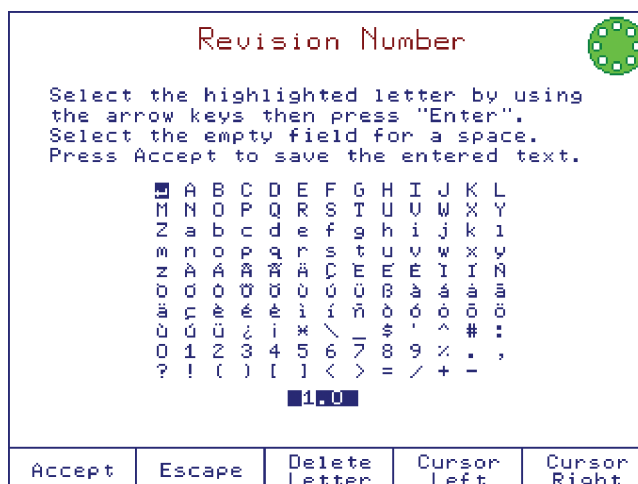
Sequence Name

Option: Up to 24 characters



Revision Number

Option: Up to 3 characters



Comments

Option: Up to 20 characters

Comments

Select the highlighted letter by using the arrow keys then press "Enter".
Select the empty field for a space.
Press Accept to save the entered text.

<input checked="" type="checkbox"/>	A	B	C	D	E	F	G	H	I	J	K	L
M	N	O	P	Q	R	S	T	U	V	W	X	Y
Z	a	b	c	d	e	f	g	h	i	j	k	l
m	n	o	p	q	r	s	t	u	v	w	x	y
z	À	Á	Â	Ã	Ä	Å	Æ	Ç	È	É	Ê	Ë
Ì	Í	Î	Ï	Ñ	Ò	Ó	Ô	Õ	Ö	×	Ù	Ú
Û	Ü	Ý	ÿ	ı	*	\	_	€	'	^	#	:
0	1	2	3	4	5	6	7	8	9	>	<	,
?	!	()	[]	<	>	=	/	+	-	

█

Accept	Escape	Delete Letter	Cursor Left	Cursor Right
--------	--------	---------------	-------------	--------------

Sequence Type

Option: Sample Analysis, pH Calibration, ISE Calibration

View / Modify Sequence

Id: SEQ0003 Modified: 18:41 Mar 06, 2026

Select the option to be modified.

Sequence Name: Default ISE Sequence

Revision Number: 1.0

Comments:

Sequence Type: ISE Calibration

Measurement Input: 1

Stirrer Type: d

Missing Beaker Behavior: p

ISE Setup:

Edit Sample Table: Disabled

Reagent Addition 1: 140 mm

Dispenser Position: 1 sec

Head Up Wait Time: 1 sec

Select	Escape			
--------	--------	--	--	--

Measurement Input

Option: Analog CH1 or Analog CH2 (if installed)

View / Modify Sequence

Id: SEQ0003 Modified: 18:41 Mar 06, 2026

Select the option to be modified.

Sequence Name: Default ISE Sequence

Revision Number: 1.0

Comments:

Sequence Type: ISE Calibration

Measurement Input: Analog CH1

Stirrer Type: d

Missing Beaker Behavior: p

pH Setup:

Edit Sample Table: Disabled

Dispenser Position: 140 mm

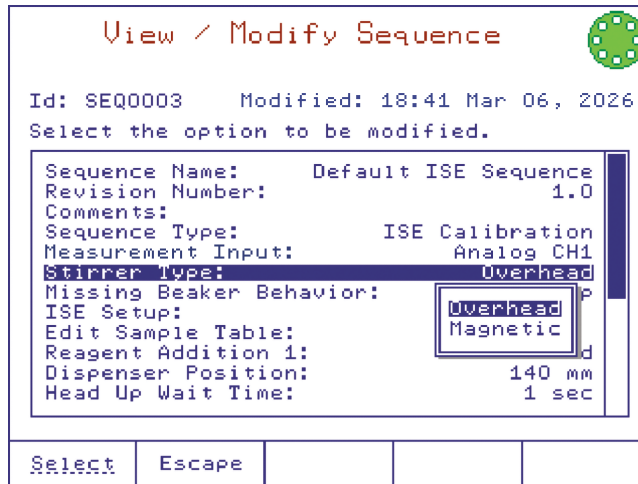
Head Up Wait Time: 1 sec

Aspirate Sample: Disabled

Select	Escape			
--------	--------	--	--	--

Stirrer Type

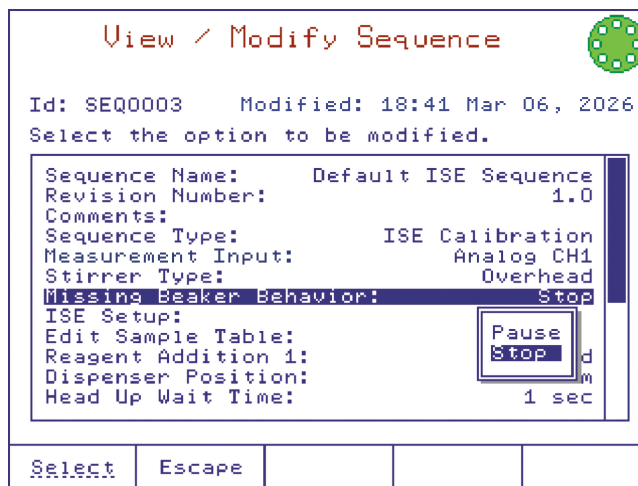
Option: Overhead or Magnetic



Missing Beaker Behavior

Option: Pause or Stop

Select type of behaviour when beaker detection is enabled and no beaker is detected.

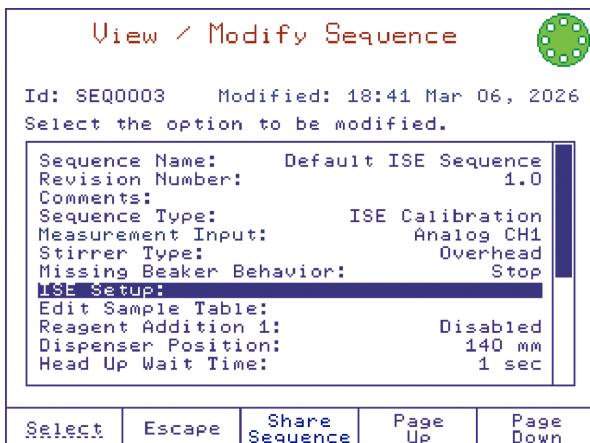


Pause The autosampler will pause the sequence at the current beaker and wait for the user before continuing the analysis.

Stop All operations will stop and the analysis will be stopped.

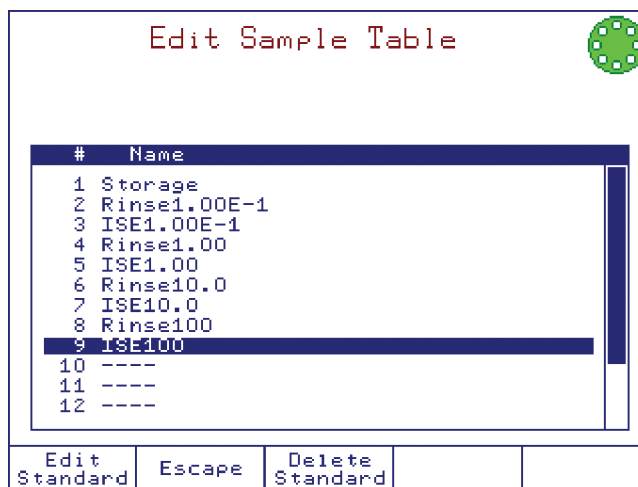
ISE Setup

This option allows the user to select ISE Setup menu. See [2.8.2. ISE Setup](#) section for more information.

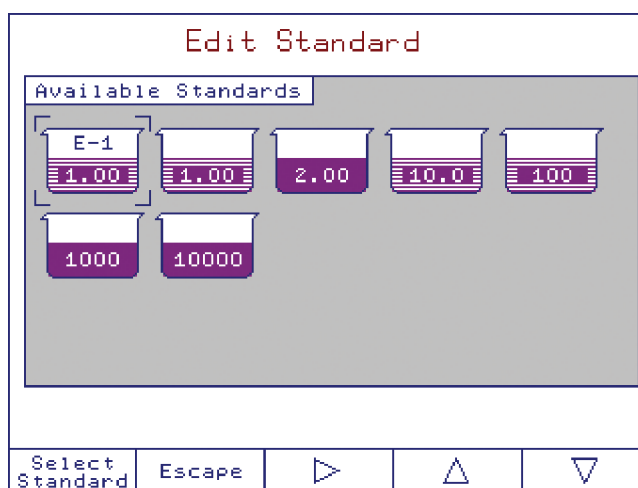


Edit Sample Table

This option allows to add, remove and edit the calibration beakers in the tray.



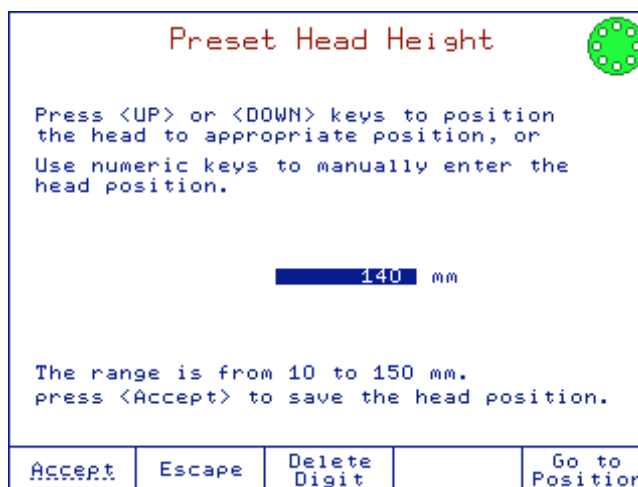
Note: Up to five calibration beakers can be added. Highlighted beakers have already been added to the calibration sequence.



Dispenser Position

Option: 10 to 150 mm

User can select the height for the dispenser head to be positioned at during titration. The default options is 140 mm.



Head Up Wait Time

Option: 1 to 30 seconds

User can set the time for the autosampler to wait, with the dispenser head in the up position to catch any residual droplets, before moving on to the next sample or rinse beaker.

Head Up Wait Time

Please enter the wait time in seconds.

2 sec

Low limit: 1 second
High limit: 30 seconds

Accept Escape Delete Digit

Sample Aspiration

Option: Disabled, Aspirate Only, Aspirate/Spray Rinse

Reacted samples may be aspirated into a waste container after each titration.

Note: Sample aspiration requires one peristaltic pump to be configured for aspiration, using [HI920-203 Tubing Set for Aspiration](#).

Aspirate Sample

Select the option to be modified.

Aspiration Option: Disabled

Disabled
Aspirate Only
Aspirate/Spray Rinse

Select Escape

Select the aspiration mode.

Aspirate Only

The existing waste from the sample beaker will be removed according to the parameters defined in this menu.

Aspirate/Spray Rinse

Reserved for future

Aspiration Pump

Option: Disabled, Auxiliary Pump 1, Auxiliary Pump 2, Auxiliary Pump 3

Select the peristaltic pump that is connected to the aspiration tube.

Aspirate Sample				
Select the option to be modified.				
Aspiration Option:	Aspirate Only			
Aspiration Pump:	Disabled			
Aspiration Time:				
Head Height:				
				<ul style="list-style-type: none"> Disabled Aux Pump 1 Aux Pump 2 Aux Pump 3
Select	Escape			

Aspiration Time

Option: 1 to 300 seconds

Set the duration that the peristaltic pump will run.

Aspiration Time				
Enter the period of time for running auxiliary pump.				
<div style="border: 1px solid black; display: inline-block; padding: 2px;">15</div> sec				
Low limit: 1 second				
High limit: 300 seconds				
Accept	Escape	Delete Digit		

Dispenser Head Height

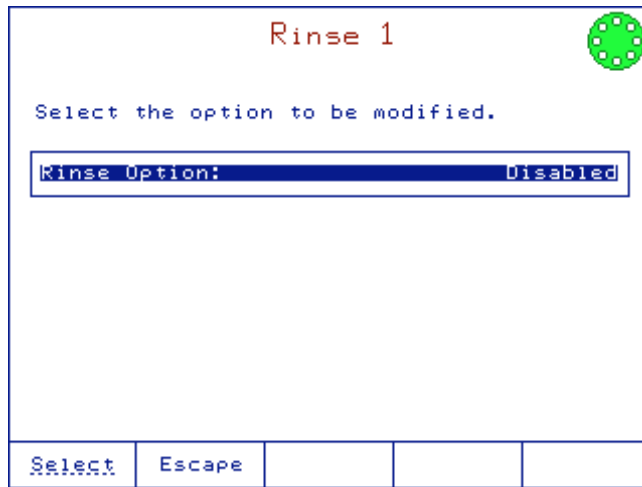
Option: 10 to 150 mm

Set the height for the dispenser head. The aspiration tube should be positioned such that it reaches the bottom of the sample beaker when the dispenser head is positioned in the range of 10 to 150 mm.

Preset Head Height				
Press <UP> or <DOWN> keys to position the head to appropriate position, or Use numeric keys to manually enter the head position.				
<div style="border: 1px solid black; display: inline-block; padding: 2px;">140</div> mm				
The range is from 10 to 150 mm. press <Accept> to save the head position.				
Accept	Escape	Delete Digit		Go to Position

Rinse

The autosampler can perform a dip rinse function after each analysis.
 Up to three dip rinses can be performed in a dedicated rinse beaker dip-rinse function.

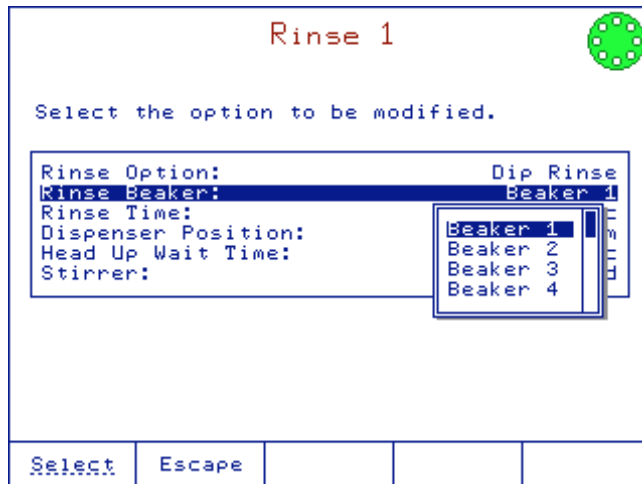


Dip rinse Dip rinse option can be used after each analysis to clean the electrodes and stirrer of contaminants, using dedicated rinsing beakers.

Spray Rinse Reserved for future

Rinse Beaker

Select the tray position for the dedicated rinse beaker.



Note: If the selected beaker is already in use for buffer, standard or calibration rinse, it will be removed from the sample table.

Rinse Time

Option: 1 to 300 seconds

Rinse Time				
Enter the period of time you would like to remain in the rinse beaker.				
10 sec				
Low limit: 1 second High limit: 300 seconds				
ACCEPT	Escape	Delete Digit		

Dispenser Position

Option: 10 to 150 mm

Set the height for the dispenser head during rinsing.

Preset Head Height				
Press <UP> or <DOWN> keys to position the head to appropriate position, or Use numeric keys to manually enter the head position.				
140 mm				
The range is from 10 to 150 mm. press <Accept> to save the head position.				
ACCEPT	Escape	Delete Digit		Go to Position

Head Up Wait Time

Option: 1 to 300 seconds

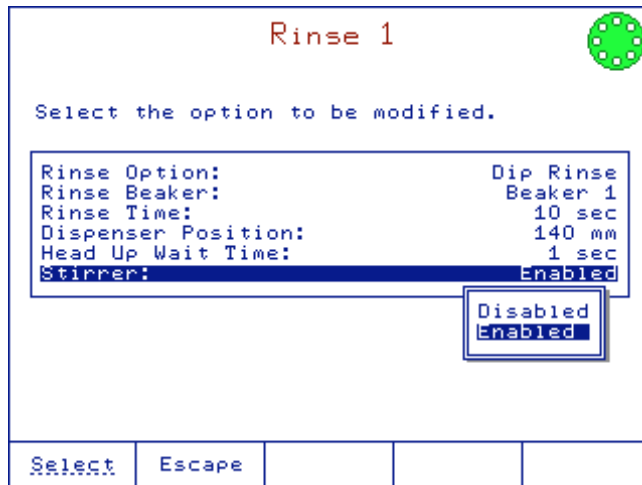
Set the duration that the autosampler will wait with the dispenser in the up position for any drops of solution to fall off of the electrodes / stirrer before moving to another sample or rinse beaker.

Head Up Wait Time				
Please enter the stirring time in seconds.				
1 sec				
Low limit: 1 second High limit: 300 seconds				
ACCEPT	Escape	Delete Digit		

Stirrer

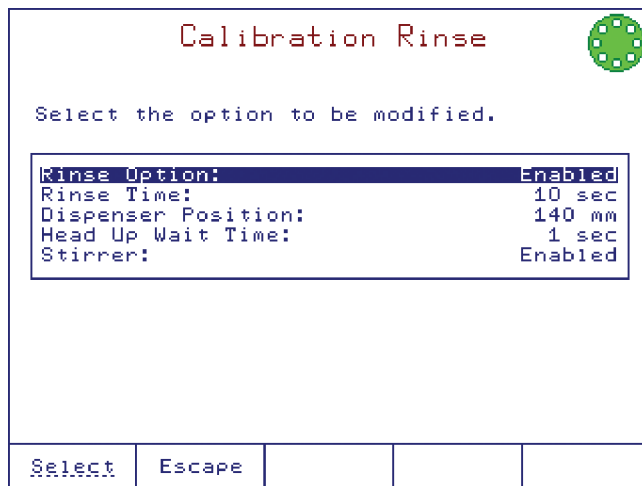
Option: Enabled or Disabled

Select if the stirrer will run during rinsing.



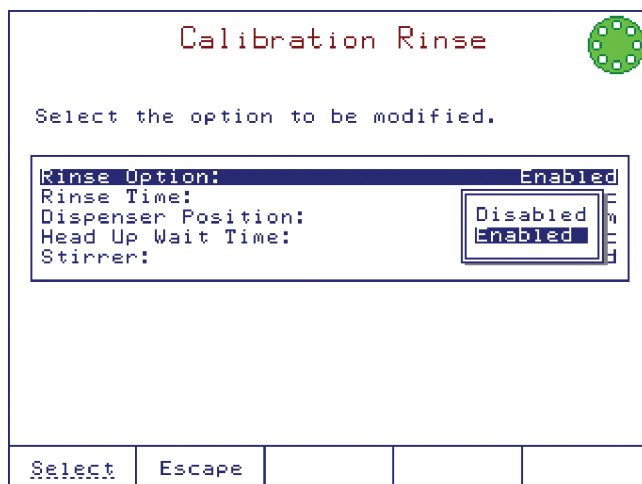
Calibration Rinse

The user can select to enable the rinse option during ISE calibrations.



Rinse Option

Option: Enabled or Disabled



Rinse Time


Option: 1 to 300 seconds

Rinse Time 				
Enter the period of time you would like to remain in the rinse beaker.				
<div style="border: 1px solid black; display: inline-block; padding: 2px;">10</div> sec				
Low limit: 1 second High limit: 300 seconds				
ACCEPT	Escape	Delete Digit		

Dispenser Position

Option: 10 to 150 mm


Set the height for the dispenser head during rinsing.

Preset Head Height 				
Press <UP> or <DOWN> keys to position the head to appropriate position, or Use numeric keys to manually enter the head position.				
<div style="border: 1px solid black; display: inline-block; padding: 2px;">140</div> mm				
The range is from 10 to 150 mm. press <Accept> to save the head position.				
ACCEPT	Escape	Delete Digit		Go to Position

Head Up Wait Time

Option: 1 to 300 seconds

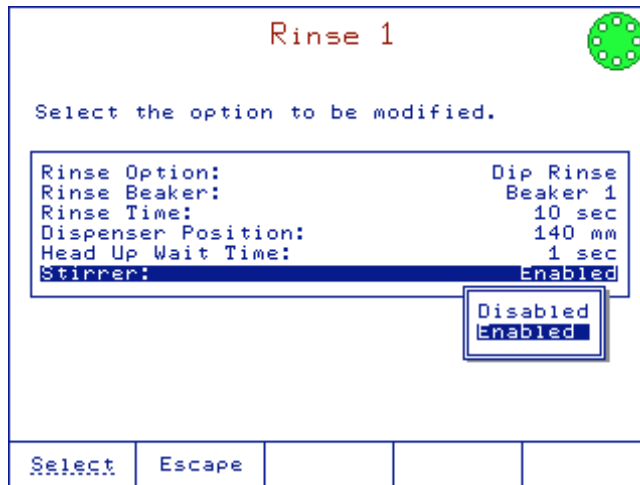
Set the duration that the autosampler will wait with the dispenser in the up position for any drops of solution to fall off of the electrodes or stirrer before moving to another sample or rinse beaker.

Head Up Wait Time 				
Please enter the stirring time in seconds.				
<div style="border: 1px solid black; display: inline-block; padding: 2px;">1</div> sec				
Low limit: 1 second High limit: 300 seconds				
ACCEPT	Escape	Delete Digit		

Stirrer

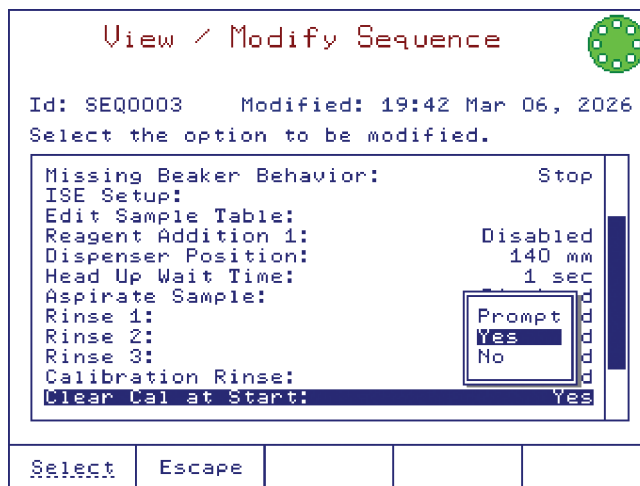
Option: Enabled or Disabled

Select if the stirrer will run during rinsing.



Clear Calibration at Start

Option: Prompt, Yes, No



Prompt Prompts the user to clear the calibration when the sequence starts

Yes Always clear the old calibration

No The new calibration data will be merged with the old data

Minimum Calibration Stability Time

Option: 1 to 600 seconds

Min. Cal Stability Time				
Please enter the minimum calibration stability time in seconds.				
1 sec				
Low limit: 1 second High limit: 600 seconds				
Accept	Escape	Delete Digit		

Beaker Height

Option: 30 to 120 mm

Set the height of the beaker being used on the autosampler.

Beaker Height				
Enter the beaker height in mm.				
100 mm				
Low limit: 30 mm High limit: 120 mm				
Accept	Escape	Delete Digit		

Position when Finished

Option: Home, Standard, Storage

View / Modify Sequence																												
Id: SEQ0003 Modified: 16:31 Mar 25, 2026																												
Select the option to be modified.																												
<table border="1"> <tbody> <tr><td>Reagent Addition 1:</td><td>Disabled</td></tr> <tr><td>Dispenser Position:</td><td>140 mm</td></tr> <tr><td>Head Up Wait Time:</td><td>1 sec</td></tr> <tr><td>Aspirate Sample:</td><td>Disabled</td></tr> <tr><td>Rinse 1:</td><td>Disabled</td></tr> <tr><td>Rinse 2:</td><td>Disabled</td></tr> <tr><td>Rinse 3:</td><td>Disabled</td></tr> <tr><td>Calibration Rinse:</td><td>Home</td></tr> <tr><td>Clear Cal at Start:</td><td>Standard</td></tr> <tr><td>Min. Cal Stability Time:</td><td>Storage</td></tr> <tr><td>Beaker Height:</td><td></td></tr> <tr><td>Position when finished:</td><td>Storage</td></tr> </tbody> </table>					Reagent Addition 1:	Disabled	Dispenser Position:	140 mm	Head Up Wait Time:	1 sec	Aspirate Sample:	Disabled	Rinse 1:	Disabled	Rinse 2:	Disabled	Rinse 3:	Disabled	Calibration Rinse:	Home	Clear Cal at Start:	Standard	Min. Cal Stability Time:	Storage	Beaker Height:		Position when finished:	Storage
Reagent Addition 1:	Disabled																											
Dispenser Position:	140 mm																											
Head Up Wait Time:	1 sec																											
Aspirate Sample:	Disabled																											
Rinse 1:	Disabled																											
Rinse 2:	Disabled																											
Rinse 3:	Disabled																											
Calibration Rinse:	Home																											
Clear Cal at Start:	Standard																											
Min. Cal Stability Time:	Storage																											
Beaker Height:																												
Position when finished:	Storage																											
Select	Escape																											

Home

The dispenser head will be in the up positioned above beaker one.

Standard

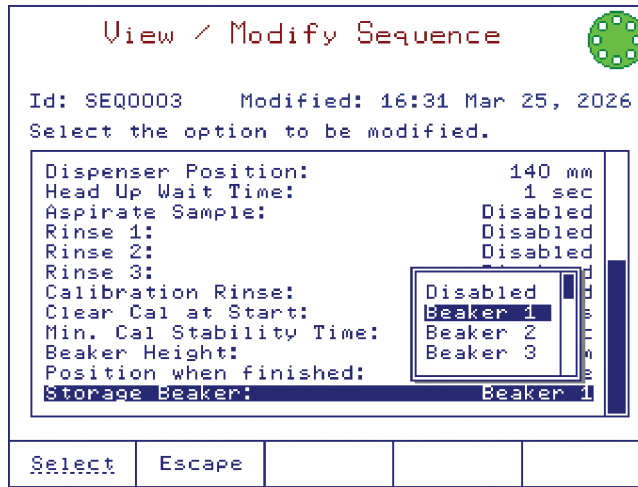
The dispenser head will remain down in the last standard that was analyzed / titrated.

Storage

The dispenser head will be down in a preset beaker containing storage solution.

Storage Beaker (Position when finished, storage only)

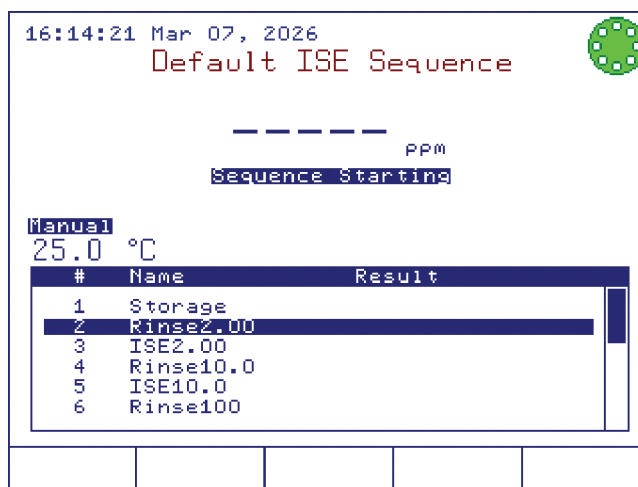
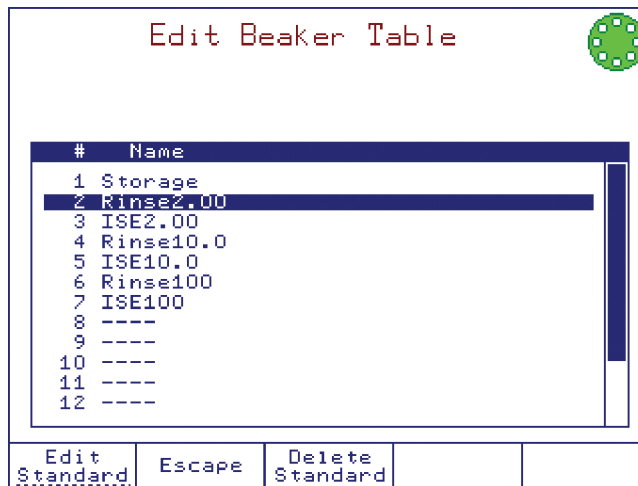
After the sequence has been completed, the autosampler will move to this position automatically and lower the dispenser head.



2.11.7.2. Running ISE Calibration Sequence

The sequence can be started by pressing the  key.

Note: When the Calibration Rinse option is enabled, the beaker used for rinsing is positioned before beaker used for buffer calibration.




The autosampler will process each calibration point according to the settings in sequence options.

While the autosampler is running, the top part of the screen shows the calibration information, and the bottom part of the screen shows a portion of the sample table.

The sample in progress is marked with symbol in the sample table. Use the and keys to scroll the sample table.

Press  to pause the current calibration. Press  to stop the calibration.

16:14:29 Mar 07, 2026 

Default ISE Sequence


----- PPM

Sequence Suspended

Manual
25.0 °C

#	Name	Result
1	Storage	
2	Rinse2.00	Suspended
3	ISE2.00	
4	Rinse10.0	
5	ISE10.0	
6	Rinse100	

Resume

16:14:44 Mar 07, 2026 

Default ISE Sequence

----- PPM

Manually Terminated


Manual
25.0 °C

2.00

#	Name	Result
1	Storage	
2	Rinse2.00	Stopped
3	ISE2.00	Rns. Stopped
4	Rinse10.0	
5	ISE10.0	
6	Rinse100	

Suspend

At the end of the sequence "Sequence Completed" will be displayed. The "Result" column will display the status for each beaker.

16:50:32 Mar 07, 2026 TRAY0029 

Default ISE Sequence



Sequence Completed

#	Name	Result
1	Storage	
2	Rinse2.00	Rinsed
3	ISE2.00	Calibrated
4	Rinse10.0	Rinsed
5	ISE10.0	Calibrated
6	Rinse100	Rinsed
7	ISE100	Calibrated
8	----	
9	----	
10	----	
11	----	
12	----	

View GLP. Clear Results AutoSmp. Setup

Note: If errors will appear during calibration the sequence will stop.

2.11.7.3. Results

Press  to view ISE GLP Data or the  key for more information.

ISE GLP Data

Analog CH2
 Last Calibration: 16:50 Mar 07, 2026
 Slope: 100.1% ISE: Iodide

2.00 ppm,	73.6mV	25.0°C	M
16:48:01 Mar 07, 2026			
10.0 ppm,	32.3mV	25.0°C	M
16:49:17 Mar 07, 2026			
100 ppm,	-27.1mV	25.0°C	M
16:50:32 Mar 07, 2026			

ESCAPE

Review Result

TRAY0036.RPT

2 Rinse2.00	Rinsed	16:47:01
3 ISE2.00	Calibrated	16:47:26
4 Rinse10.0	Rinsed	16:48:17
5 ISE10.0	Calibrated	16:48:42
6 Rinse100	Rinsed	16:49:32
7 ISE100	Calibrated	16:49:57

GLP Data

Last Calibration: 16:50 Mar 07, 2026
 Average Slope: 100.1%
 ISE: Iodide

2.00 ppm	73.6mV	25.0°C	M
----------	--------	--------	---

ESCAPE Share Report Page Up Page Down

2.11.8. SHARING OPTIONS IN AUTOSAMPLER MODE

Autosampler sequences, method parameters, tray reports, and meter information can be printed via USB, shared over Ethernet using FTP /FTPS or SMB; or saved to a USB storage device.

Files can be sent as raw text formats (.txt or .rpt) or converted to PDF or signed PDF format (if a valid signature file has been imported). Titration reports can be compiled into a CSV summary file that can be transferred over FTP/FTPS or SMB.

2.11.8.1. Share Sequence

When in *Default Sample Sequence* screen press .

Press  then select sharing type (print, FTP or SMB transfer, raw text or pdf file format, or raw text via USB flash drive).

View / Modify Sequence

Id: SEQ0004 Modified: 14:46 Mar 24, 2026
 Select the option to be modified.

Sequence Name: copy of Default Sample S	
Revision Number:	1.0
Comments:	
Sequence Type:	Sample Analysis
Stirrer Type:	Overhead
Missing Beaker Behavior:	Stop
Sample Leveling:	Aux Pump 1, 1 sec
Reagent Addition 1:	Disabled
Reagent Addition 2:	Disabled
Method:	HI0001EN, 0.1N Sodium Hydroxide
Method Options:	
Dispenser Position:	140 mm

Select Escape Share Sequence Page Up Page Down

Share


Select the menu option:

Print
As Raw over FTP
As Raw over SMB
As PDF over FTP
As PDF over SMB
As Signed pdf over FTP
As Signed pdf over SMB
As Raw to USB flash drive

Select Escape View Summary

2.11.8.2. Share Method

From main screen press , highlight *Method Options* then press .

Perform desired changes on selected method and press .

View/Modify Method

Id: USER0008 Modified: 17:55 Mar 12, 2026
Select the option to be modified.

Name:	copy of 0.1N Sodium Hydr
Method Revision:	1.0
Analysis Type:	Standard Titration
Measurement Input:	Analog CH1
Stirrer Configuration	
Titrant pump:	Pump 1
Reagent Addition 1:	Disabled
Reagent Addition 2:	Disabled
Dosing Type:	Dynamic
End Point Mode:	pH 1EQ point,1st Der
Recognition Options	
Pre-Titration Volume:	5.000 mL
Pre-Titration Stir Time:	60 sec
Measurement Mode:	Signal Stability

Select Escape Share Method Page Up Page Down

Share

Select the menu option:

Print
As Raw over FTP
As Raw over SMB
As PDF over FTP
As PDF over SMB
As Signed pdf over FTP
As Signed pdf over SMB
As Raw to USB flash drive

Select Escape View Summary

2.11.8.3. Share Tray Report

From main screen press  key and the **Data Parameters** screen will be displayed.

Highlight *Review Available Tray Report* and press .

Data Parameters

Select a menu option.

Review Last Tray Reports
Review Available Tray Report
GLP Data
Autosampler Information
Setup pH/mV/ISE Report
Setup Titration Report
Setup Calibration Report



Select Escape

Available Tray Reports

Highlight a Tray Result and press <Select> to see the tray reports.

Direct Reading Sequence	TRAY0033
9 analysis	13:42 May 26, 2026
Direct Reading Sequence	TRAY0032
27 analysis	11:41 May 26, 2026
Linked titrations	TRAY0018
6 analysis	11:02 May 25, 2026
Back Sequence	TRAY0017
4 analysis	10:49 May 25, 2026
Multiple eq points	TRAY0016
3 analysis	10:37 May 25, 2026
Mixed Sequence	TRAY0015
3 analysis	10:29 May 25, 2026
Default ISE Sequence	TRAY0014
ISE Calibration	10:20 May 25, 2026

Select Escape Share Report Share List Delete


- Press  to share the list of available tray reports as raw text, pdf, or signed pdf format.
- Press  to share reports from a tray.

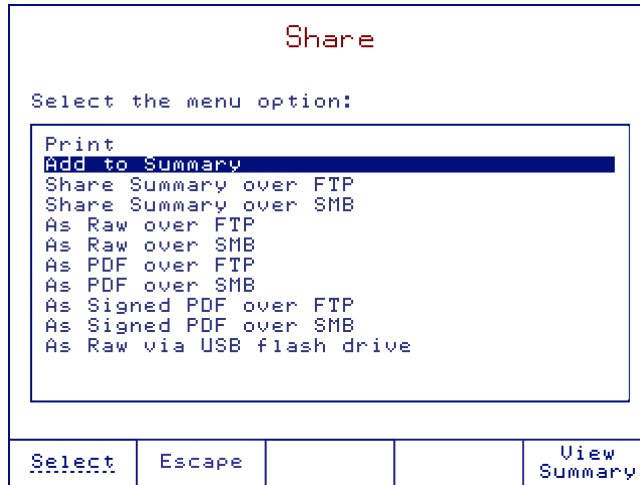
Analysis Reports

Sequence Name: Back Sequence
Tray ID: TRAY0017
Date & Time: 10:49 May 25, 2026

#	Sample Name	Result	Report Id
1	Chlorine ORP1	0.46975	Ti_11701
2	Chlorine ORP2	0 ppt (g/kg)	Ti_11702
8	Chlorine ORP8	0.25000	Ti_11703
11	Chlorine OR11	0.15591	Ti_11704

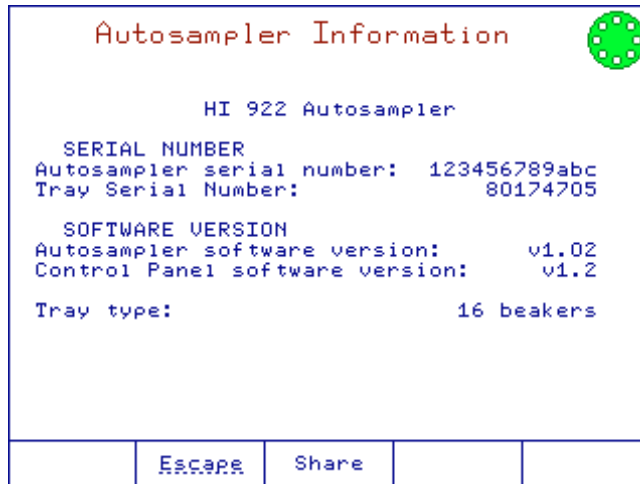
View Graph Escape View Report Share Report Delete Report

- » Press  to share the selected report from the tray.
 Select *Add to summary* option to have all reports ready for share in a single .csv file.
 Summary transfer is allowed via FTP/FTPS or SMB.

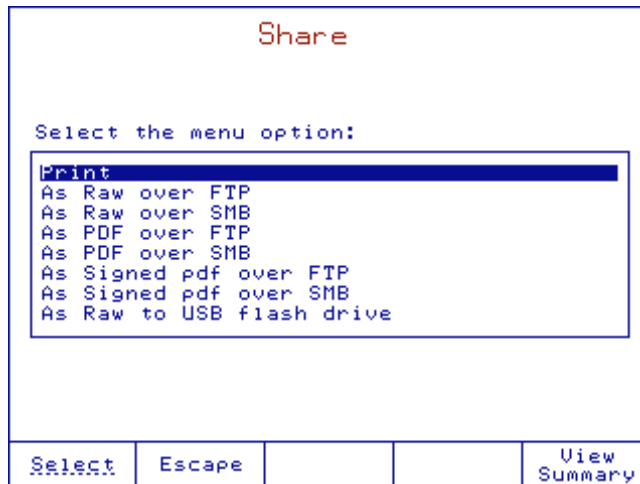


2.11.8.4. Share Autosampler Information

From main screen press  key.
 Highlight *Autosampler Information* and press .



Press  and select sharing type.



2.12. ACCESSORIES

2.12.1. SOLUTIONS

2.12.1.1. pH Calibration Buffers

Ordering Information	Description
HI7001M	pH 1.68 buffer solution, 230 mL
HI7001L	pH 1.68 buffer solution, 500 mL
HI7004M	pH 4.01 buffer solution, 230 mL
HI7004L	pH 4.01 buffer solution, 500 mL
HI7006M	pH 6.86 buffer solution, 230 mL
HI7006L	pH 6.86 buffer solution, 500 mL

Ordering Information	Description
HI7007M	pH 7.01 buffer solution, 230 mL
HI7007L	pH 7.01 buffer solution, 500 mL
HI7009M	pH 9.18 buffer solution, 230 mL
HI7009L	pH 9.18 buffer solution, 500 mL
HI7010M	pH 10.01 buffer solution, 230 mL
HI7010L	pH 10.01 buffer solution, 500 mL

2.12.1.2. pH Calibration Buffers in FDA Approved Bottle

Ordering Information	Description
HI8004L	pH 4.01 buffer solution, 500 mL
HI8006L	pH 6.86 buffer solution, 500 mL
HI8007L	pH 7.01 buffer solution, 500 mL

Ordering Information	Description
HI8009L	pH 9.18 buffer solution, 500 mL
HI8010L	pH 10.01 buffer solution, 500 mL

2.12.1.3. pH Technical Calibration Buffers

Ordering Information	Description
HI5016	pH 1.68 buffer solution, 500 mL
HI5003	pH 3.00 buffer solution, 500 mL
HI5004	pH 4.01 buffer solution, 500 mL
HI5068	pH 6.86 buffer solution, 500 mL

Ordering Information	Description
HI5007	pH 7.01 buffer solution, 500 mL
HI5091	pH 9.18 buffer solution, 500 mL
HI5010	pH 10.01 buffer solution, 500 mL
HI5124	pH 12.45 buffer solution, 500 mL

2.12.1.4. pH Millesimal Calibration Buffers

Ordering Information	Description
HI6016	pH 1.679 buffer solution, 500 mL
HI6003	pH 3.000 buffer solution, 500 mL
HI6004	pH 4.010 buffer solution, 500 mL
HI6068	pH 6.862 buffer solution, 500 mL

Ordering Information	Description
HI6007	pH 7.010 buffer solution, 500 mL
HI6091	pH 9.177 buffer solution, 500 mL
HI6010	pH 10.010 buffer solution, 500 mL
HI6124	pH 12.450 buffer solution, 500 mL

2.12.1.5. Electrode Cleaning Solutions

Ordering Information	Description
HI7061M	General purpose cleaning solution, 230 mL
HI7061L	General purpose cleaning solution, 500 mL
HI7073M	Protein cleaning solution, 230 mL
HI7073L	Protein cleaning solution, 500 mL
HI7074M	Inorganic cleaning solution, 230 mL
HI7074L	Inorganic cleaning solution, 500 mL
HI7077M	Oil & fat cleaning solution, 230 mL
HI7077L	Oil & fat cleaning solution, 500 mL

2.12.1.6. Electrode Cleaning Solutions in FDA Approved Bottle

Ordering Information	Description
HI8061L	General purpose solution, 500 mL
HI8073L	Protein cleaning solution, 500 mL
HI8077L	Oil & fat cleaning solution, 500 mL

2.12.1.7. Electrode Storage Solutions

Ordering Information	Description
HI70300M	Storage solution, 230 mL
HI70300L	Storage solution, 500 mL

2.12.1.8. Electrode Storage Solutions in FDA Approved Bottle

Ordering Information	Description
HI80300M	Storage solution, 230 mL
HI80300L	Storage solution, 500 mL

2.12.1.9. Electrode Refill Electrolyte Solutions

Ordering Information	Description
HI7071	3.5 M KCl with AgCl reference electrolyte solution, 30 mL x 4
HI7072	1 M Potassium nitrate electrode fill solution, 30 mL x 4
HI7075	1.7 M Potassium nitrate, 0.7 M potassium chloride electrode fill solution, 30 mL x 4
HI7076	1 M Sodium chloride electrode fill solution, 30 mL x 4
HI7078	0.5 M Ammonium sulfate electrode fill solution, 30 mL x 4
HI7082	3.5 M KCl reference electrolyte solution, 30 mL, 30 mL x 4
HI7094	1M LiCl in ethanol fill solution, 30 mL x 4

2.12.1.10. Electrode Refill Electrolyte Solutions in FDA Approved Bottle

Ordering Information	Description
HI8071	3.5 M KCl with AgCl reference electrolyte solution, 30 mL
HI8082	3.5 M KCl reference electrolyte solution, 30 mL

2.12.1.11. ORP Pretreatment Solutions

Ordering Information	Description
HI7091L	Reducing pretreatment solution, 500 mL
HI7092M	Oxidizing pretreatment solution, 230 mL
HI7092L	Oxidizing pretreatment solution, 500 mL

2.12.1.12. Titration Reagents

Ordering Information	Description
HI70429	0.05M Silver Nitrate titration reagent, 1 Litre
HI70433	0.01N Stabilized iodine titration reagent, 1 Litre
HI70439	0.1M Sodium Thiosulfate titration reagent, 1 Litre
HI70440	0.02N Stabilized Iodine titration reagent, 1 Litre
HI70441	0.04N Stabilized Iodine titration reagent, 1 Litre
HI70448	0.02M Silver Nitrate titration reagent, 1 Litre
HI70449	0.02M EDTA titration reagent, 1 Litre
HI70455	0.01N Sodium Hydroxide titration reagent, 1 Litre
HI70456	0.1N Sodium Hydroxide titration reagent, 1 Litre
HI70457	1N Sodium Hydroxide titration reagent, 1 Litre
HI70458	0.01M Sulfuric Acid titration reagent, 1 Litre
HI70459	0.05M Sulfuric Acid titration reagent, 1 Litre
HI70462	0.01N Hydrochloric Acid titration reagent, 1 Litre
HI70463	0.1N Hydrochloric Acid titration reagent, 1 Litre
HI70464	1N Hydrochloric Acid titration reagent, 1 Litre

2.12.1.13. Ion-Selective Electrode Calibration Standards

Ordering Information	Description
HI4001-01	0.1 M Ammonia standard
HI4001-02	100 ppm Ammonia standard (as N)
HI4001-03	1000 ppm Ammonia standard (as N)
HI4002-01	0.1 M Bromide standard
HI4003-01	0.1 M Cadmium standard
HI4004-01	0.1 M Calcium standard
HI4005-01	0.1 M Carbon dioxide standard
HI4005-03	1000 ppm Carbon dioxide standard (as CaCO ₃)
HI4007-01	0.1 M Chloride standard
HI4007-02	100 ppm Chloride standard
HI4007-03	1000 ppm Chloride standard
HI4008-01	0.1 M Cupric standard
HI4010-01	0.1 M Fluoride standard
HI4010-02	100 ppm Fluoride standard
HI4010-03	1000 ppm Fluoride standard
HI4011-01	0.1 M Iodide standard
HI4012-01	0.1 M Lead standard
HI4012-21	0.1 M Sulfate standard
HI4013-01	0.1 M Nitrate standard
HI4013-02	100 ppm Nitrate standard
HI4013-03	1000 ppm Nitrate standard
HI4014-01	0.1 M Potassium standard
HI4015-01	0.1 M Silver standard

2.12.2. SENSORS

2.12.2.1. pH Electrodes

Ordering Information	Description
HI1043B	Glass-body, double junction, refillable, combination pH electrode Use: strong acid and base, paint and solvents
HI1053B	Glass-body, triple ceramic, conic shape, refillable, combination pH electrode Use: emulsions, fats and creams, soil and semi-solids samples
HI1083B	Glass-body, micro, viscolene, nonrefillable, combination pH electrode Use: biotechnology and micro titration
HI1131B	Glass-body, double junction, refillable, combination pH electrode Use: general purpose
HI1330B	Glass-body, semi-micro diameter, single junction, refillable, combination pH electrode Use: laboratory, vials, and test tubes
HI1331B	Glass-body, semi-micro diameter, single junction, refillable, combination pH electrode Use: flasks
HI1230B	Plastic-body (PEI), double junction, gel-filled, combination pH electrode Use: general purpose
HI2031B	Glass-body, conical tip, refillable, combination pH electrode. Use: dairy and semi-solid products
HI1332B	Plastic-body (PEI), double junction, refillable, combination pH electrode Use: chemicals, field applications and quality control testing
FC100B	Plastic-body (PVDF), double junction, refillable, combination pH electrode Use: cheese
FC200B	Plastic-body (PVDF), single junction, conical tip, non-refillable viscolene electrolyte, combination pH electrode. Use: milk, yogurt, dairy products, and semi-solid foods
FC210B	Glass-body, double junction, conical tip, non-refillable viscolene electrolyte, combination pH electrode. Use: milk, yogurt, and cream
FC220B	Glass-body, single junction, refillable, combination pH electrode Use: milk, yogurt, cream, sauce, and fruit juices
FC911B	Plastic-body (PVDF), double junction, refillable, combination pH electrode Use: sauce, juices, dairy products and other liquid or slurry forms of food
HI1413B	Glass-body, single junction, flat tip, non-refillable viscolene electrolyte, combination pH electrode Use: surfaces, skin, leather, paper, and emulsions

Digital pH Probes

Ordering Information	Description
HI10530	Triple ceramic, double junction, low temperature glass body, refillable pH electrode with conical tip and temperature sensor
HI10430	Triple ceramic, double junction, high temperature glass body, refillable pH electrode with temperature sensor
HI11310	Glass body, double junction, refillable pH/temperature electrode
HI11311	Glass body, double junction, refillable pH/temperature electrode with enhanced diagnostics
HI12300	Plastic body, double junction, gel filled, non refillable pH/temperature electrode

Ordering Information	Description
HI12301	Plastic body, double junction, gel filled, non refillable pH/temperature electrode with enhanced diagnostics
HI10480	Glass body, double junction with temperature sensor for wine analysis
FC2320	Double junction, open reference, non refillable, electrolyte viscolene, PVDF body with conical tip, pH/temperature electrode
FC2100	Double junction, open reference, non refillable, electrolyte viscolene, glass body with conical tip, pH/temperature electrode
FC2020	Double junction, open reference, non refillable, electrolyte viscolene, PVDF body with conical tip, pH/temperature electrode

2.12.2.2. ORP Electrodes

Ordering Information	Description
HI3131B	Glass-body, refillable, combination platinum ORP electrode Use: laboratories and general purpose
HI3230B	Plastic-body (PEI), gel-filled, combination platinum ORP electrode Use: municipal water and quality control
HI4430B	Plastic-body (PEI), gel-filled, combination gold ORP electrode Use: oxidants and ozone

Digital ORP Probes

Ordering Information	Description
HI36180	Glass body, double junction, refillable ORP/temperature probe
HI36200	Plastic body, single junction gel filled, non refillable ORP/temperature probe

2.12.2.3. Photometric Electrodes

Ordering Information	Description
HI900601	Photometric electrode with green LED (525 nm)
HI900602	Photometric electrode with red LED (625 nm)
HI900603	Photometric electrode with yellow LED (590 nm)
HI900604	Photometric electrode with blue LED (470 nm)

2.12.2.4. Half-Cell Electrodes

Ordering Information	Description
HI5311	Glass-body, silver/silver chloride (Ag/AgCl) reference half-cell electrode, double junction, refillable, with 4 mm banana plug, 1 m (3.3') cable Use: general purpose with wide temperature range
HI5315	Plastic-body (PEI), double junction, silver/silver chloride (Ag/AgCl) reference half-cell electrode, refillable, with 4 mm plug, 1 m (3.3') cable Use: Ion-Selective Electrodes
HI5412	Glass-body, single Calomel reference half-cell electrode, refillable, with 4mm plug, 1 m (3.3') cable Use: general purpose with constant temperature range

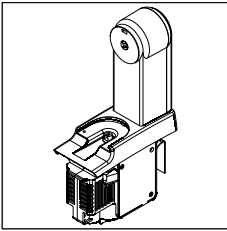
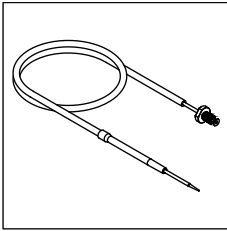
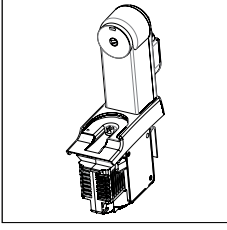
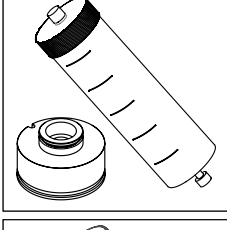
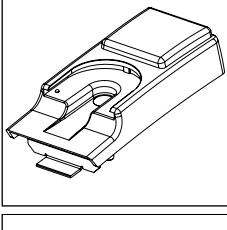
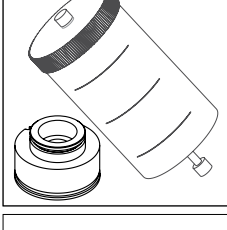
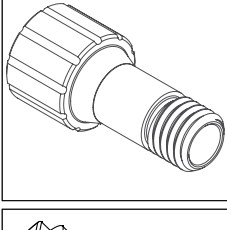
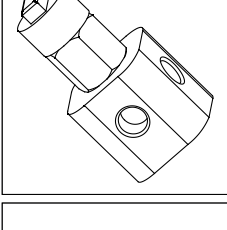
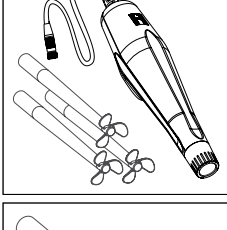
2.12.2.5. Ion-Selective Electrodes

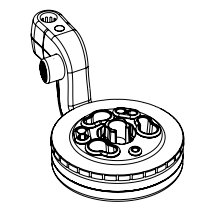
Ordering Information	Description
HI4101	Ammonia combination ion selective electrode
HI4002 / HI4102	Bromide combination ion selective electrode
HI4003 / HI4103	Cadmium half-cell ion selective electrode
HI4004 / HI4104	Calcium combination ion selective electrode
HI4105	Carbon dioxide combination ion selective electrode
HI4007 / HI4107	Chloride combination ion selective electrode
HI4008 / HI4108	Copper (cupric) combination ion selective electrode
HI4009 / HI4109	Cyanide combination ion selective electrode
HI4010 / HI4110	Fluoride combination ion selective electrode
HI4011 / HI4111	Iodide combination ion selective electrode
HI4012 / HI4112	Lead half-cell ion selective electrode
HI4013 / HI4113	Nitrate combination ion selective electrode
HI4014 / HI4114	Potassium combination ion selective electrode
HI4015 / HI4115	Silver / Sulfide combination ion selective electrode
FC300B	Sodium electrode

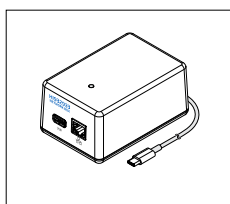
2.12.2.6. Temperature Sensor

Ordering Information	Description
HI7662-TW	Temperature probe with 1 m (3.3') paneled cable

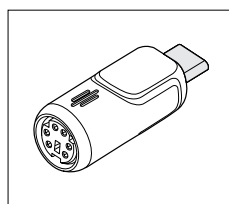
2.12.3. TITRATOR COMPONENTS

	HI930100 Pump assembly		HI930280 Dispensing tube with dispensing tip, fitting, protection tube and tube guide
	HI930101 Pump assembly with peristaltic pump		HI900205 Syringe, 5 mL
	Burette with: HI930105 - 5 mL syringe HI930110 - 10 mL syringe HI930125 - 25 mL syringe HI930150 - 50 mL syringe		HI900210 Syringe, 10 mL
	HI930191 Blank support		HI900225 Syringe, 25 mL
	HI930190 Blank burette support		HI900250 Syringe, 50 mL
	HI900942 Tool for burette cap removal		HI930160 Burette holder
	HI900260 Three-way valve		HI930301 Overhead stirrer and 3 propellers
	HI900270S Aspiration tube with fitting and protection tube		HI930302 Replacement propellers (3 pieces)

	<p>HI930303 High chemical resistance propellers (3 pieces)</p>		<p>HI930202 Titrator peristaltic pump complete tubing set</p>
	<p>HI930320 Overhead holder support rod</p>		<p>HI7662-TW Temperature probe</p>
	<p>HI930310 Overhead electrode holder</p>		<p>HI900945 Shorting cap</p>
	<p>HI930330 Titrant bottle cap assembly</p>		<p>HI900947 Power adapter (European plug)</p>
	<p>HI930311 Electrode adapter for overhead holder</p>		<p>HI900946 Power adapter (USA plug)</p>
	<p>HI930315 Titrant bottle holder</p>		<p>HI920018 USB-C to USB-C cable</p>
	<p>HI930201 Replacement cap and rotor for peristaltic pump</p>		<p>HI930901 USB flash drive</p>
	<p>HI930204 Roller tube for titrator peristaltic pump (3 pieces)</p>		<p>HI930812 Instruction manual binder for HI932 (version 2.2.2 and later)</p>

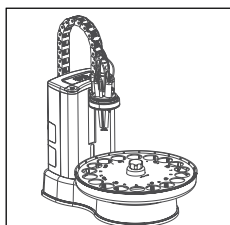


HI932933
Network box

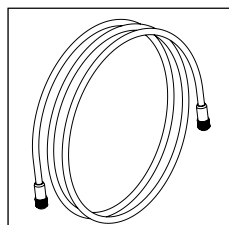


HI932934
USB-C to PS/2 adapter for
HI90060X photometric
electrodes use with HI932
(software version 2.2.2 and later)

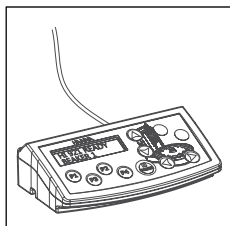
2.12.4. AUTOSAMPLER COMPONENTS



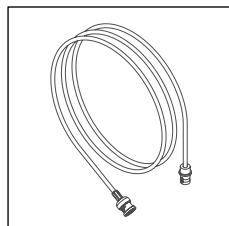
HI922-XYZ
Autosampler



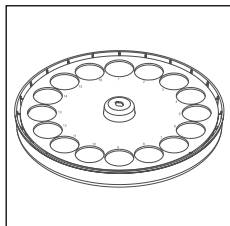
HI920-933
(HI932 to HI921/HI922)
Communication cable



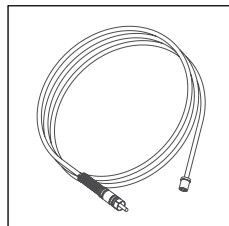
HI920-922
Control panel



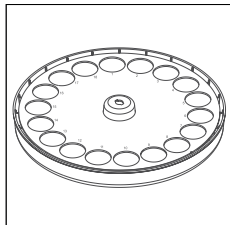
HI920-931
BNC Extension cable (1 m)



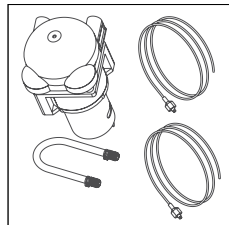
HI920-11660W
16 beaker tray, 60 mm diameter
Single row with RFID



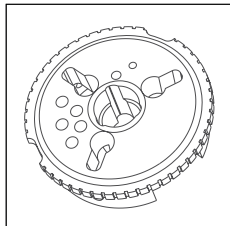
HI920-932
Reference extension cable (1 m)



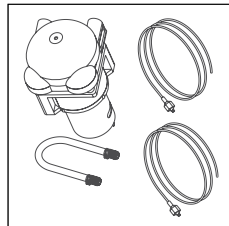
HI920-11853W
18 beaker tray, 53 mm diameter
Single row with RFID



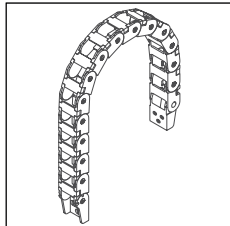
HI920-103
Peristaltic pump with dispensing
tubing



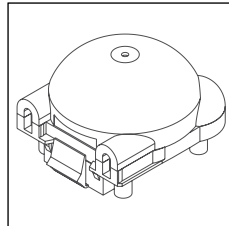
HI920-310
Electrode holder



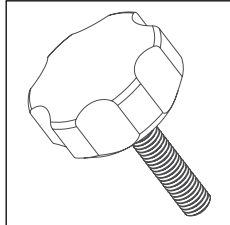
HI920-104
Peristaltic pump with aspiration
tubing



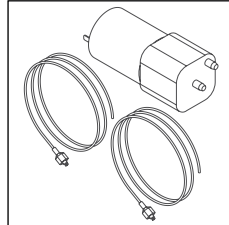
HI920-320
Cable chain



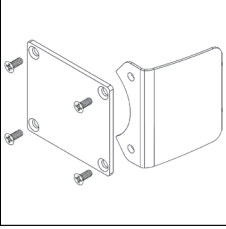
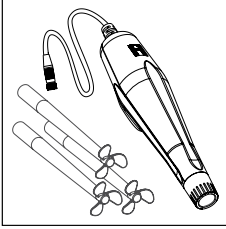
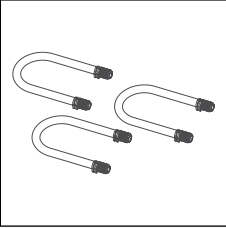
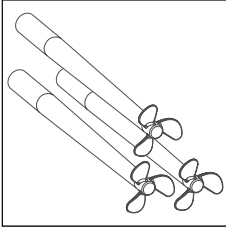
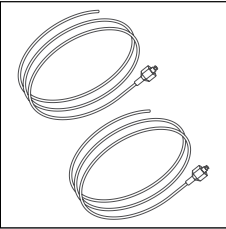
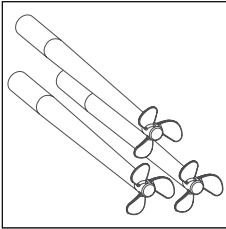
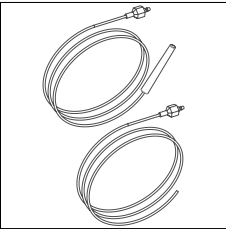
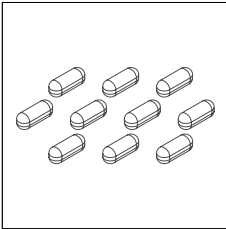
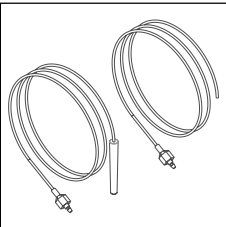
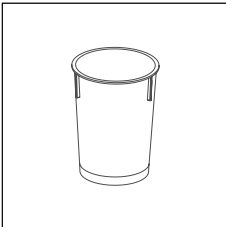
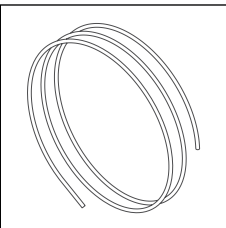
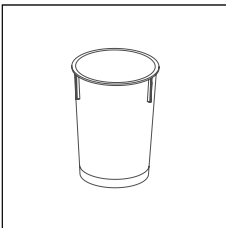
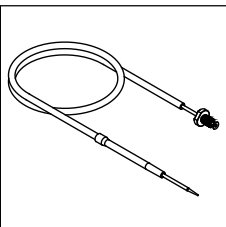
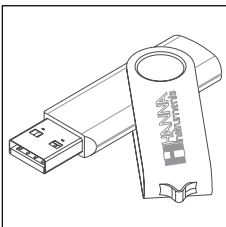
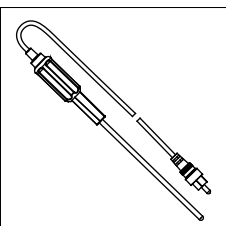
HI920-201
Replacement cap and rotor for
peristaltic pump



HI920-960
Tray locking screw



HI920-113
Membrane pump with tubing

	<p>HI920-191 Pump covers</p>		<p>HI930301 Overhead stirrer + 3 propellers</p>
	<p>HI920-204 Roller tube for autosampler peristaltic pump (3 pieces)</p>		<p>HI930302 Replacement propellers (3 pieces)</p>
	<p>HI920-212 Membrane pump complete tubing set</p>		<p>HI930303 High chemical resistance propellers (3 pieces)</p>
	<p>HI920-208 Tubing set with plastic dispensing tube guide for peristaltic pump</p>		<p>HI731319 Stir bars, 25 mm x 7 mm (10 pieces)</p>
	<p>HI920-203 Tubing set with stainless steel aspiration tube for peristaltic pump</p>		<p>HI920-053 Plastic beaker for HI920-11853 (20 pieces)</p>
	<p>HI920-290 TYGON Tube (5 m)</p>		<p>HI920-060 Plastic beaker for HI920-11660 (20 pieces)</p>
	<p>HI920-281 Titrant dispensing tube (1.5 m)</p>		<p>HI920-901 USB flash drive</p>
	<p>HI7662-AW Temperature sensor</p>		

PART 3. APPLICATIONS

HI0001EN – 0.1N SODIUM HYDROXIDE TITRANT CONCENTRATION

Description

Method for the standardization (titer determination) of 0.1N Sodium Hydroxide (NaOH) titrant solution against Potassium Hydrogen Phthalate (KHP). The results are expressed in **N (eq/L)**.

Reference

AOAC Official Methods of Analysis, Official Method 936.16

Electrode

- [HI1131B](#) Combination pH Electrode
- [HI7662-T](#) Temperature Probe

Reagents

- [HI70456](#) 0.1N Sodium Hydroxide (1 L)
- [HI70401](#) Potassium Hydrogen Phthalate (20 g)
- [HI70436](#) Deionized Water (1 gal)

Accessories

- [HI70300L](#) Storage Solution (500 mL)
- [HI7082](#) Electrode Fill Solution (30 mL × 4)
- [HI7004L](#) pH 4.01 Buffer Solution (500 mL)
- [HI7007L](#) pH 7.01 Buffer Solution (500 mL)
- [HI7010L](#) pH 10.01 Buffer Solution (500 mL)
- [HI920-053](#) 100-mL Plastic Beaker (20 pcs.)
- Analytical Balance with 0.0001 g resolution

Device Preparation

- Connect the pH electrode and temperature probe to the titrator.
- Install a 25 mL burette filled with 0.1N sodium hydroxide ([HI70456](#)) on pump one and verify that no air bubbles are present in the burette or tubing. If necessary prime the burette until all the air has been removed completely.
- Press HI0001EN 0.1N Sodium Hydroxide and press .

Electrode Preparation

- Press .
- Calibrate the electrode using pH 4.01, 7.01 and 10.01 buffers. Refer to the instruction manual for calibration procedure.

Sample Preparation

- Crush approximately 3 grams of potassium hydrogen phthalate ([HI70401](#)) and dry it for 2 hours at 120°C. Cool to room temperature in a desiccator.
- Place a clean 100 mL plastic beaker on the analytical balance.
- Zero the balance.
- Carefully weigh approximately 0.20 grams of dried potassium hydrogen phthalate into the beaker. Ensure that all of the potassium hydrogen phthalate is on the bottom of the beaker.
- Record the exact weight of the sample once the balance has stabilized with an accuracy of 0.0001 grams.
- Remove the beaker from the balance and add deionized water to the 50 mL mark on the beaker.

Analysis

- Place the beaker under the stirrer assembly and lower it to immerse the pH electrode, temperature probe and stirrer. Ensure that the reference junction of the pH electrode is 5 to 6 mm below the surface. If necessary add extra deionized water.

Note: The dispensing tip should be slightly submerged in the sample.

- Press start stop. You will be prompted to enter the weight of the analyte (weight of potassium hydrogen phthalate). Use the numeric keypad to enter the exact weight and press enter to start the analysis.

Note: Ensure that the potassium hydrogen phthalate dissolves completely during the pre-titration stir time. Erroneous results may occur if the sample does not dissolve completely prior to titration. If necessary the pre-titration stir time can be increased.

- At the end of the titration, after detection of the equivalence point, "Titration Completed" will appear with the result. The result is expressed in **N (eq/L) of sodium hydroxide**.
- Remove the pH electrode, temperature probe and stirrer from the sample and rinse them thoroughly with deionized water.
- Record the result.

Note: For improved accuracy, repeat this procedure a minimum of three times and calculate the average value.

For methods utilizing 0.1N sodium hydroxide titrant solution, follow the steps below to enter the titer/standardized value.

- Select the method utilizing 0.1N sodium hydroxide.
- Press Method Options from the main screen.
- Using the arrow keys, highlight Titrant Conc. and press Select.
- Use the numeric keypad to enter the standardized (titer) value of the titrant then press Accept.
- Press Escape to exit the **View/Modify Method** screen. Use the arrow keys to highlight **Save Method** and press Select.

METHOD PARAMETERS

Name: 0.1N Sodium Hydroxide
 Method Revision: 4.0
 Analysis Type: Standard Titration
 Measurement Input: Analog CH1
 Stirrer Configuration:
 Stirrer: Stirrer 1
 Stirring Speed: 1400 RPM
 Titrant Pump: Pump 1
 Reagent Addition 1: Disabled
 Reagent Addition 2: Disabled
 Dosing Type: Dynamic
 Min Vol: 0.030 mL
 Max Vol: 0.500 mL
 delta E: 4.500 mV
 End Point Mode: pH 1EQ point, 1st Der
 Recognition Options:
 Threshold: 500 mV/mL
 Range: NO
 Filtered Derivatives: NO
 Pre-Titration Volume: 5.000 mL
 Pre-Titration Stir Time: 60 sec
 Measurement Mode: Signal Stability
 delta E: 0.3 mV
 delta t: 2 sec
 Min wait: 3 sec
 Max wait: 30 sec
 Electrode Type: pH
 Blank Option: No Blank
 Calculations: Stdz. Titrant by Weight
 Dilution Option: Disabled
 Titrant Name: 0.1N NaOH
 Analyte Size: 0.20000 g
 Analyte Entry: Manual
 Maximum Titrant Volume: 15.000 mL
 Potential Range: -2000.0 to 2000.0 mV
 Volume/Flow Rate: 25 mL/50.0 mL/min
 Signal Averaging: 1 Reading
 Significant Figures: XXXXX

CALCULATIONS

Calculations: Stdz. Titrant by Weight
 Titrant units: N (eq/L)
 Titrant volume dosed: V (L)
 Standard weight: 0.200 g
 mw of standard: 204.23 g/mol
 Titrant/Standard: 1.000 eq/mol

$$\frac{\text{eq}}{\text{L}} \text{NaOH} = \frac{0.200 * 1.000}{204.23 * V (\text{L})}$$

RESULTS

Titration Report

Method Name: 0.1N Sodium Hydroxide
 Time & Date: 17:03 May 07, 2026
 Report ID: Ti_00053

Titration Results

Method Name: 0.1N Sodium Hydroxide
 Time & Date: 17:03 May 07, 2026
 Analyte Size: 0.20920 g
 End Point Volume: 10.215 mL
 pH Equivalence Point: 8.394
 Result: 0.10027 N(eq/L)
 Initial & Final pH: 4.173 to 9.570
 Titration Duration: 6:25 [mm:ss]
 Titration went to Completion:

Analyst Signature: _____

HI0002EN — 0.1N HYDROCHLORIC ACID TITRANT CONCENTRATION

Description

Method for the standardization (titer determination) of 0.1N Hydrochloric Acid (HCl) titrant solution against Sodium Hydroxide (NaOH). The results are expressed in **N (eq/L)**.

Reference

AOAC Official Methods of Analysis, Official Method 936.15

Electrode

- [HI1131B](#) Combination pH Electrode
- [HI7662-T](#) Temperature Probe

Reagents

- [HI70463](#) 0.1N Hydrochloric Acid (1 L)
- [HI70456](#) 0.1N Sodium Hydroxide (1 L)
- [HI70436](#) Deionized Water (1 gal)

Accessories

- [HI70300L](#) Storage Solution (500 mL)
- [HI7082](#) Electrode Fill Solution (30 mL × 4)
- [HI7004L](#) pH 4.01 Buffer Solution (500 mL)
- [HI7007L](#) pH 7.01 Buffer Solution (500 mL)
- [HI7010L](#) pH 10.01 Buffer Solution (500 mL)
- [HI920-053](#) 100-mL Plastic Beaker (20 pcs.)
- 10 mL Class A Volumetric Pipette

Device Preparation

- Connect the pH electrode and temperature probe to the titrator.
- Install a 25 mL burette filled with 0.1N hydrochloric acid ([HI70463](#)) on pump one and verify that no air bubbles are present in the burette or tubing.
If necessary prime the burette until all the air has been removed completely.
- Press from the main screen. Use the arrow keys to highlight [HI0002EN 0.1N Hydrochloric Acid](#) and press .

Electrode Preparation

- Press from the main screen.
- If necessary, select the measurement input and press .
- Calibrate the electrode using pH 4.01, 7.01, and 10.01 buffers.
Refer to the instruction manual for calibration procedure.


Sample Preparation

- Use a Class A volumetric pipette to transfer exactly 10.00 mL of 0.1N sodium hydroxide ([HI70456](#)) to a clean 100 mL beaker.
- Add deionized water to the 50 mL mark on the beaker.

Analysis






- Place the beaker under the stirrer assembly and lower it to immerse the pH electrode, temperature probe and stirrer.
- Ensure that the reference junction of the pH electrode is 5 to 6 mm below the surface.
- If necessary add extra deionized water.

Note: The dispensing tip should be slightly submerged in the sample.

- Press . The titrator start the analysis.
- At the end of the titration, after detection of the equivalence point, "Titration Completed" will appear with the result. The result is expressed in **N (eq/L) of hydrochloric acid**.
- Remove the pH electrode, temperature probe and stirrer from the sample and rinse them thoroughly with deionized water.
- Record the result.

Note: For improved accuracy, repeat this procedure a minimum of three times and calculate the average value.

For methods utilizing 0.1N hydrochloric acid titrant solution, follow the steps below to enter the titer/standardized value.

- Select the method utilizing 0.1 N hydrochloric acid.
- Press  from the main screen.
- Using the arrow keys, highlight Titrant Conc. and press .
- Use the numeric keypad to enter the standardized (titer) value of the titrant then press .
- Press  to exit the **View/Modify Method** screen.
- Use the arrow keys to highlight *Save Method* and press .

METHOD PARAMETERS

Name: 0.1N Hydrochloric Acid
 Method Revision: 4.0
 Analysis Type: Standard Titration
 Measurement Input: Analog CH1
 Stirrer Configuration:
 Stirrer: Stirrer 1
 Stirring Speed: 1400 RPM
 Titrant Pump: Pump 1
 Reagent Addition 1: Disabled
 Reagent Addition 2: Disabled
 Dosing Type: Dynamic
 Min Vol: 0.030 mL
 Max Vol: 0.500 mL
 delta E: 6.000 mV
 End Point Mode: pH 1EQ point, 1st Der
 Recognition Options:
 Threshold: 500 mV/mL
 Range: NO
 Filtered Derivatives: NO
 Pre-Titration Volume: 5.000 mL
 Pre-Titration Stir Time: 0 sec
 Measurement Mode: Signal Stability
 delta E: 1.0 mV
 delta t: 2 sec
 Min wait: 3 sec
 Max wait: 15 sec
 Electrode Type: pH
 Blank Option: No Blank
 Calculations: Stdz. Titrant by Volume
 Dilution Option: Disabled
 Titrant Name: 0.1N HCl
 Analyte Size: 10.0000 mL
 Analyte Entry: Fixed
 Maximum Titrant Volume: 15.000 mL
 Potential Range: -2000.0 to 2000.0 mV
 Volume/Flow Rate: 25 mL/50.0 mL/min
 Signal Averaging: 1 Reading
 Significant Figures: XXXXX

CALCULATIONS

Calculations: Stdz. Titrant by Volume
 Titrant units: N (eq/L)
 Titrant volume dosed: V (L)
 Standard volume: 10.000 mL
 Standard conc.: 0.100 eq/L

$$\frac{\text{eq}}{\text{L}} \text{HCl} = \frac{10.000 * 0.100}{\text{V(L)} * 1000}$$

RESULTS

Titration Report
 Method Name: 0.1N Hydrochloric Acid
 Time & Date: 14:55 May 30, 2026
 Report ID: Ti_00002

Titration Results
 Method Name: 0.1N Hydrochloric Acid
 Time & Date: 14:55 May 30, 2026
 Analyte Size: 10.000 mL
 End Point Volume: 9.979 mL
 pH Equivalence Point: 5.059
 Result: 0.10020 N(eq/L)
 Initial & Final pH: 12.135 to 4.989
 Titration Duration: 2:45 [mm:ss]
 Titration went to Completion:

Analyst Signature: _____

HI0003EN – 0.1M SODIUM THIOSULFATE TITRANT CONCENTRATION

Description

Method for the standardization (titer determination) of 0.1M Sodium Thiosulfate ($\text{Na}_2\text{S}_2\text{O}_3$) titrant solution against Potassium Iodate (KIO_3). The results are expressed in **M (mol/L)**.

Reference

Standard Methods for the Examination of Water and Wastewater 19th Edition, Method 4500-Cl B

Electrode

- HI3131B Combination ORP Electrode

Reagents

- HI70439 0.1M Sodium Thiosulfate (1 L)
- HI70407 Potassium Iodate (20 g)
- HI70425 16% Sulfuric Acid (500 mL)
- HI70468 Potassium Iodide (35 g)
- HI70436 Deionized Water (1 gal)

Accessories

- HI70300L Storage Solution (500 mL)
- HI7071 Electrode Fill Solution (30 mL × 4)
- HI920-053 100-mL Plastic Beaker (20 pcs.)
- Analytical Balance 0.0001 g
- 100 mL Class A Volumetric Flask
- 10 mL Class A Volumetric Pipette

Device Preparation

- Connect the ORP electrode to the titrator.
- Install a 25 mL burette filled with 0.1M sodium thiosulfate (HI70439) on pump one and verify that no air bubbles are present in the burette or tubing. If necessary prime the burette until all the air has been removed completely.
- Press Select from the main screen.
- Use the arrow keys to highlight HI0003EN 0.1M Sodium Thiosulfate and press Select.

Electrode Preparation

- Prepare the ORP electrode according to the procedure in the manual.

Sample Preparation



- Crush approximately 2 grams of potassium iodate (HI70407) and dry it for 2 hours at 120°C. Cool to room temperature in a desiccator.
- Carefully weigh approximately 0.35 grams of dried potassium iodate.
- Record the exact weight of the sample once the balance has stabilized with an accuracy of 0.0001 grams.
- Carefully transfer the salt to a 100 mL Class A volumetric flask. Add approximately 80 mL of deionized water, and mix to dissolve. Once the salt is completely dissolved bring the flask to volume with deionized water. Mix well.
- Use a Class A volumetric pipette to transfer exactly 10.00 mL of the solution to a clean 100 mL plastic beaker.
- Add deionized water to the 50 mL mark on the beaker.
- Add 5.00 mL of 16% sulfuric acid (HI70425) and 1.5 grams of potassium iodide (HI70468) to the beaker.

Analysis

- Place the beaker under the stirrer assembly and lower it to immerse the ORP electrode and stirrer.
- Ensure that the reference junction of the ORP electrode is 5 to 6 mm below the surface.






- If necessary add extra deionized water.

Note: The dispensing tip should be slightly submerged in the sample.

- Press . The user is prompted to enter the weight of the analyte (weight of potassium iodate). Use the numeric keypad to enter the exact weight and press  to start the analysis.
- At the end of the titration, after detection of the equivalence point, "Titration Completed" will appear with the result. The result is expressed in **M (mol/L) of sodium thiosulfate**.
- Remove the ORP electrode and stirrer from the sample and rinse them thoroughly with deionized water.
- Record the result.

Note: For improved accuracy, repeat this procedure a minimum of three times and calculate the average value.

For methods utilizing 0.1M sodium thiosulfate titrant solution, follow the steps below to enter the titer/standardized value.

- Select the method utilizing 0.1M sodium thiosulfate.
- Press  from the main screen.
- Using the arrow keys, highlight Titrant Conc. and press .
- Use the numeric keypad to enter the standardized (titer) value of the titrant then press .
- Press  to exit the **View/Modify Method** screen.
- Use the arrow keys to highlight *Save Method* and press .

METHOD PARAMETERS

Name: 0.1M Sodium Thiosulfate
 Method Revision: 4.0
 Analysis Type: Standard Titration
 Measurement Input: Analog CH1
 Stirrer Configuration:
 Stirrer: Stirrer 1
 Stirring Speed: 1400 RPM
 Titrant Pump: Pump 1
 Reagent Addition 1: Disabled
 Reagent Addition 2: Disabled
 Dosing Type: Dynamic
 Min Vol: 0.030 mL
 Max Vol: 0.600 mL
 delta E: 6.500 mV
 End Point Mode: mV 1EQ point, 1st Der
 Recognition Options:
 Threshold: 50 mV/mL
 Range: NO
 Filtered Derivatives: NO
 Pre-Titration Volume: 5.000 mL
 Pre-Titration Stir Time: 0 sec
 Measurement Mode: Signal Stability
 delta E: 0.3 mV
 delta t: 2 sec
 Min wait: 2 sec
 Max wait: 20 sec
 Electrode Type: ORP
 Blank Option: No Blank
 Calculations: Stdz. Titrant by Weight
 Dilution Option: Enabled
 Final Dilution Volume: 100.000 mL
 Aliquot Volume: 10.000 mL
 Titrant Name: 0.1M Na₂S₂O₃
 Analyte Size: 0.35000 g
 Analyte Entry: Manual
 Maximum Titrant Volume: 15.000 mL
 Potential Range: -2000.0 to 2000.0 mV

Volume/Flow Rate: 25 mL/50.0 mL/min
 Signal Averaging: 1 Reading
 Significant Figures: XXXXX

CALCULATIONS

Calculations: Stdz. Titrant by Weight
 Titrant units: M (mol/L)
 Titrant volume dosed: V (L)
 Standard weight: 0.350 g
 Dilution Factor: 0.100
 Final Dilution volume: 100.000 mL
 Aliquot Volume: 10.000 mL
 mw of standard: 214.00 g/mol
 Titrant/Standard: 6.000 mol/mol

$$\frac{\text{mol}}{\text{L}} \text{Na}_2\text{S}_2\text{O}_3 = \frac{0.350 \cdot 0.10 \cdot 6.0}{214.00 \cdot V(\text{L})}$$

RESULTS

Titration Report
 Method Name: 0.1M Sodium Thiosulfate
 Time & Date: 17:03 May 07, 2026
 Report ID: Ti_00073

Titration Results
 Method Name: 0.1M Sodium Thiosulfate
 Time & Date: 17:03 May 07, 2026
 Analyte Size: 0.35020 g
 End Point Volume: 9.635 mL
 mV Equivalence Point: 233.0
 Result: 0.10191 M (mol/L)
 Initial & Final mV: 361.8 to 173.4
 Titration Duration: 2:51 [mm:ss]
 Titration went to Completion:

Analyst Signature: _____

HI0010EN — 0.1M FERROUS AMMONIUM SULFATE TITRANT CONCENTRATION

Description

Method for the standardization (titer determination) of 0.1M Ferrous Ammonium Sulfate (FAS) titrant solution against Potassium Dichromate ($K_2Cr_2O_7$). The results are expressed in **M (mol/L)**.

Reference

Standard Methods for the Examination of Water and Wastewater 21st Edition, Method 5220B

Electrode

- [HI3131B](#) Combination ORP Electrode

Reagents

- [HI70444](#) 25% Sulfuric Acid
- [HI70436](#) Deionized Water (1 gal)
- Ferrous Ammonium Sulfate (ACS Grade)
- Potassium Dichromate (ACS Grade)



Accessories

- [HI70300L](#) Storage Solution (500 mL)
- [HI7071](#) Electrode Fill Solution (30 mL × 4)
- [HI7092](#) Pretreatment Oxidizing Solution (230 mL)
- [HI920-053](#) 100-mL Plastic Beaker (20 pcs.)
- Analytical Balance with 0.0001 g resolution
- 100 mL Class A Volumetric Flask
- 500 mL Class A Volumetric Flask
- 10 mL Class A Volumetric Pipette

Titration Preparation

- Carefully weigh 19.607 grams of ferrous ammonium sulfate.
- Carefully transfer the salt to a 500 mL Class A volumetric flask.
- Add approximately 300 mL of deionized water. Mix to dissolve.
- Add 40.00 mL of 25% sulfuric acid ([HI70444](#)) to the flask. Invert the solution to mix.
- Allow the flask to return to room temperature.
- Bring the flask to volume with deionized water. Mix well.

Device Preparation

- Connect the ORP electrode to the titrator.
- Install a 25 mL burette filled with 0.1M ferrous ammonium sulfate on pump one and verify that no air bubbles are present in the burette or tubing. If necessary prime the burette until all the air has been removed completely.
- Press  from the main screen. Use the arrow keys to highlight [HI0010EN 0.1M FAS TITR. CONC](#) and press .

Electrode Preparation

- Prepare the ORP electrode by soaking in [HI7092](#) Pretreatment oxidizing solution for 30 minutes.

Sample Preparation

- Carefully weigh approximately 0.49 grams of dried potassium dichromate.
- Record the exact weight of the sample once the balance has stabilized with an accuracy of 0.0001 grams.
- Carefully transfer the salt to a 100 mL Class A volumetric flask. Add approximately 80 mL of deionized water, and mix to dissolve. Once the salt is completely dissolved bring the flask to volume with deionized water, mix well.
- Use a Class A volumetric pipette to transfer exactly 10.00 mL of the solution to a clean 100 mL plastic beaker.
- Add 25.00 mL of 25% sulfuric acid ([HI70444](#)) to the beaker.
- Add deionized water to the 50 mL mark on the beaker.

Analysis

- Place the beaker under the stirrer assembly and lower it to immerse the electrode and stirrer.
Ensure that the reference junction of the ORP electrode is 5 to 6 mm below the surface. If necessary add extra deionized water.

Note: The dispensing tip should be slightly submerged in the sample.

- Press start stop. The user is prompted to enter the weight of the analyte (weight of potassium dichromate).
Use the numeric keypad to enter the exact weight. Press enter to start the analysis.
- At the end of the titration, after detection of the equivalence point, "Titration Completed" will appear with the result.
The result is expressed in **M (mol/L) of ferrous ammonium sulfate**.
- Remove the ORP electrode and stirrer from the sample and rinse them thoroughly with deionized water.
- Record the result.

Note: For improved accuracy, repeat this procedure a minimum of three times and calculate the average value.

For methods utilizing 0.1M ferrous ammonium sulfate titrant solution, follow steps below to enter titer/standardized value.

- Select the method utilizing 0.1M ferrous ammonium sulfate.
- Press Method Options from the main screen.
- Using the arrow keys to highlight Titrant Conc. Press Select.
- Use the numeric keypad to enter the standardized (titer) value of the titrant then press Accept.
- Press Escape to exit the **View/Modify Method** screen. Use the arrow keys to highlight **Save Method** and press Select.

METHOD PARAMETERS

```
Name: 0.1M FAS
Method Revision: 4.0
Analysis Type: Standard Titration
Measurement Input: Analog CH1
Stirrer Configuration:
  Stirrer: Stirrer 1
  Stirring Speed: 1400 RPM
Titrant Pump: Pump 1
Reagent Addition 1: Disabled
Reagent Addition 2: Disabled
Dosing Type: Dynamic
  Min Vol: 0.030 mL
  Max Vol: 0.150 mL
  delta E: 4.000 mV
End Point Mode: mV 1EQ point, 1st Der
Recognition Options:
  Threshold: 500 mV/mL
  Range: NO
  Filtered Derivatives: YES
Pre-Titration Volume: 7.000 mL
Pre-Titration Stir Time: 20 sec
Measurement Mode: Signal Stability
delta E: 4.0 mV
delta t: 2 sec
Min wait: 3 sec
Max wait: 20 sec
Electrode Type: ORP
Blank Option: No Blank
Calculations: Stdz. Titrant by Weight
Dilution Option: Enabled
  Final Dilution Volume: 100.000 mL
  Aliquot Volume: 10.000 mL
Titrant Name: 0.1M FAS
Analyte Size: 0.49000 g
Analyte Entry: Manual
```

```
Maximum Titrant Volume: 15.000 mL
Potential Range: -2000.0 to 2000.0 mV
Volume/Flow Rate: 25 mL/50.0 mL/min
Signal Averaging: 1 Reading
Significant Figures: XXXXX
```

CALCULATIONS

```
Calculations: Stdz. Titrant by Weight
Titrant units: M (mol/L)
Titrant volume dosed: V (L)
Standard weight: 0.490 g
Dilution Factor: 0.100
  Final Dilution volume: 100.000 mL
  Aliquot Volume: 10.000 mL
mw of standard: 294.18 g/mol
Titrant/Standard: 6.000 mol/mol

$$\frac{\text{mol}}{\text{L}} \text{ FAS} = \frac{0.490 * 0.10 * 6.0}{294.18 * V(\text{L})}$$

```

RESULTS

```
Titration Report
Method Name: 0.1M FAS
Time & Date: 15:59 May 1, 2026
Report ID: Ti_00015

Titration Results
Method Name: 0.1M FAS
Time & Date: 15:59 May 1, 2026
Analyte Size: 0.491 g
End Point Volume: 9.879 mL
mV Equivalence Point: 667.4
Result: 0.10137 M (mol/L)
Initial & Final mV: 791.3 to 598.0
Titration Duration: 3:05 [mm:ss]
Titration went to Completion:

Analyst Signature: _____
```

HI0200EN — 0.02M SILVER NITRATE TITRANT CONCENTRATION

Description

Method for the standardization (titer determination) of 0.02M Silver Nitrate (AgNO_3) titrant solution against Sodium Chloride (NaCl). The results are expressed in **M (mol/L)**.

Reference

AOAC Official Methods of Analysis, Official Method 941.18

Electrode

- [HI4115](#) Silver/Sulfide Combination ISE

Reagents

- [HI70448](#) 0.02M Silver Nitrate (1 L)
- [HI70406](#) Sodium Chloride (20 g)
- [HI70427](#) 1.5M Nitric Acid Solution (500 mL)
- [HI70436](#) Deionized Water (1 gal)

Accessories

- [HI7072](#) Electrode Fill Solution (4 × 30 mL)
- Analytical Balance with 0.0001 g resolution
- 150 mL Glass Beaker
- 100 mL Class A Volumetric Flask
- 5 mL Class A Volumetric Pipette

Device Preparation

- Connect the Silver/Sulfide electrode to the titrator.
- Install a 25 mL burette filled with 0.02M silver nitrate ([HI70448](#)) on pump one and verify that no air bubbles are present in the burette or tubing. If necessary prime the burette until all the air has been removed completely.
- Press Select Method from the main screen. Use the arrow keys to highlight [HI0200EN 0.02M Silver Nitrate](#) and press Select.

Electrode Preparation

- Prepare the Silver/Sulfide electrode according to the procedure in the manual.

Sample Preparation

- Crush approximately 2 grams of sodium chloride ([HI70406](#)) and dry it for 2 hours at 140°C. Cool to room temperature in a desiccator.
- Weigh 0.20 g of dried sodium chloride with an accuracy of 0.0001 g. Transfer the salt to a 100 mL volumetric flask. Add approximately 80 mL of distilled water and mix. Dissolve completely before bringing to volume.
- Use a Class A volumetric pipette to transfer exactly 5.00 mL of prepared standard solution to a 150 mL glass beaker and add distilled water to the 100 mL mark on the beaker.
- Add 10.00 mL of 1.5M nitric acid ([HI70427](#)) to the beaker.

Analysis

- Place the beaker under the stirrer assembly and lower it to immerse the Silver/Sulfide electrode and stirrer. Ensure that the reference junction of the electrode is 5 to 6 mm below the surface. If necessary add extra deionized water.

Note: The dispensing tip should be slightly submerged in the sample.

- Press start stop. You will be prompted to enter the weight of the analyte (weight of sodium chloride). Use the numeric keypad to enter the exact weight and press enter to start the analysis.

- At the end of the titration, after detection of the equivalence point, "Titration Completed" will appear with the result. The result is expressed in **M (mol/L) of silver nitrate**.
- Remove the electrode and stirrer from the sample and rinse them thoroughly with deionized water.
- Record the result.

Note: For improved accuracy, repeat this procedure a minimum of three times and calculate the average value.

For methods utilizing 0.02M silver nitrate titrant solution, follow the steps below to enter the titer/standardized value.

- Select the method utilizing 0.02M silver nitrate.
- Press Method Options from the main screen.
- Using the arrow keys, highlight Titrant Conc. and press Select.
- Use the numeric keypad to enter the standardized (titer) value of the titrant then press Accept.
- Press Escape to exit the **View/Modify Method** screen. Use the arrow keys to highlight **Save Method** and press Select.

METHOD PARAMETERS

```
Name: 0.02M Silver Nitrate
Method Revision: 4.0
Analysis Type: Standard Titration
Measurement Input: Analog CH1
Stirrer Configuration:
  Stirrer: Stirrer 1
  Stirring Speed: 1400 RPM
Titrant Pump: Pump 1
Reagent Addition 1: Disabled
Reagent Addition 2: Disabled
Dosing Type: Dynamic
  Min Vol: 0.030 mL
  Max Vol: 0.500 mL
  delta E: 8.000 mV
End Point Mode: mV 1EQ point, 1st Der
Recognition Options:
  Threshold: 100 mV/mL
  Range: NO
  Filtered Derivatives: YES
Pre-Titration Volume: 6.000 mL
Pre-Titration Stir Time: 0 sec
Measurement Mode: Signal Stability
  delta E: 1.0 mV
  delta t: 2 sec
  Min wait: 2 sec
  Max wait: 20 sec
Electrode Type: Silver/Sulfide
Blank Option: No Blank
Calculations: Stdz. Titrant by Weight
Dilution Option: Enabled
  Final Dilution Volume: 100.000 mL
  Aliquot Volume: 5.000 mL
Titrant Name: 0.02M AgNO3
Analyte Size: 0.20000 g
Analyte Entry: Manual
Maximum Titrant Volume: 15.000 mL
Potential Range: -2000.0 to 2000.0 mV
Volume/Flow Rate: 25 mL/50.0 mL/min
Signal Averaging: 1 Reading
Significant Figures: XXXXX
```

CALCULATIONS

```
Calculations: Stdz. Titrant by Weight
Titrant units: M (mol/L)
Titrant volume dosed: V (L)
Standard weight: 0.200 g
Dilution Factor: 0.05
  Final Dilution volume: 100.000 mL
  Aliquot Volume: 5.000 mL
mw of standard: 58.440 g/mol
Titrant/Standard: 1.000 mol/mol

$$\frac{\text{mol}}{\text{L}} \text{AgNO}_3 = \frac{0.200 * 0.05 * 1.0}{58.440 * V(\text{L})}$$

```

RESULTS

```
Titration Report
Method Name: 0.02M Silver Nitrate
Time & Date: 15:52 May 1, 2026
Report ID: Ti_00037

Titration Results
Method Name: 0.02M Silver Nitrate
Time & Date: 15:52 May 1, 2026
Analyte Size: 0.1923 g
End Point Volume: 9.065 mL
mV Equivalence Point: 273.1
Result: 0.01815 M (mol/L)
Initial & Final mV: 146.9 to 291.0
Titration Duration: 2:21 [mm:ss]
Titration went to Completion:

Analyst Signature: _____
```

HI1004EN – ALKALINITY OF WATER

0 to 2500 mg/L CaCO₃, pH 4.5 Endpoint

Description

Method for the determination of total (methyl red) alkalinity in water by titration of a sample to pH 4.5. The results are expressed in mg/L (ppm) as calcium carbonate.

For the determination of phenolphthalein alkalinity, set the endpoint to pH 8.3.

Reference

Standard Methods for the Examination of Water and Wastewater 21st edition, Method 2320B

Electrode

- HI1131B Combination pH Electrode
- HI7662-T Temperature Probe

Reagents

- HI70463 0.1N Hydrochloric Acid (1 L)
- HI70436 Deionized Water (1 gal)

Accessories

- HI70300L Storage Solution (500 mL)
- HI7082 Electrode Fill Solution (4 × 30 mL)
- HI7004L pH 4.01 Buffer Solution (500 mL)
- HI7007L pH 7.01 Buffer Solution (500 mL)
- HI7010L pH 10.01 Buffer Solution (500 mL)
- HI920-053 100-mL Plastic Beaker (20 pcs.)
- 50 mL Class A Volumetric Pipette

Device Preparation

- Connect the pH electrode and temperature probe to the titrator.
- Install a 25 mL burette filled with 0.1N hydrochloric acid (HI70463) on pump one and verify that no air bubbles are present in the burette or tubing. If necessary prime the burette until all the air has been removed completely.
- For the determination of the exact concentration of the 0.1N hydrochloric acid, follow [HI0002EN 0.1N Hydrochloric Acid Titrant Concentration](#).
- Press from the main screen. Use the arrow keys to highlight [HI1004EN Alkalinity of Water](#) and press .

Electrode Preparation

- Press from the main screen, if necessary select the analog board and press .
- Calibrate the electrode using pH 4.01, 7.01 and 10.01 buffers. Refer to the instruction manual for calibration procedure.


Sample Preparation

- Use a Class A volumetric pipette to transfer exactly 50.00 mL of sample to a clean 100 mL plastic beaker.

Analysis

- Place the beaker under the stirrer assembly and lower it to immerse the pH electrode, temperature sensor and stirrer. Ensure that the reference junction of the pH electrode is 5 to 6 mm below the surface. If necessary add extra deionized water.

Note: The dispensing tip should be slightly submerged in the sample.

- Press . The titrator will start the analysis.
- At the end of the titration, when pH 4.50 is reached, "Titration Completed" will appear with the result. The result is expressed in **mg/L as calcium carbonate**.
- Remove the electrode and stirrer from the sample and rinse them thoroughly with deionized water.
- Record the result.

METHOD PARAMETERS

Name: Alkalinity of Water
 Method Revision: 4.0
 Analysis Type: Standard Titration
 Measurement Input: Analog CH1
 Stirrer Configuration:
 Stirrer: Stirrer 1
 Stirring Speed: 1400 RPM
 Titrant Pump: Pump 1
 Reagent Addition 1: Disabled
 Reagent Addition 2: Disabled
 Dosing Type: Dynamic
 Min Vol: 0.050 mL
 Max Vol: 0.500 mL
 delta E: 5.000 mV
 End Point Mode: Fixed 4.500 pH
 Pre-Titration Volume: 0.000 mL
 Pre-Titration Stir Time: 0 sec
 Measurement Mode: Signal Stability
 delta E: 1.0 mV
 delta t: 2 sec
 Min wait: 2 sec
 Max wait: 20 sec
 Electrode Type: pH
 Blank Option: No Blank
 Calculations: Sample Calc. by Volume
 Dilution Option: Disabled
 Titrant Name: 0.1N HCl
 Titrant Conc.: 0.1000 N(eq/L)
 Analyte Size: 50.000 mL
 Analyte Entry: Fixed
 Maximum Titrant Volume: 25.000 mL
 Potential Range: -2000.0 to 2000.0 mV
 Volume/Flow Rate: 25 mL/50.0 mL/min
 Signal Averaging: 1 Reading
 Significant Figures: XXXXX

CALCULATIONS

Calculations: Sample Calc. by Volume
 Titrant units: N (eq/L)
 Titrant volume dosed: V (L)
 Final result units: mg/L
 Titrant Conc.: 0.1000 N(eq/L)
 Sample/Titrant: 0.500 mol/eq
 mw of standard: 100.09 g/mol
 Sample Volume: 50.000 mL

$$\frac{\text{mg}}{\text{L}} \text{CaCO}_3 = \frac{V(\text{L}) * 1000 * 0.10 * 0.5 * 100.09 * 1000}{50.00}$$

RESULTS

Titration Report

Method Name: Alkalinity of Water
 Time & Date: 14:36 May 1, 2026
 Report ID: Ti_00036

Titration Results

Method Name: Alkalinity of Water
 Time & Date: 14:36 May 1, 2026
 Analyte Size: 50.000 mL
 End Point Volume: 9.336 mL
 pH Fixed End Point: 4.500
 Result: 934.44 mg/L
 Initial & Final pH: 10.232 to 4.419
 Titration Duration: 3:23 [mm:ss]
 Titration went to Completion:

Analyst Signature: _____

HI1005EN – ACIDITY OF WATER

0 to 2500 mg/L, pH 8.3 Endpoint

Description

Method for the determination of total (phenolphthalein) acidity in water by titration of a sample to pH 8.3. The results are expressed in **mg/L (ppm) as calcium carbonate**.

For the determination of methyl orange acidity, set the endpoint to pH 3.7.

Reference

Standard Methods for the Examination of Water and Wastewater 21st edition, Method 2310B

Electrode

- [HI1131B](#) Combination pH Electrode
- [HI7662-T](#) Temperature Probe

Reagents

- [HI70456](#) 0.1N Sodium Hydroxide (1 L)
- [HI70436](#) Deionized Water (1 gal)

Accessories

- [HI70300L](#) Storage Solution (500 mL)
- [HI7082](#) Electrode Fill Solution (4 × 30 mL)
- [HI7004L](#) pH 4.01 Buffer Solution (500 mL)
- [HI7007L](#) pH 7.01 Buffer Solution (500 mL)
- [HI7010L](#) pH 10.01 Buffer Solution (500 mL)
- [HI920-053](#) 100-mL Plastic Beaker (20 pcs.)
- 50 mL Class A Volumetric Pipette

Device Preparation

- Connect the pH electrode and temperature probe to the titrator.
- Install a 25 mL burette filled with 0.1N sodium hydroxide ([HI70456](#)) on pump one and verify that no air bubbles are present in the burette or tubing. If necessary prime the burette until all the air has been removed completely.
- For the determination of the exact concentration of the 0.1N sodium hydroxide, follow [HI0001EN 0.1N Sodium Hydroxide Titrant Concentration](#).
- Press from the main screen. Use the arrow keys to highlight [HI1005EN Acidity in Water](#) and press .

Electrode Preparation

- Press from the main screen, if necessary select the analog board and press .
- Calibrate the electrode using pH 4.01, 7.01 and 10.01 buffers. Refer to the instruction manual for calibration procedure.


Sample Preparation

- Use a Class A volumetric pipette to transfer exactly 50.00 mL of sample to a clean 100 mL plastic beaker.

Analysis

- Place the beaker under the stirrer assembly and lower it to immerse the pH electrode, temperature sensor and stirrer. Ensure that the reference junction of the pH electrode is 5 to 6 mm below the surface. If necessary add extra deionized water.

Note: The dispensing tip should be slightly submerged in the sample.

- Press , the titrator will start the analysis.
- At the end of the titration, when pH 8.30 is reached, "Titration Completed" will appear with the result. The result is expressed in **mg/L as calcium carbonate**.
- Remove the pH electrode, temperature probe and stirrer from the sample and rinse them thoroughly with deionized water.
- Record the result.

METHOD PARAMETERS

Name: Acidity of Water
 Method Revision: 4.0
 Analysis Type: Standard Titration
 Measurement Input: Analog CH1
 Stirrer Configuration:
 Stirrer: Stirrer 1
 Stirring Speed: 1400 RPM
 Titrant Pump: Pump 1
 Reagent Addition 1: Disabled
 Reagent Addition 2: Disabled
 Dosing Type: Dynamic
 Min Vol: 0.050 mL
 Max Vol: 0.500 mL
 delta E: 5.000 mV
 End Point Mode: Fixed 8.300 pH
 Pre-Titration Volume: 0.000 mL
 Pre-Titration Stir Time: 0 sec
 Measurement Mode: Signal Stability
 delta E: 1.0 mV
 delta t: 2 sec
 Min wait: 2 sec
 Max wait: 20 sec
 Electrode Type: pH
 Blank Option: No Blank
 Calculations: Sample Calc. by Volume
 Dilution Option: Disabled
 Titrant Name: 0.1N NaOH
 Titrant Conc.: 0.1000 N(eq/L)
 Analyte Size: 50.000 mL
 Analyte Entry: Fixed
 Maximum Titrant Volume: 25.000 mL
 Potential Range: -2000.0 to 2000.0 mV
 Volume/Flow Rate: 25 mL/50.0 mL/min
 Signal Averaging: 1 Reading
 Significant Figures: XXXXX

CALCULATIONS

Calculations: Sample Calc. by Volume
 Titrant units: N (eq/L)
 Titrant volume dosed: V (L)
 Final result units: (mg/L)
 Titrant Conc.: 0.1000 N(eq/L)
 Sample/Titrant: 0.500 mol/eq
 mw of standard: 100.09 g/mol
 Sample Volume: 50.000 mL

$$\frac{\text{mg}}{\text{L}} \text{CaCO}_3 = \frac{V(\text{L}) * 1000 * 0.10 * 0.5 * 100.09 * 1000}{50.00}$$

RESULTS

Titration Report

Method Name: Acidity of Water
 Time & Date: 14:54 May 1, 2026
 Report ID: Ti_00023

Titration Results

Method Name: Acidity of Water
 Time & Date: 14:54 May 1, 2026
 Analyte Size: 50.000 mL
 End Point Volume: 5.879 mL
 pH Fixed End Point: 8.300
 Result: 588.43 (mg/L)
 Initial & Final pH: 2.465 to 8.398
 Titration Duration: 3:42 [mm:ss]
 Titration went to Completion:

Analyst Signature: _____

HI1007EN – CHLORIDE IN WATER

0 to 150 ppm (mg/L)

Description

Method for the determination of chloride in water. The results are expressed as **ppm (mg/L) as Chloride**.

Reference

Standard Methods for the Examination of Water and Wastewater 21st edition, Method 4500-Cl

Electrode

- [HI4115](#) Silver/Sulfide Combination ISE



Reagents

- [HI70448](#) 0.02M Silver Nitrate (1 L)
- [HI70427](#) 1.5M Nitric Acid Solution (500 mL)
- [HI70436](#) Deionized Water (1 gal)

Accessories

- [HI7072](#) Electrode Fill Solution (4×30 mL)
- 150 mL Glass Beaker
- 100 mL Class A Volumetric Pipette
- 10 mL Class A Volumetric Pipette

Device Preparation

- Connect the Silver/Sulfide electrode to the titrator.
- Install a 25 mL burette filled with 0.02M silver nitrate ([HI70448](#)) on pump one and verify that no air bubbles are present in the burette or tubing. If necessary prime the burette until all the air has been removed completely.
- For the determination of the exact concentration of the 0.02M Silver Nitrate, follow [HI0200EN 0.02M Silver Nitrate Titrant Concentration](#).
- Press  from the main screen. Use the arrow keys to highlight [HI1007EN Chloride in Water](#) and press .

Electrode Preparation

- Prepare the Silver/Sulfide electrode according to the procedure in the manual.


Sample Preparation

- Use a class A volumetric pipette to transfer exactly 100.00 mL of sample to a clean 150 mL beaker.
- Add 10.00 mL of 1.5M nitric acid ([HI70427](#)) to the beaker.

Analysis

- Place the beaker under the stirrer assembly and lower it to immerse the electrode and stirrer. Ensure that the reference junction of the electrode is 5 to 6 mm below the surface. If necessary add extra deionized water.

Note: *The dispensing tip should be slightly submerged in the sample.*

- Press , the titrator will start the analysis.
- At the end of the titration, after detection of the equivalence point, “Titration Completed” will appear with the result. The result is expressed in **ppm (mg/L) of chloride**.
- Remove the electrode and stirrer from the sample and rinse them thoroughly with deionized water.
- Record the result.

METHOD PARAMETERS

Name: Chloride in Water
 Method Revision: 4.0
 Analysis Type: Standard Titration
 Measurement Input: Analog CH1
 Stirrer Configuration:
 Stirrer: Stirrer 1
 Stirring Speed: 1400 RPM
 Titrant Pump: Pump 1
 Reagent Addition 1: Disabled
 Reagent Addition 2: Disabled
 Dosing Type: Dynamic
 Min Vol: 0.030 mL
 Max Vol: 0.500 mL
 delta E: 5.000 mV
 End Point Mode: mv 1EQ point, 1st Der
 Recognition Options:
 Threshold: 100 mV/mL
 Range: NO
 Filtered Derivatives: NO
 Pre-Titration Volume: 0.000 mL
 Pre-Titration Stir Time: 0 sec
 Measurement Mode: Signal Stability
 delta E: 1.0 mV
 delta t: 2 sec
 Min wait: 2 sec
 Max wait: 20 sec
 Electrode Type: Silver/Sulfide
 Blank Option: No Blank
 Calculations: Sample Calc. by Volume
 Dilution Option: Disabled
 Titrant Name: 0.02M AgNO3
 Titrant Conc.: 2.0000E-2 M (mol/L)
 Analyte Size: 100.000 mL
 Analyte Entry: Manual
 Maximum Titrant Volume: 25.000 mL
 Potential Range: -2000.0 to 2000.0 mV
 Volume/Flow Rate: 25 mL/50.0 mL/min
 Signal Averaging: 1 Reading
 Significant Figures: XXXXX

CALCULATIONS

Calculations: Sample Calc. by Volume
 Titrant units: M (mol/L)
 Titrant volume dosed: V (L)
 Final result units: (mg/L)
 Titrant Conc.: 2.0000E-2 M (mol/L)
 Sample/Titrant: 1.000 mol/mol
 mw of sample: 35.453 g/mol
 Sample Volume: 100.000 mL

$$\frac{\text{mg}}{\text{L}} = \frac{\text{V(L)} * 1000 * 0.02 * 1.0 * 35.45 * 1000}{100.0}$$

RESULTS

Titration Report

Method Name: Chloride in Water
 Time & Date: 15:11 May 1, 2026
 Report ID: Ti_00052

Titration Results

Method Name: Chloride in Water
 Time & Date: 15:11 May 1, 2026
 Analyte Size: 100.000 mL
 End Point Volume: 4.781 mL
 mV Fixed End Point: 280.3
 Result: 33.897 ppm (mg/L)
 Initial & Final mV: 94.8 to 298.5
 Titration Duration: 1:24 [mm:ss]
 Titration went to Completion:

Analyst Signature: _____

HI1008EN – NEUTRALIZATION WITH SULFURIC ACID

0 to 200 meq/L

Description

Method for the determination of strong or weak base concentration by titration of a sample to the equivalence point with sulfuric acid. The results are expressed as **meq/L**.

Electrode

- [HI1131B](#) Combination pH Electrode
- [HI7662-T](#) Temperature Probe

Reagents

- [HI70459](#) 0.05M Sulfuric Acid (1 L)
- [HI70436](#) Deionized Water (1 gal)

Accessories

- [HI70300L](#) Storage Solution (500 mL)
- [HI7082](#) Electrode Fill Solution (4×30 mL)
- [HI7004L](#) pH 4.01 Buffer Solution
- [HI7007L](#) pH 7.01 Buffer Solution
- [HI7010L](#) pH 10.01 Buffer Solution
- [HI920-053](#) 100-mL Plastic Beaker (20 pcs.)
- 10 mL Class A Volumetric Pipette

Device Preparation

- Connect the pH electrode and temperature probe to the titrator.
- Install a 25 mL burette filled with 0.05M sulfuric acid ([HI70459](#)) on pump one and verify that no air bubbles are present in the burette or tubing. If necessary prime the burette until all the air has been removed completely.
- For the determination of the exact concentration of the 0.05M sulfuric acid, follow [HI0103EN 0.05M Sulfuric Acid Titrant Concentration](#).
- Press from the main screen. Use the arrow keys to highlight [HI1008EN Neutralization w/H2SO4](#) and press .

Electrode Preparation

- Press from the main screen, if necessary select the analog board and press .
- Calibrate the electrode using pH 4.01, 7.01 and 10.01 buffers. Refer to the instruction manual for calibration procedure.

Sample Preparation

- Use a class A volumetric pipette to transfer exactly 10.00 mL of sample to a clean 100 mL plastic beaker.
- Add deionized water to the 50 mL mark on the beaker.

Analysis

- Place the beaker under the stirrer assembly and lower it to immerse the pH electrode, temperature probe and stirrer. Ensure that the reference junction of the pH electrode is 5 to 6 mm below the surface.

Note: The dispensing tip should be slightly submerged in the sample.

- Press , the titrator will start the analysis.
- At the end of the titration, after detection of the equivalence point, "Titration Completed" will appear with the result. The result is expressed in **meq/L**.
- Remove the pH electrode and stirrer from the sample and rinse them thoroughly with deionized water.
- Record the result.

METHOD PARAMETERS

Name: Neutralization w/ H2SO4
 Method Revision: 4.0
 Analysis Type: Standard Titration
 Measurement Input: Analog CH1
 Stirrer Configuration:
 Stirrer: Stirrer 1
 Stirring Speed: 1400 RPM
 Titrant Pump: Pump 1
 Reagent Addition 1: Disabled
 Reagent Addition 2: Disabled
 Dosing Type: Dynamic
 Min Vol: 0.050 mL
 Max Vol: 0.500 mL
 delta E: 20.000 mV
 End Point Mode: pH 1EQ point, 1st Der
 Recognition Options:
 Threshold: 50 mV/mL
 Range: NO
 Filtered Derivatives: NO
 Pre-Titration Volume: 0.000 mL
 Pre-Titration Stir Time: 0 sec
 Measurement Mode: Signal Stability
 delta E: 1.0 mV
 delta t: 2 sec
 Min wait: 2 sec
 Max wait: 15 sec
 Electrode Type: pH
 Blank Option: No Blank
 Calculations: Sample Calc. by Volume
 Dilution Option: Disabled
 Titrant Name: 0.05M H2SO4
 Titrant Conc.: 5.0000E-2 M (mol/L)
 Analyte Size: 10.000 mL
 Analyte Entry: Fixed
 Maximum Titrant Volume: 20.000 mL
 Potential Range: -2000.0 to 2000.0 mV
 Volume/Flow Rate: 25 mL/50.0 mL/min
 Signal Averaging: 1 Reading
 Significant Figures: XXXXX

CALCULATIONS

Calculations: Sample Calc. by Volume
 Titrant units: M (mol/L)
 Titrant volume dosed: V (L)
 Final result units: meq/L
 Titrant Conc.: 5.0000E-2 M (mol/L)
 Sample/Titrant: 2.000 eq/mol
 Sample Volume: 10.000 mL

$$\frac{\text{meq}}{\text{L}} = \frac{V(\text{L}) * 1000 * 0.05 * 2.0 * 1000}{10.0}$$

RESULTS

Titration Report

Method Name: Neutralization w/ H2SO4
 Time & Date: 09:46 May 1, 2026
 Report ID: Ti_00027

Titration Results

Method Name: Neutralization w/ H2SO4
 Time & Date: 09:46 May 1, 2026
 Analyte Size: 10.000 mL
 End Point Volume: 9.562 mL
 mV Equivalence Point: 7.966
 Result: 95.620 meq/L
 Initial & Final pH: 11.655 to 6.248
 Titration Duration: 1:24 [mm:ss]
 Titration went to Completion:

Analyst Signature: _____

HI1009EN – NEUTRALIZATION WITH SODIUM HYDROXIDE

0 to 200 meq/L

Description

Method for the determination of strong or weak acid concentration by titration of a sample to the equivalence point with sodium hydroxide. The results are expressed as **meq/L**.

Electrode

- [HI1131B](#) Combination pH Electrode
- [HI7662-T](#) Temperature Probe

Reagents

- [HI70456](#) 0.1N Sodium Hydroxide (1 L)
- [HI70436](#) Deionized Water (1 gal)

Accessories

- [HI70300L](#) Storage Solution (500 mL)
- [HI7082](#) Electrode Fill Solution (4×30 mL)
- [HI7004L](#) pH 4.01 Buffer Solution
- [HI7007L](#) pH 7.01 Buffer Solution
- [HI7010L](#) pH 10.01 Buffer Solution
- [HI920-053](#) 100-mL Plastic Beaker (20 pcs.)
- 10 mL Class A Volumetric Pipette

Device Preparation

- Connect the pH electrode and temperature probe to the titrator.
- Install a 25 mL burette filled with 0.1N sodium hydroxide ([HI70456](#)) on pump one and verify that no air bubbles are present in the burette or tubing. If necessary prime the burette until all the air has been removed completely.
- For the determination of the exact concentration of the 0.1N sodium hydroxide, follow [HI0001EN 0.1N Sodium Hydroxide Titrant Concentration](#)
- Press from the main screen. Use the arrow keys to highlight [HI1009EN Neutralization w/NaOH](#) and press .

Electrode Preparation

- Press from the main screen, if necessary select the analog board and press .
- Calibrate the electrode using pH 4.01, 7.01 and 10.01 buffers. Refer to the instruction manual for calibration procedure.

Sample Preparation

- Use a class A volumetric pipette to transfer exactly 10.00 mL of sample to a clean 100 mL plastic beaker.
- Add deionized water to the 50 mL mark on the beaker.

Analysis

- Place the beaker under the stirrer assembly and lower it to immerse the pH electrode, temperature probe and stirrer. Ensure that the reference junction of the pH electrode is 5 to 6 mm below the surface.

Note: The dispensing tip should be slightly submerged in the sample.

- Press , the titrator will start the analysis.
- At the end of the titration, after detection of the equivalence point, "Titration Completed" will appear with the result. The result is expressed in **meq/L**.
- Remove the pH electrode and stirrer from the sample and rinse them thoroughly with deionized water.
- Record the result.

METHOD PARAMETERS

Name: Neutralization w/ NaOH
 Method Revision: 4.0
 Analysis Type: Standard Titration
 Measurement Input: Analog CH1
 Stirrer Configuration:
 Stirrer: Stirrer 1
 Stirring Speed: 1400 RPM
 Titrant Pump: Pump 1
 Reagent Addition 1: Disabled
 Reagent Addition 2: Disabled
 Dosing Type: Dynamic
 Min Vol: 0.050 mL
 Max Vol: 0.500 mL
 delta E: 20.000 mV
 End Point Mode: pH 1EQ point, 1st Der
 Recognition Options:
 Threshold: 50 mV/mL
 Range: NO
 Filtered Derivatives: NO
 Pre-Titration Volume: 0.000 mL
 Pre-Titration Stir Time: 0 sec
 Measurement Mode: Signal Stability
 delta E: 1.0 mV
 delta t: 2 sec
 Min wait: 2 sec
 Max wait: 15 sec
 Electrode Type: pH
 Blank Option: No Blank
 Calculations: Sample Calc. by Volume
 Dilution Option: Disabled
 Titrant Name: 0.1N NaOH
 Titrant Conc.: 0.1000 N(eq/L)
 Analyte Size: 10.000 mL
 Analyte Entry: Fixed
 Maximum Titrant Volume: 20.000 mL
 Potential Range: -2000.0 to 2000.0 mV
 Volume/Flow Rate: 25 mL/50.0 mL/min
 Signal Averaging: 1 Reading
 Significant Figures: XXXXX

CALCULATIONS

Calculations: Sample Calc. by Volume
 Titrant units: N (eq/L)
 Titrant volume dosed: V (L)
 Final result units: meq/L
 Titrant Conc.: 5.0000E-2 M (mol/L)
 Sample/Titrant: 0.1000 N(eq/L)
 Sample Volume: 10.000 mL

$$\frac{\text{meq}}{\text{L}} = \frac{V(\text{L}) * 1000 * 0.1 * 1.0 * 1000}{10.0}$$

RESULTS

Titration Report

Method Name: Neutralization w/ NaOH
 Time & Date: 10:29 May 2, 2026
 Report ID: Ti_00017

Titration Results

Method Name: Neutralization w/ NaOH
 Time & Date: 10:29 May 2, 2026
 Analyte Size: 10.000 mL
 End Point Volume: 15.970 mL
 pH Equivalence Point: 8.431
 Result: 159.70 meq/L
 Initial & Final pH: 2.675 to 10.316
 Titration Duration: 3:20 [mm:ss]
 Titration went to Completion:

Analyst Signature: _____

HI1011EN – TROUBLESHOOTING 1

Description

Method for verifying the dosing and potentiometric signal accuracy of the titrator. This method should be used to troubleshoot a titrator equipped with a 25 mL burette. The titrator dispenses a 20.00 mL pre-titration volume, waits 20 seconds and dispenses an additional 20.00 mL dose, bringing the total volume to 40.00 mL. This procedure can also be used to check the stability of the mV and temperature channels.

The dosing accuracy of the 25 mL burette is ± 0.025 mL ($\pm 0.1\%$ of the full volume).

If the results are not correct, check all fittings for leakage, and burette and tubing for air bubbles. Repeat the measurement.



Reference

ISO/TC 48/SC1N 380E and 383E: "Piston and/or Plunger Operated Volumetric Apparatus"


Accessories

- HI762000C 0 °C Temperature Key
- HI762070C 70 °C Temperature Key
- HI70436 Deionized Water (1 gal)
- HI7662-T Temperature Probe
- Shorting Cap
- Narrow Neck Beaker
- Analytical Balance with 0.0001g resolution

Device Preparation

- Connect the shorting cap to the BNC socket on Analog Board 1.
- Install a 25 mL burette filled with room temperature deionized water (HI70436) on pump one and verify that no air bubbles are present in the burette or tubing. If necessary prime the burette until all the air has been removed completely.
- Press  from the main screen. Use the arrow keys to highlight **HI1011EN Troubleshooting 1** and press .

Large Dose Dispensing Procedure

- Add a small amount of deionized water to a narrow neck beaker.
- Place the narrow neck beaker on an analytical balance and zero the balance.
- Place the dosing tip through the neck of the beaker. Take care not to immerse it in the liquid during dispensing and not to touch the beaker walls.
- Press .
- Write down the exact weight displaced on the balance after each dose.
- This procedure can be repeated on pump 2.

Other burette sizes can be checked using the following settings, see instruction manual for accuracy:

Burette Volume	Pre-titration Volume	Max. Titrant Volume
5 mL	4.000 mL	8.000 mL
10 mL	8.000 mL	16.000 mL

METHOD PARAMETERS

Name:	Troubleshooting 1	Pre-Titration Stir Time:	0 sec
Method Revision:	4.0	Measurement Mode:	Timed Increment
Analysis Type:	Standard Titration	Time interval:	20 sec
Measurement Input:	Analog CH1	Electrode Type:	Shorting Cap
Stirrer Configuration:		Blank Option:	No Blank
Stirrer:	Stirrer 1	Calculations:	No Formula (mL only)
Stirring Speed:	0 RPM	Titrant Name:	DI Water
Titrant Pump:	Pump 1	Maximum Titrant Volume:	40.000 mL
Reagent Addition 1:	Disabled	Potential Range:	-2000.0 to 2000.0 mV
Reagent Addition 2:	Disabled	Volume/Flow Rate:	25 mL/50.0 mL/min
Dosing Type:	Linear - 20.000 mL	Signal Averaging:	1 Reading
End Point Mode:	Fixed 10.0 mV	Significant Figures:	XXXXXX
Pre-Titration Volume:	20.000 mL		

Calculations

$$V = m * \frac{1}{\rho} * \left(1 + \frac{\rho_{\text{air}}}{\rho_{\text{L}}} - \frac{\rho_{\text{air}}}{\rho_{\text{std}}} \right)$$

- V Volume of measure mass of water (mL)
 m Measure mass of water (g)
 ρ_{L} Density of dispensed water (g/mL)
 ρ_{air} Density of ambient air (g/mL)
 ρ_{std} Density of calibration standard weight (g/mL)

Alternative Calculations

If the actual values of the above parameters are not accessible the following equation can be used:

$$V = M * F$$

- V Volume of measured mass of water (mL)
 F Transformation factor

The values from the table have been calculated by correcting the air and water density with temperature, assuming the density of dry air $\rho_{\text{air}} = 0.0012 \text{ g/mL}$ and density of calibration steel standard weigh $\rho_{\text{STD}} = 8 \text{ g/mL}$.

Temperature (°C)	Factor
17.0	1.002290
18.0	1.002467
19.0	1.002654
20.0	1.002853
21.0	1.003061
22.0	1.003282
23.0	1.003512

Temperature (°C)	Factor
24.0	1.003752
25.0	1.004002
26.0	1.004261
27.0	1.004531
28.0	1.004809
29.0	1.005097
30.0	1.005395

Temperature Channel Fast Check Procedure

- Connect the shorting cap to the BNC socket on Analog Board 1.
- Connect the [HI762000C](#) 0°C temperature key to the RCA socket (temperature sensor input) on Analog Board 1.
- On the main screen select Mode, if necessary select the analog board and press mV.
- The titrator should display ATC 0.0 ±0.4°C with no fluctuations or drift.
- Connect the [HI762070C](#) 70°C temperature key to the RCA socket (temperature sensor input) on Analog Board 1.
- The titrator should display ATC 70.0 ±0.4°C with no fluctuations or drift.
- This procedure can be repeated on analog board 2.

Temperature & mV Channel Logging Procedure

- Connect the shorting cap to the BNC socket on Analog Board 1.
- Connect the [HI762000C](#) 0°C temperature key to the RCA socket (temperature sensor input) on Analog Board 1.
- On the main screen select Mode, if necessary select the analog board and press mV.
- Press mV Setup and use the arrow keys to highlight Logging Interval. Set the logging interval to 15 seconds and press Accept. Press Escape to return to the main screen.
- Press the results key and use the arrow keys to highlight Setup pH/mV/ISE Report, press Select.
- Select Potential and Temperature and Units. All other fields should be unselected.
- Press Save Report to return to the Data Parameters screen.
- Press Escape to return to the main screen.
- Once on the main screen press Start Log to start the automatic log.
- Let the log run for about 10 minutes. Press Stop Log to stop the automatic log.
- Press results, use the arrow keys to highlight Review Last Analysis Report, and press Select.
- The mV column should display 0.0 ±0.1 mV and the temperature column should display 0.0°C ±0.4°C.
- This procedure can be repeated using the [HI762070C](#) 70°C temperature key and on analog board 2.

HI1012EN – TROUBLESHOOTING 2

Description

Method for verifying the dosing of the titrator. This method should be used to troubleshoot a titrator equipped with a 25 mL burette. The titrator dispenses a 10.00 mL pre-titration volume, waits 20 seconds and dispenses an additional 0.5 mL dose twenty times, waiting 20 seconds between each dose, bringing the total volume to 20 mL. This procedure can also be used to check the stirrer functionality. The dosing accuracy of the 25 mL burette is ± 0.025 mL ($\pm 0.1\%$ of the full volume). If the results are not correct, check all fittings for leakage, and burette and tubing for air bubbles. Repeat the measurement.

Reference

ISO/TC 48/SC1N 380E and 383E: "Piston and/or Plunger Operated Volumetric Apparatus"

Accessories

- HI762000C 0 °C Temperature Key
- HI70436 Deionized Water (1 gal)
- HI7662-T Temperature Probe
- Shorting Cap
- Narrow Neck Beaker
- Analytical Balance with 0.0001g resolution

Device Preparation

- Connect the shorting cap to the BNC socket on Analog Board 1
- Install a 25 mL burette filled with room temperature deionized water (HI70436) on pump one and verify that no air bubbles are present in the burette or tubing. If necessary prime the burette until all the air has been removed completely.
- Press Select Method from the main screen. Use the arrow keys to highlight **HI1012EN Troubleshooting 2** and press Select.

Small Dose Dispensing Procedure

- Add a small amount of deionized water to a narrow neck beaker. By doing this the air space in the beaker will be vapor-saturated minimizing evaporation.
- Place the narrow neck beaker on an analytical balance and zero the balance.
- Place the dosing tip through the neck of the beaker. Take care not to immerse it in the liquid during dispensing and not to touch the beaker walls.
- Press start stop.
- Write down the exact weight displaced on the balance after each dose.
- This procedure can be repeated on pump 2.

Other burette sizes can be checked using the following settings, see instruction manual for accuracy:

Burette Volume	Pre-titration Volume	Max. Titrant Volume
5 mL	4.000 mL	8.000 mL
10 mL	8.000 mL	16.000 mL

METHOD PARAMETERS

Name:	Troubleshooting 2	Pre-Titration Stir Time:	0 sec
Method Revision:	4.0	Measurement Mode:	Timed Increment
Analysis Type:	Standard Titration	Time interval:	10 sec
Measurement Input:	Analog CH1	Electrode Type:	Shorting Cap
Stirrer Configuration:		Blank Option:	No Blank
Stirrer:	Stirrer 1	Calculations:	No Formula (mL only)
Stirring Speed:	0 RPM	Titrant Name:	DI Water
Titrant Pump:	Pump 1	Maximum Titrant Volume:	20.000 mL
Reagent Addition 1:	Disabled	Potential Range:	-2000.0 to 2000.0 mV
Reagent Addition 2:	Disabled	Volume/Flow Rate:	25 mL/50.0 mL/min
Dosing Type:	Linear - 0.500 mL	Signal Averaging:	1 Reading
End Point Mode:	Fixed 10.0 mV	Significant Figures:	XXXXXX
Pre-Titration Volume:	10.000 mL		

Calculations

$$V = m * \frac{1}{\rho} * \left(1 + \frac{\rho_{\text{air}}}{\rho_{\text{L}}} - \frac{\rho_{\text{air}}}{\rho_{\text{std}}} \right)$$

- V Volume of measure mass of water (mL)
 m Measure mass of water (g)
 ρ_{L} Density of dispensed water (g/mL)
 ρ_{air} Density of ambient air (g/mL)
 ρ_{std} Density of calibration standard weight (g/mL)

Alternative Calculations

If the actual values of the above parameters are not accessible the following equation can be used:

$$V = M * F$$

- V Volume of measured mass of water (mL)
 F Transformation factor

The values from the table have been calculated by correcting the air and water density with temperature, assuming the density of dry air $\rho_{\text{air}} = 0.0012$ g/mL and density of calibration steel standard weigh $\rho_{\text{STD}} = 8$ g/mL.

Temperature (°C)	Factor
17.0	1.002290
18.0	1.002467
19.0	1.002654
20.0	1.002853
21.0	1.003061
22.0	1.003282
23.0	1.003512
24.0	1.003752
25.0	1.004002
26.0	1.004261
27.0	1.004531
28.0	1.004809
29.0	1.005097
30.0	1.005395

Stirring Speed Fast Check Procedure

- On the main screen select Mode, if necessary select the analog board and press mV.
- Press mV Setup and use the arrow keys to highlight Stirrer Configuration.
Use the arrow keys to highlight Stirrer 1.
Press Accept
- Use the arrow keys to highlight String Speed. Use the numeric keypad to enter 200 rpms then press Accept.
- Press Escape to exit the mV Setup screen.
- From the main screen, press stir, use the up arrow key to increase the stir speed slowly to 2500 rpms.
- Check that the propeller continues to increase speed, following the commands.
- This procedure can be repeated on stirrer 2.

HI1014EN – CONCENTRATION OF PHOSPHORIC ACID

0.00 to 0.01 M (mol/L)

Description

Method for the determination of phosphoric acid (H_3PO_4), by titration of a sample to the point of inflection with sodium hydroxide. The first inflection point corresponds to the H_3PO_4 content and the difference between the first and second corresponds to H_2PO_4^- . The results are expressed as **M (mol/L) phosphoric acid**.

If only phosphoric acid and no other acids or bases are present in the sample, then $\text{H}_3\text{PO}_4 = \text{H}_2\text{PO}_4^-$. If H_3PO_4 is greater than H_2PO_4^- this means other weak acids or bases are present (e.g. citric acid / citrate or ascorbic acid / ascorbate).

Electrode

- [HI1131B](#) Combination pH Electrode
- [HI7662-T](#) Temperature Probe

Reagents

- [HI70456](#) 0.1N Sodium Hydroxide (1 L)
- [HI70436](#) Deionized Water (1 gal)

Accessories

- [HI70300L](#) Storage Solution (500 mL)
- [HI7082](#) Electrode Fill Solution (4 x 30 mL)
- [HI7004L](#) pH 4.01 Buffer Solution (500 mL)
- [HI7007L](#) pH 7.01 Buffer Solution (500 mL)
- [HI7010L](#) pH 10.01 Buffer Solution (500 mL)
- 100 mL Class A Volumetric Pipette
- 150 mL Glass Beaker

Device Preparation

- Connect the pH electrode and temperature probe to the titrator.
- Install a 25 mL burette filled with 0.1N sodium hydroxide ([HI70456](#)) on pump one and verify that no air bubbles are present in the burette or tubing. If necessary prime the burette until all the air has been removed completely.
- For the determination of the exact concentration of the 0.1 N sodium hydroxide, follow [HI0001EN 0.1N Sodium Hydroxide Titrant Concentration](#).
- Press from the main screen. Use the arrow keys to highlight [#1014EN Concentration of H3PO4](#) and press .

Electrode Preparation

- Press from the main screen, if necessary select the analog board and press .
- Calibrate the electrode using pH 4.01, 7.01 and 10.01 buffers. Refer to the instruction manual for calibration procedure.


Sample Preparation

- Use a Class A volumetric pipette to transfer exactly 100.00 mL of sample to a clean 150 mL glass beaker.

Analysis

- Place the beaker under the stirrer assembly and lower it to immerse the pH electrode, temperature probe and stirrer. Ensure that the reference junction of the pH electrode is 5 to 6 mm below the surface.

Note: The dispensing tip should be slightly submerged in the sample.

- Press , the titrator will start the analysis.
- At the end of the titration, after detection of the equivalence point, "Titration Completed" will appear with the result. The result is expressed in **M (mol/L) of phosphoric acid**.
- Remove the pH electrode and stirrer from the sample and rinse them thoroughly with deionized water.
- Record the result.

METHOD PARAMETERS

Name: Concentration of H3PO4
 Method Revision: 4.0
 Analysis Type: Standard Titration
 Measurement Input: Analog CH1
 Stirrer Configuration:
 Stirrer: Stirrer 1
 Stirring Speed: 1400 RPM
 Titrant Pump: Pump 1
 Reagent Addition 1: Disabled
 Reagent Addition 2: Disabled
 Dosing Type: Dynamic
 Min Vol: 0.030 mL
 Max Vol: 0.500 mL
 delta E: 8.000 mV
 End Point Mode: pH 2EQ points, 1st Der
 Recognition Options:
 Threshold: 50 mV/mL
 Range: NO
 Filtered Derivatives: NO
 Pre-Titration Volume: 0.000 mL
 Pre-Titration Stir Time: 10 sec
 Measurement Mode: Signal Stability
 delta E: 0.8 mV
 delta t: 2 sec
 Min wait: 2 sec
 Max wait: 20 sec
 Electrode Type: pH
 Blank Option: No Blank
 Calculations: Sample Calc. by Volume
 Dilution Option: Disabled
 Titrant Name: 0.1N NaOH
 Titrant Conc.: 0.1000 N(eq/L)
 Analyte Size: 100.000 mL
 Analyte Entry: Fixed
 Maximum Titrant Volume: 20.000 mL
 Potential Range: -2000.0 to 2000.0 mV
 Volume/Flow Rate: 25 mL/50.0 mL/min
 Signal Averaging: 1 Reading
 Significant Figures: XXXXX

CALCULATIONS

Calculations: Sample Calc. by Volume
 Titrant units: N (eq/L)
 Titrant volume dosed: V (L)
 Final result units: M (mol/L)
 Titrant Conc.: 0.1000 N(eq/L)
 Sample/Titrant: 1.000 mol/eq
 Sample Volume: 100.000 mL

$$\frac{\text{mol}}{\text{L}} = \frac{V(\text{L}) * 100 * 0.1 * 1.0}{100.0}$$

RESULTS

Titration Report
 Method Name: Concentration of H3PO4
 Time & Date: 11:56 May 2, 2026
 Report ID: Ti_00034

Titration Results
 Method Name: Concentration of H3PO4
 Time & Date: 11:56 May 2, 2026
 Analyte Size: 100.000 mL
 Equivalence point 1:
 pH: 4.677
 Volume: 4.397 mL
 Result: 4.3972E-03 M (mol/L)
 Equivalence point 2:
 pH: 8.916
 Volume: 4.429 mL
 Result: 4.4293E-03 M (mol/L)
 Titration Duration: 1:24 [mm:ss]
 Titration went to Completion:

Analyst Signature: _____

PART 4. TITRATION THEORY

4.1. TITRATION THEORY

4.1.1. INTRODUCTION

A titration is a quantitative, volumetric procedure used in analytical chemistry to determine the concentration of an analyte (the species being measured) in solution. The concentration of the analyte is determined by slowly adding a titrant (reagent) to the solution. As the titrant is added, a chemical reaction occurs between the titrant and the analyte.

Titration reactions are relatively fast, simple reactions that can be expressed using a chemical equation. The titration reaction continues as the titrant is added until all of the analyte is consumed and the analyte reacts completely and quantitatively with the titrant.

The point at which all of the analyte has been reacted is called the equivalence point, also known as the theoretical or stoichiometric endpoint. This point is accompanied by an abrupt physical change in the solution, which sharply defines the endpoint of the reaction. The physical change associated with the titration endpoint can be produced by the titrant, or an indicator, and can be detected visually or by physical measurements.

Titration cannot be used to determine the quantity of all analytes.

The chemical reaction between the titrant and analyte must fulfill four requirements:

- Must be fast and occur within approximately one second after the titrant is added
- Must go to completion
- Must have well-known stoichiometry (reaction ratios)
- A convenient endpoint or inflection point

Titration provides many advantages over alternative methods; they are highly precise, quickly performed and require relatively simple apparatus and instrumentation.

4.1.2. USES OF TITRATIONS

- Acid content of plant effluents, food (e.g.: cheese and wine), plating and etching baths, petroleum products, drugs
- Base content of fertilizer (containing ammonia), bleach, minerals
- Hardness in water
- Metal content of alloys, minerals, ores, clays, waters, plating baths, paints, paper, plant materials, biological fluids, petroleum products
- Moisture content in foodstuffs, petrochemicals, plastics, pharmaceutical products
- Redox reagent concentrations such as available chlorine in potable water, peroxide, traces of oxidants and reductants in food, reductants in high temperature or high pressure boiler water, vitamin analysis

4.1.3. ADVANTAGES & DISADVANTAGES

Advantages of titration as an analytical technique:

- More precise results than many instrumental methods, such as measurement by electrode, the accuracy of the measurement is up to 0.1%
- Simple methods, reasonable capital costs and easy training
- Suitability to measure major components of a mixture or product
- Automation can reduce time and labor spent on each analysis

Disadvantages of titration as an analytical technique:

- The time it takes to prepare standards and titrants
- Good technique is required to achieve precise results (training and practice required)
- Not suitable for determining trace or minor components of a mixture or product
- Limited dynamic range, it may require additional sample preparation (dilution) and repeat analyses

4.2. TYPES OF TITRATIONS

4.2.1. TITRATIONS ACCORDING TO THE MEASUREMENT METHOD

4.2.1.1. Amperometric Titrations

An amperometric titration is performed by placing two electrodes (typically a metal ion-selective electrode and a reference electrode) into the sample solution and keeping the potential of the metal electrode at a selected voltage.

The current that flows, due to the oxidation or reduction of a reactant or product, is plotted vs. volume of titrant to provide the titration curve and locate the equivalence point.

Changes in the current are due to changes in the concentration of a particular species (being oxidized or reduced at the electrode). Generally, the reaction between the analyte and titrant forms a new species. Depending on the titration, the reactants are electroactive and the products are not or vice-versa. Amperometric titration curves look like two straight lines intersecting at the equivalence point, this is due to the change in the electroactivity of the solution.

Figure 1A shows an active analyte and non-reactive titrant.

Figures 1B and 1D show a nonreactive analyte and a reactive titrant.

Figure 1C shows a reactive analyte and titrant.

Many metal ions can be amperometrically titrated using a precipitation, complexation or redox reaction. Some metal ions and species that can be determined in this manner include silver, barium, halides, potassium, magnesium, palladium, molybdate, sulfate, tungstate, zinc, bismuth, cadmium, fluoride, indium, thallium, iodine and gold.

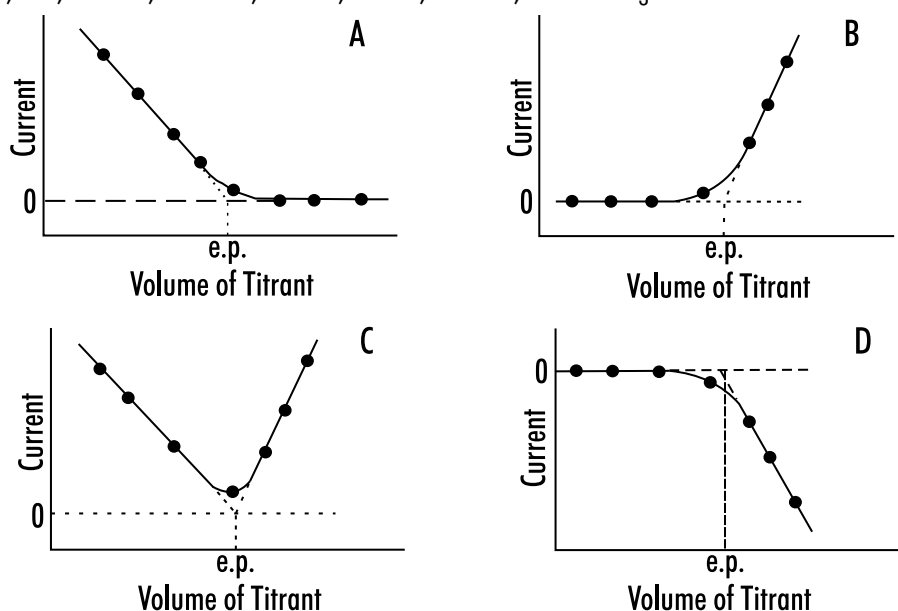


Figure 1: Amperometric titrations

4.2.1.2. Potentiometric Titrations

Potentiometric titrations are done by measuring the voltage across the solution using an electrode system.

An electrode system consists of an indicator electrode and a reference electrode. As titrant is added, the variations in the potential of the indicator electrode, with respect to the reference electrode, are monitored to show the progress of the titration.

Potentiometry is the measurement of a potential under conditions of zero current flow. The measured potential can then be used to determine the analytical quantity of interest, generally a component concentration of the analyte solution. The potential that develops in the electrochemical cell is the result of the free energy change that would occur if the chemical phenomena were to proceed until the equilibrium condition has been satisfied.

There are many types of titrations where potentiometry can be used, e.g. pH electrodes for acid-base titrations, platinum ORP electrodes in redox titrations, ion-selective electrodes, such as chloride or fluoride for a specific ion titration, and silver electrodes for argentometric (silver-based) titrations.

In **Figure 2A** the pH of the solution is plotted against the volume of titrant.

In **Figure 2B** the electrode potential is plotted against the volume of titrant.

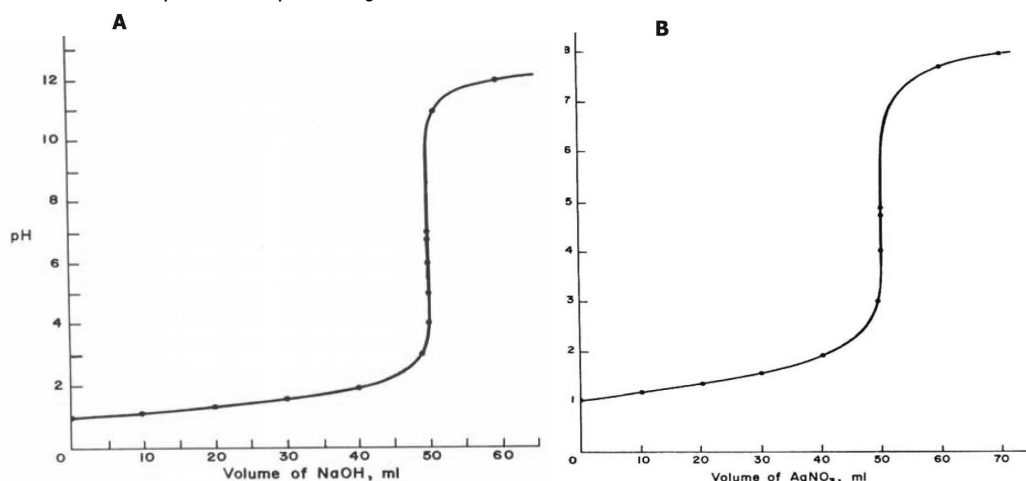


Figure 2: Potentiometric titrations

4.2.1.3. Spectrophotometric Titrations

The name comes from the method used to detect the endpoint of the titration, not its chemistry. Highly colored indicators that change color during the course of the titration are available for many titrations. More accurate data on the titration curve can be obtained if the light absorption is monitored instrumentally using a light source, a simple monochromator and a photodetector, rather than visually determining the color or light absorption change. Light absorption by either an indicator or by one of the reactants or products can be used to monitor the titration.

Figure 3A shows the absorption of a metal-indicator complex is being monitored.

The absorption is constant while the metal is complexed by the ethylenediaminetetraacetic acid (EDTA) titrant. The metal indicator complex was stripped, causing a sharp break in the titration curve. The point where all the metal is complexed and stripped from the indicator is the equivalence point. This point is marked by "e.p." on the graph.

Figure **Figure 3B** shows the metal complex is being measured while being titrated with EDTA.

The new complex being formed is not colored and does not absorb light.

The extrapolated intersection of the two lines determines the equivalence point.

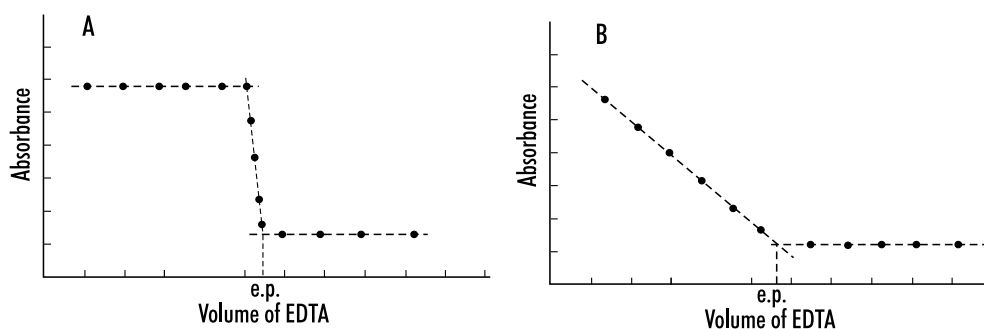


Figure 3: Spectrophotometric titrations

4.2.2. TITRATIONS ACCORDING TO THE REACTION TYPE

4.2.2.1. Acid-Base Titrations

Acid-base titrations are the most common type of titrations. They are based upon a reaction between an acid and a base, a stoichiometric neutralization or the exchange of protons. Virtually all acid-base titrations are carried out using a strong acid or a strong base as the titrant. The endpoint of a titration carried out with a weak acid or a weak base would be difficult to detect due to a small change in pH at the equivalence point.

Chemical indicators can be used to determine the endpoint. The indicator will change color to signify that the end of the titration has been reached. The color of the indicator is dependent upon the concentration of ions in the solution. An acid-base indicator is composed of a conjugate weak acid-weak base pair, where the two forms exhibit different colors depending on the pH of the solution. For an indicator, the acid ionization constant (K_a) is usually written as:

$$K_a = \frac{[H_3O^+][In^-]}{[HIn]}$$

HIn is the acid form of the indicator and In^- is the base form. At the center of the change region, the ratio of $[In^-]$ to $[HIn]$ is one, $[H_3O^+] = K_a$ and $pH = pK_a$. The color change region is usually ± 1 pH unit around this point.

Table 1 contains a list of aqueous acid-base chemical indicators, the pH range, the pK_a and the expected color (acid and base form). It is generally recommended to select a chemical indicator that has a pK_a as close to the endpoint of the titration as possible. When chemical indicators are not suitable, a potentiometric pH titration can also be used. The pH of the solution is plotted versus the volume of titrant added.

Table 1: Aqueous acid-base chemical indicators

pH Range	Indicator	pK_a	Acid Form	Base Form
0.0 - 1.6	Methyl Violet		Yellow	Blue
1.2 - 2.8	Thymol Blue	1.65	Red	Yellow
3.2 - 4.4	Methyl Orange	3.46	Red	Yellow
3.8 - 5.4	Bromocresol Green	4.90	Yellow	Blue
4.8 - 6.0	Methyl Red	5.00	Red	Yellow
5.2 - 6.8	Chlorophenol Blue	6.25	Yellow	Red
6.0 - 7.6	Bromothymol Blue	7.30	Yellow	Blue
6.6 - 8.0	Phenol Red	8.00	Yellow	Red
7.4 - 9.0	Metacresol Purple	8.30	Yellow	Purple
8.0 - 9.6	Thymol Blue	9.20	Yellow	Blue
8.2 - 10.0	Phenolphthalein	9.50	Clear	Pink
9.4 - 10.6	Thymolphthalein		Clear	Blue
10.1 - 12.0	Alizarin Yellow R		Yellow	Red
11.4 - 12.6	Indigo Carmine		Blue	Yellow

Figure 4 shows a traditional strong acid-strong base titration curve, the volume of sodium hydroxide (NaOH) added to the solution is plotted against the pH of the solution. Note the abrupt change in the pH at the equivalence point.

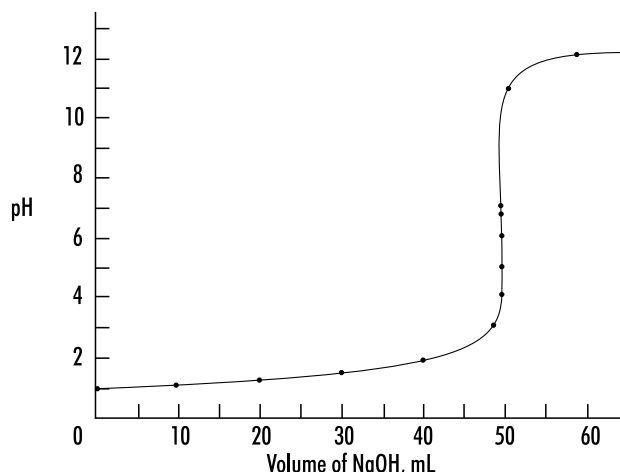


Figure 4: Acid-base titration

4.2.2.2. Argentometric Titrations

Argentometric titrations use silver (nitrate) as the titrant and are generally precipitation titrations, as many silver salts are insoluble. These titrations are commonly used to titrate and determine the concentration of bromide, chloride, cyanide, iodide and sulfide. Argentometric titrations can be done with Mohr's indicator. After all of the chloride has reacted, a red silver chromate precipitate is formed or the titration can be easily followed with a silver ISE (or chloride ISE for chloride titrations) and a reference electrode.

Figure 5 shows the titration of a sodium chloride solution with silver nitrate (AgNO_3). The volume of AgNO_3 is plotted against the potentiometric signal from a chloride ISE.

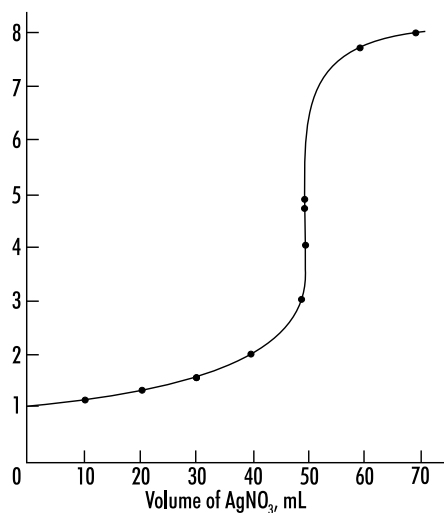


Figure 5: Argentometric titration

4.2.2.3. Complexometric Titrations

A complex is a species where a central metal ion is covalently bonded to one or more electron donating groups called ligands. In a complexometric titration, metal ions are titrated using a titrant that binds strongly to it. Often these titrants contain EDTA or CDTA, polydentate ligands that form very stable coordination compounds with metal ions. The complexation reaction must be fast in order to be useful for direct titration. Some metal ions react too slowly with EDTA for a direct titration.

An indicator electrode that responds to the metal ion can be used to monitor the titration progress. The titration curve will appear similar to a potentiometric titration. Complexation indicators change color at the endpoint as all metal ions are "consumed" or complexed by the titrant.

Figure 6 shows a typically complexometric titration curve when using an indicator electrode that responds to the metal ion.

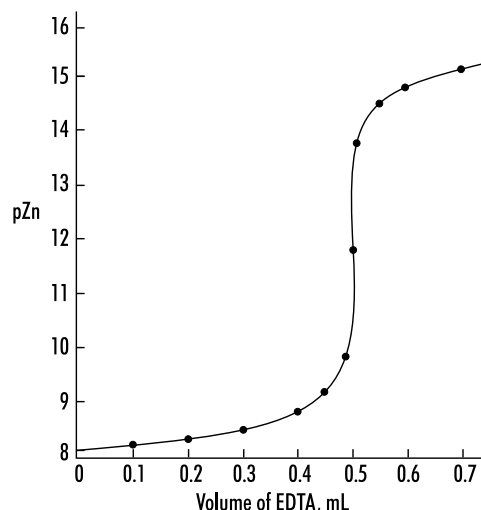


Figure 6: Complexometric titration

4.2.2.4. Ion-selective Titrations

The most popular ion-selective titration is an acid-base titration. The hydrogen ion concentration is specifically measured and monitored during the titration process to locate the equivalence point. Using an ion-selective electrode (ISE) as the indicator electrode, the potentiometric signal (in mV) is used to directly follow a specific ion's concentration (or activity).

Examples of ISE titrations include titrating fluoride with an aluminum titrant using a fluoride ISE, chloride with silver nitrate using a chloride ISE, sodium with a sodium ISE, etc. The equivalence point can be determined by plotting the mV value vs. the amount of titrant added.

4.2.2.5. Non-aqueous Solvent Acid-Base Titrations

Non-aqueous solvents must be used to titrate very weak acids and bases due to the inherent leveling effect water has on all acids and bases dissolved in it. A wide variety of weak acids and bases can be titrated using non-aqueous solvents. Mixtures of acids or bases can often be individually analyzed in a single sequential titration.

Titration of Acids

Weak acids with pK_a 's up to about 11 can be titrated in non-aqueous solvents. These include carboxylic acids, enols, phenols, imides, sulfonic acids and inorganic acids. Water or lower alcohols are suitable for titrating medium to strong acids (pK_a less than 5). Titrating a weaker acid with a strong base titrant requires a solvent less acidic than water or ethanol/methanol. Solvents such as acetone, acetonitrile, *t*-butyl alcohol, dimethylformamide, isopropanol and pyridine have been found to work well for acid-base titrations of strong, medium and weak acids/bases. Titrants include alcoholic potassium hydroxide and various sodium or potassium alkoxides in a 10:1 mixture of benzene/methanol. The best titrants are quaternary ammonium hydroxides (such as tetrabutylammonium hydroxide) due to good solubility of tetraalkylammonium salts of the titrated acids and the clean potentiometric titration curve obtained.

Figure 7, shows an example of titration with tributylmethylammonium hydroxide titrant.

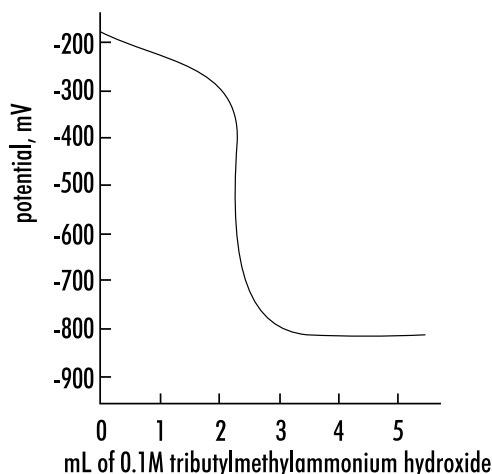


Figure 7: Non-aqueous titration

Titration of Bases

Weak bases with pK_b 's up to about 11, which do not ionize with water, can be titrated in non-aqueous solvents. These bases include aliphatic and aromatic amines, basic nitrogen heterocycles, alkali metal and amine salts of acids, and many other organic basic compounds. Titrating a weak base with a strong acid titrant requires a basic solvent that is as weak as possible. Water and alcohols allow the titration of medium strength bases, such as aliphatic amines ($pK_b = 4$ to 5), but not the titration of weaker bases such as pyridine ($pK_b = 8.8$). Glacial acetic acid works well for weak bases and has been used extensively. Less basic solvents such as acetone, acetonitrile and nitromethane extend the range of titrable compounds.

The endpoint for non-aqueous titrations are usually determined potentiometrically using a pH glass electrode, a modified calomel or double junction reference electrode with a low-flow rate reference junction. Good potentiometric titration curves are obtained in most solvents, except those with very low dielectric constants such as benzene or chloroform, when high electrical resistance of the solvent causes unstable potentials.

4.2.2.6. Precipitation Titrations

Precipitation titrations allow for faster analysis when compared to gravimetric analysis, where a precipitate is formed, filtered, dried and weighed to analyze a compound. Typically silver halides, silver thiocyanate and a few mercury, lead, and zinc salts are titrated using this method. The chemical reactions must form an insoluble salt and precipitate out quickly in order to be analyzed by this method. When the reaction is not quick, a back titration can be used. A measured excess of the precipitating reagent (titrant) is added to force the reaction to occur, and then unreacted titrant is titrated with a standard solution of another reagent.

4.2.2.7. Redox Titrations

There are a number of oxidation-reduction reactions that can be used to determine unknown concentration by titration. If the reaction goes to completion, is fast and has an analytical signal available to follow it, a titration can be performed. The term “fast” means that each addition of titrant is reacted completely and the sensing electrode is able to detect the change in solution in less than one second.

Figure 8 shows an example of a redox titration using Cerium (IV) as a titrant.

Redox titrations are potentiometric titrations where the mV signal from a combination ORP (redox) electrode (usually with a platinum indicator electrode) is used to follow the reaction of oxidant/reductant. The electrode potential is determined by the Nernst equation and is controlled by the oxidant/reductant ratio.

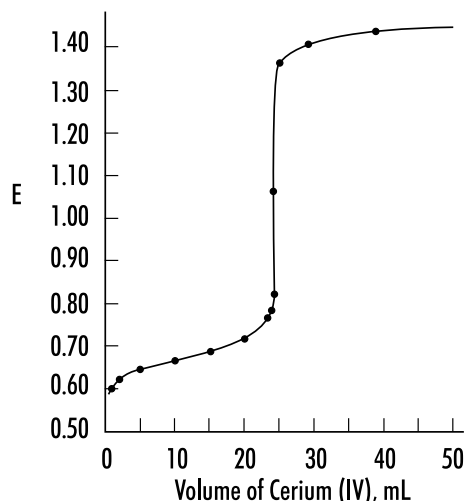


Figure 8: Redox titration

Visual indicators, such as Ferroin, are also available. The oxidized and reduced form of the indicator will have different colors and can be used to determine the endpoint.

Various reductants can be determined by titrants with oxidants such as potassium permanganate, potassium chromate or iodine. Commonly used reductants that are used as titrants include sodium thiosulfate and ferrous ammonium sulfate.

As with acid-base titrations, the potential changes dramatically at the equivalence point.

4.2.2.8. Karl Fischer Titrations

This method is based on a well-defined chemical reaction between water and the Karl Fischer reagent. The chemistry provides excellent specificity for water determination. The method can be used to determine free and bound water in a sample matrix. The Karl Fischer method is widely considered to produce the most rapid, accurate and reproducible results and has the largest detectable concentration range spanning 1 ppm to 100%.

The determination of water content is one of the most commonly practiced methods in laboratories around the world. Knowledge of water content is critical to understanding chemical and physical properties of materials and ascertaining product quality. Water content determination is conducted on many sample types including pharmaceuticals and cosmetics, foods and natural products, organic and inorganic compounds, chemicals, solvents and gases, petroleum and plastic products as well as paints and adhesives. The KF method is verifiable and can be fully documented. As a result, Karl Fischer titration is the standard method for analysis of water in a multitude of samples as specified by numerous organizations including the Association of Official Analytical Chemists, the United States and European Pharmacopoeia, ASTM, American Petroleum Institute, British Standards, and DIN.

4.2.3. TITRATIONS ACCORDING TO THE TITRATION SEQUENCE

4.2.3.1. Back Titrations

Back titrations are generally used when a reaction is too slow to be directly accomplished during a “direct” titration, where the reaction goes to completion within a few seconds. In a back titration, a large excess of a reagent is added to the sample solution, helping a slow reaction to go to completion. The unreacted, excess reagent is then titrated. The difference in the total volume of the first reagent added and amount determined from the second titration is the quantity of reagent required to complete the first reaction.

4.2.3.2. Multiple Endpoint Titrations

Under certain conditions, some titrations can exhibit more than one equivalence point and be titratable to the individual endpoints to determine the concentration of each individual component. Examples of these types of titrations include acid-base (different strengths acids or bases are in a mixture), redox (each species has a different reduction potential), complexometric (different species are separately titratable), and acid-base, using polyprotic acids (the pK_a of the different protons varies enough to separate them).

In **Figure 9A** a titration of a polyprotic acid is shown, the different acid strengths of the first and second proton can be determined.

Figure 9B shows a titration with two different metal redox species, the different redox potentials allow the species to be separated.

In **Figure 9C** the solution being titrated contains a mixture of strong, weak, and very weak acids, the different pK_a 's allow the species to be separated.

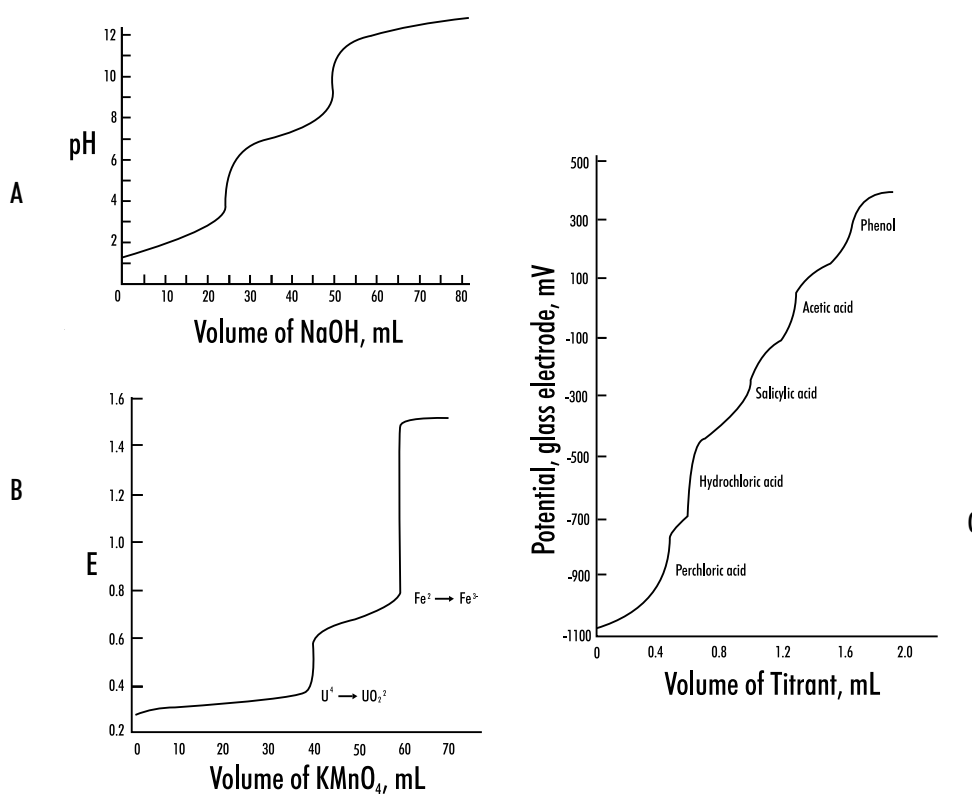


Figure 9: Multiple endpoint titrations

4.3. TITRATION PROCEDURE

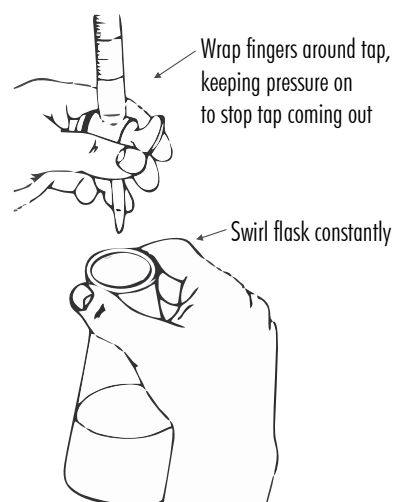
4.3.1. MANUAL TITRATION

Apparatus required for manual titration include:

- Volumetric burette, for precisely controlled delivery of titrant to the reaction vessel
- Erlenmeyer, or similar flask, that facilitates constant mixing or swirling required to ensure solution homogeneity
- Volumetric pipettes for the precise addition of samples and indicator solutions
- Titrant solutions of known concentration
- A visual or instrumental indicator for detecting the completion of the reaction

A typical manual titration consists of the following steps:

1. A volumetric pipette is used to add a known volume of sample to the flask.
2. An indicator solution or instrument probe is added to the flask.
3. A burette is used to measure the addition of titrant to the flask and dispense titrant in a controlled manner.
4. Titrant is added via the burette until the method indication signals the reaction endpoint.
5. Analyte concentration is calculated based on the concentration and volume of titrant required to reach the endpoint.



4.3.2. AUTOMATIC TITRATION

Automatic titrators are high-precision analytical instruments that deliver the titrant, monitor the physical change associated with the titration reaction, automatically stop at the endpoint and calculates the concentration of the analyte. Automatic titrators are best for repetitive titrations and high-accuracy analyses.

An automatic titrator must have an accurate liquid dispensing system. In high-accuracy systems, the liquid dispensing system consists of three main components: motor-driven syringe burette capable of accurately and precisely dispensing very small volumes of titrant, valve system capable of switching between titrant intake and outlet and a dispensing tip. These three main subsystem components must be as accurate as possible, with very low gear backlash in the burette pump, minimal piston seal flexing, precision ground inner diameter of the glass syringe, a low dead-volume valve, minimal evaporation/permeation, and chemically resistant tubing.

Apparatus required for automatic titration include:

- An automatic titrator, equipped with a burette
- A beaker
- An electronic stirring system, either a propeller stirrer or a magnetic stir bar and stir plate
- Volumetric pipettes for the precise addition of samples
- Standard titrant solutions of known concentration
- An electrode system that can be used to determine the endpoint of the titration

A typical automatic titration consists of the following steps:

1. Set up the automatic titrator according to the manufacturer's instructions.
2. Use a volumetric pipette to add a known volume of sample to the beaker.
3. Submerge the propeller stirrer or add the stir bar to the beaker and turn on.
4. Start the titration.
5. The titrator will automatically stop at the endpoint and determine the concentration of the analyte.

4.4. TITRATION RESULTS

4.4.1. ACCURACY

The factors most critical to achieving accurate results are the concentration of the sample, size of the sample, and having an optimized set of method parameters.

4.4.2. REPEATABILITY

Repeatability or the agreement between replicate determinations, is expressed quantitatively as the relative standard deviation (RSD).

4.4.3. SOURCES OF ERROR

One of the advantages of volumetric analysis is excellent accuracy and precision. The sources of error can be grouped into sampling, titrant and standards, chemical reactions, endpoint determination and calculations.

4.4.3.1. Sampling Errors

- Selection of a non-homogeneous or non-representative sample
- Sample changed or was contaminated during collection, storage or transfers
- Poor technique when transferring sample to beaker or flask
- Errors in the balance (calibrate and check balance regularly)

4.4.3.2. Preparation Errors

Incorrect preparation due to:

- Poor technique in weighing the salt or when transferring to volumetric glassware
- Low-purity salts or water used to make titrant and standard
- Dirty or wet glassware
- Improper storage of titrant or standard which allows water gain, evaporation or deterioration
- Failure to standardize frequently, to adjust for change in titrant
- Failure to flush titrator tubing with a volume of titrant before standardizing
- Volume errors from pipettes and volumetric flasks (grade A glassware is required)
- Balance errors when weighing out salts (calibrate and check balance regularly)

4.4.3.3. Dispensing Errors

Incorrect dispensing due to:

- Dead valve volume and leaking valve
- Inaccuracy in motor drive and gear lash or backlash
- Poor burette or piston seal
- Non-uniform diameter of burette glass cylinder
- Chemical incompatibility with tubing or bubble generation
- Density or temperature changes in titrant
- Inadequate volume to cover electrode

4.4.3.4. Chemical Reaction Errors

- Inappropriate solvent or sample, resulting in side reactions
- Poor mixing in the titration vessel
- Reaction between titrant and sample is not rapid
- Reaction does not go to completion
- Reaction has side reactions

4.4.3.5. Endpoint Determination Errors

Most manual titrations use a visual indicator to indicate when the endpoint is reached and the titration should be stopped. Automatic titrators use instrumental methods to determine the end of a titration and the equivalence point. There are two predominant methods used to determine the equivalence point, first derivative and second derivative.

The first derivative is often used to determine the inflection point. The inflection point of the titration curve (mV vs. volume) is normally assumed to be the equivalence point. The maximum value of the first derivative (ΔmV vs. ΔV) corresponds to the theoretical equivalence point. During a titration it is rare to have a data point exactly at the first derivative maximum, the maximum value is determined by interpolating the first derivative data points.

The second derivative (ΔmV^2 vs. ΔV^2) can also be used to determine the equivalence point, and can offer advantages over the first derivative method. Second derivatives have increased sensitivity to smaller inflection points and easier numerical evaluation of the actual equivalence point. The value where the second derivative is equal to zero is the equivalence point. The second derivative requires fewer points located near the equivalence point, where data is often not obtained or not as reliable.

Errors in determining the endpoint can result from:

- Incorrect signals from the sensor
- Sensor drift
- Sensor or instrument has slow response (it is recommended to keep the sensors in good condition)
- Inappropriate setting on the titrator

4.5. CALCULATIONS

The main variables used in calculating a result from a titration are the sample volume, the concentration of the titrant, and the volume of titrant required to reach the equivalence point. At the equivalence point, an equal number of equivalents of the analyte and titrant has been added.

4.5.1. SAMPLE CALCULATION BY MASS

$$C_{\text{sample}} = \frac{V_{\text{titrant}} \times C_{\text{titrant}} \times \text{Ratio} \times \text{FW}_{\text{analyte}}}{m_{\text{sample}}} \times 100$$

C_{sample}	Sample concentration (g/100g)
V_{titrant}	Volume of titrant
C_{titrant}	Titrant concentration (eq/L)
Ratio	Equivalence ratio of analyte / titrant (mol analyte / eq titrant)
$\text{FW}_{\text{analyte}}$	Formula weight of the analyte (g/mol)
m_{sample}	Mass of sample (g)

4.5.2. SAMPLE CALCULATION BY VOLUME

$$C_{\text{sample}} = \frac{V_{\text{titrant}} \times C_{\text{titrant}} \times \text{Ratio} \times \text{FW}_{\text{analyte}}}{V_{\text{sample}}} \times 100$$

C_{sample}	Sample concentration (g/100mL)
V_{titrant}	Volume of titrant
C_{titrant}	Titrant concentration (eq/L)
Ratio	Equivalence ratio of analyte / titrant (mol analyte / eq titrant)
$\text{FW}_{\text{analyte}}$	Formula weight of the analyte (g/mol)
V_{sample}	Volume of sample (mL)

4.5.3. STANDARDIZE TITRANT BY MASS

Titrant standardization is the second most important calculation in titrations. A primary standard is titrated in order to determine the concentration of the titrant. This is essentially a typical titration calculated in “reverse”, where the concentration of the solution is known and the titrant is unknown.

$$C_{\text{titrant}} = \frac{m_{\text{standard}} \times \text{Ratio}}{FW_{\text{standard}} \times V_{\text{titrant}}}$$

C_{titrant}	Titrant concentration (N)
m_{standard}	Mass of standard (g)
Ratio	Equivalence ratio of titrant / standard (eq titrant / mol standard)
FW_{standard}	Formula weight of the standard (g/mol)
V_{titrant}	Volume of titrant (L)

4.5.4. STANDARDIZE TITRANT BY VOLUME

Titrant standardization is the second most important calculation in titrations. A primary standard is titrated in order to determine the concentration of the titrant. This is essentially a typical titration calculated in “reverse”, where the concentration of the solution is known and the titrant is unknown.

$$C_{\text{titrant}} = \frac{V_{\text{standard}} \times (1 \text{ L}/1000 \text{ mL}) \times C_{\text{standard}}}{V_{\text{titrant}}}$$

C_{titrant}	Titrant concentration (N)
V_{standard}	Volume of standard (mL)
C_{standard}	Concentration of standard (eq/L)
V_{titrant}	Volume of titrant (L)

4.5.5. BLANK TITRATION

In a blank titration a pre-titration is performed, often times on the solvent to be used for the sample titration, and the titrant volume required to reach the endpoint is noted. This blank value nullifies error due to titrant required to react with the components of the titration solution matrix. The basic titration equation can be used for a blank titration, with the single modification that the volume of titrant used in the blank titration should be subtracted from the regular titration titrant volume.

$$C_{\text{sample}} = \frac{C_{\text{titrant}} \times (V_{\text{sample}} - V_{\text{blank}}) \times \text{Ratio} \times FW_{\text{analyte}}}{m_{\text{sample}}} \times 100$$

C_{sample}	Sample concentration (g/100 g)
C_{titrant}	Titrant concentration (eq/L)
V_{sample}	Volume of titrant required for the sample (L)
V_{blank}	Volume of titrant required for the blank (L)
Ratio	Equivalence ratio of analyte / titrant (mol analyte / eq titrant)
FW_{analyte}	Formula weight of the analyte (g/mol)
m_{sample}	Mass of sample (g)

4.5.6. MULTIPLE ENDPOINT TITRATION

Some titrations have two or more endpoints, each corresponding to the equivalence point for a specific reaction. Multiple endpoint titrations are similar to a blank titration in that the volume of titrant required to reach the first endpoint is subtracted from the titrant volume used to reach the next sequential endpoint.

$$C_{\text{sample1}} = \frac{V_{\text{titrant1}} \times C_{\text{titrant}} \times \text{Ratio} \times \text{FW}_{\text{analyte1}}}{m_{\text{sample}}} \times 100$$

$$C_{\text{sample2}} = \frac{(V_{\text{titrant2}} - V_{\text{titrant1}}) \times C_{\text{titrant}} \times \text{Ratio} \times \text{FW}_{\text{analyte2}}}{m_{\text{sample}}} \times 100$$

$$C_{\text{sample3}} = \frac{(V_{\text{titrant3}} - V_{\text{titrant2}}) \times C_{\text{titrant}} \times \text{Ratio} \times \text{FW}_{\text{analyte3}}}{m_{\text{sample}}} \times 100$$

C_{sample1}	Sample 1 concentration (g/100g)
C_{sample2}	Sample 2 concentration (g/100g)
C_{sample3}	Sample 3 concentration (g/100g)
V_{titrant1}	Volume of titrant required to reach the first endpoint (L)
V_{titrant2}	Volume of titrant required to reach the second endpoint (L)
V_{titrant3}	Volume of titrant required to reach the third endpoint (L)
C_{titrant}	Concentration of titrant (N)
Ratio	Equivalence ratio of analyte / titrant (mol analyte / eq titrant)
$\text{FW}_{\text{analyte1}}$	Formula weight of the analyte 1 (g/mol)
$\text{FW}_{\text{analyte2}}$	Formula weight of the analyte 2 (g/mol)
$\text{FW}_{\text{analyte3}}$	Formula weight of the analyte 3 (g/mol)
m_{sample}	Mass of sample (g)

4.5.6.1. Back Titration

The equation used in back titration calculations is also similar to the equation for a blank titration. Instead of subtracting the initial amount of titrant needed to react with the blank, the amount of second titrant needed to react with the excess titrant added in the first titration is subtracted from the amount of the first titrant added. The difference between the two amounts is the amount of titrant necessary to reach the first equivalence point.

$$C_{\text{sample}} = \frac{(C_{\text{titrant1}} \times V_{\text{titrant1}} - C_{\text{titrant2}} \times V_{\text{titrant2}}) \times \text{Ratio} \times \text{FW}_{\text{analyte}}}{V_{\text{sample}}} \times 100$$

C_{sample}	Sample concentration (g/100mL)
C_{titrant1}	Concentration of titrant 1 (N)
V_{titrant1}	Volume of titrant 1 (L)
C_{titrant2}	Concentration of titrant 2 (N)
V_{titrant2}	Volume of titrant 2 (L)
Ratio	Equivalence ratio of analyte / titrant (mol analyte / eq titrant)
$\text{FW}_{\text{analyte}}$	Formula weight of the analyte (g/mol)
V_{sample}	Volume of sample (mL)

4.6. GLOSSARY

Acid	A chemical species that can donate one or more protons (hydrogen ions).
Acid-Base Titration	Stoichiometric neutralization titrations, based upon the reaction that occurs between an acid and a base.
Activity	A physical property corresponding to the concentration of all ions in a solution. Electrodes respond to activity.
Amperometric Titration	Titrations where the current flow between two electrodes (often a metal electrode and a reference electrode) are used to monitor the titration progress.
Analyte	The chemical species being measured in a titration.
Argentometric Titration	Titrations that use silver (nitrate) as the titrant. These titrations are typically precipitation titrations.
Automatic Titrator	An instrument designed to automatically carry out a titration. It will add the appropriate amount of titrant, determine the endpoint and calculate the results.
Back Titration	A type of titration where an excess amount of titrant is added to a sample, forcing a sluggish reaction to go to completion. The excess reagent is then "back" titrated with a second titrant.
Base	A chemical species that can accept one or more protons (hydrogen ions).
Biamperometric Indication	Uses a dual platinum pin electrode to measure the current flow through a titration solution.
Bivoltametric Indication	Uses a dual platinum pin electrode to measure the voltage required to maintain a constant current flow through a titration solution while constant voltage is applied across the platinum elements of the electrode.
Burette	A graduated cylindrical piece of laboratory glassware that is used to dispense precise amounts of solution.
Complex Ion	A species where a central metal ion is covalently bonded to one or more electron donating groups called ligands.
Complexometric Titrations	Metal ions are titrated using a titrant that binds strongly to it. The titrants often contain Ethylenediaminetetraacetic Acid (EDTA) or Cyclohexylenedinitrilotetraacetic Acid (CDTA).
Endpoint	The point where a titration is stopped because a physical change in the solution has indicated a completed titration. Titration endpoints typically coincide with the equivalence point. A fixed value endpoint (pH or mV) can be used as well. The titration will stop at the desired point regardless if the titration is complete.
Equivalence Point	The point where the quantity of titrant is stoichiometrically equal to the quantity of analyte.
Formal	The theoretical number of equivalents per liter of the solution. It is used in solutions where the exact concentration of a species may be affected by the other ions present, therefore the stated concentration may not be exactly correct.
Gravimetric Analysis	A quantitative determination of an analyte based on the mass of the solid.

Indicator Electrode	An electrode that responds to the species of interest. The electrode potential is proportional to the concentration or activity of that ion in the solution being measured.
Indicators	Chemical indicators are typically organic dyes that change form under different physical conditions, causing a color change that can be seen by an analyst. Typically used in manual titrations, chemical indicators have been replaced with electrometric indicators, which are used with automatic titrators.
Inflection Point	The point on a titration curve where the second derivative curve changes signs.
Ion-Selective Electrode (ISE)	An electrode that responds to a specific ion. The electrode potential is proportional to the concentration or activity of that ion in the solution being measured.
Karl Fischer Titration	A titration that uses a chemical reaction that is specific for determining water.
Manual Titration	A titration that is carried out by hand. The analyst must add the appropriate amount of titrant, determine the endpoint and calculate the results.
Molar	The concentration of a solute in a solution.
Mole (mol)	A quantity of a chemical species. The molecular weight of a substance in grams is equal to the mass of one mole of the substance. One mole is equal to 6.022×10^{23} atoms or molecules.
Monochromator	A device that allows only a narrow range of wavelengths to pass through it by separating the light into different wavelengths.
Multiple Endpoint Titration	A titration that reacts multiple species in solution, sequentially, using the same titrant. The concentration of each analyte can be determined from their respective endpoints.
Nernst Equation	The fundamental equation relating cell voltage to the concentration of a solution.
Neutralization	A chemical reaction where an acid and a base react to form a neutral salt and water.
Non-aqueous	A solution that does not contain water.
Non-aqueous Titration	A titration that is performed in non-aqueous solutions, typically used to titrate very weak acids and bases to eliminate the leveling effect water has on all acids and bases dissolved in it.
Normal	The concentration of a solution which accounts for any stoichiometric difference between the various species in a solution.
Oxidation / Reduction Potential (ORP)	The measurement describing whether a species wants to donate or accept electrons from other species in a redox reaction. If a solution's reduction potential is higher than the species it is reacting with, it will typically gain electrons or be reduced. If the potential is lower than the species it is reacting with, it will typically lose electrons or be oxidized.
Oxidant	The species that is accepting electrons in a redox reaction.
Pipette	Scientific apparatus that is used to deliver precise volumes of liquids.
Polyprotic Acid	Acids that are capable of donating more than one proton per acid molecule.
Potentiometric Titration	A titration in which the endpoint is determined by monitoring the voltage of the solution using an electrode.

Precipitation Titration	A titration in which the analyte reacts with the titrant to form an insoluble compound. The endpoint is typically detected with an ISE sensitive to either the analyte or titrant.
Reagent	The chemical added in a titration that causes the given reaction to occur.
Reduction-Oxidation Reaction (redox)	A chemical reaction in which the atoms involved in the reaction have their oxidation numbers changed. Reduction is the gain of electrons, which decreases the oxidation number. Oxidation is the loss of electrons, which increases the oxidation number.
Reductants	The electron donor in a redox reaction.
Reference Electrode	An electrode that supplies a constant electrode potential. It is used in combination with an "indicator" electrode, allowing for the "indicator" electrode potential to be measured.
Relative Standard Deviation (RSD)	A measure of the amount of relative variation in a set of data. It is calculated by dividing the standard deviation by the mean: $RSD = (\text{Standard Deviation of } X) * 100 / (\text{Mean of } X)$
Repeatability	The variation in sample measurements taken by a single person or instrument under the same conditions.
Spectrophotometric Titration	A titration in which the endpoint is marked by a change in color and/or color intensity.
Stoichiometry	The quantitative relationship of the reactants and products in a chemical reaction.
Titrant	The chemical added in a titration that causes the given reaction to occur.
Titration	A quantitative, volumetric procedure used in analytical chemistry to determine the concentration of an analyte in solution. The concentration of the analyte is determined by slowly adding a titrant to the solution. As the titrant is added, a chemical reaction between the titrant and the analyte occurs.
Titration Curve	A graph containing the physical data obtained for a titration. The data plotted is often an independent variable (volume of titrant) vs. a dependent variable (pH of the solution). From the titration curve, the equivalence point or endpoint can be determined.

4.7. LIST OF FIGURES

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CERTIFICATION

All Hanna[®] instruments conform to the **CE European Directives**.



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